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AMPLIBIOTHERM DNA POLYMERASE FS-T-002

DESCRIPTION:
Amplibiotherm DNA Polymerase is a thermostable 94 kDa DNA Polymerase purified from E.coli PVG-AI recombinant strain expressing ***Thermus aquatiqus*** polymerase gene.
The enzyme catalyzes polymerisation of nucleotides into duplex DNA in the 5'-3' direction in presence of Mg++ ions. The enzyme possesses also a 5'-3' exonuclease activity. Amplification of target **DNA fragments <100 b.p. up to 10.000 b.p.** can be achieved with this enzyme.

CONCENTRATION:
5 units/ul

Description	FS-T-002
Amplibiotherm Taq DNA Polymerase	250 U
10X Reaction Buffer	1 vial
25mM MgCl2 separately	1 vial

UNIT DEFINITION:
One unit is defined as the amount of enzyme that incorporates 10 nmoles of dNTPs into acid-insoluble form in 30 minutes at 72°C.

STORAGE AND DILUTION BUFFER:
20 mM Tris-HCl , 1 mM DTT; 0.1 mM EDTA;
100 mM NaCl , Stabilizer ; 50% glycerol pH: 7.5(25°C)
buffer is optimized to use with 0.2mM for each dNTPs

STORAGE TEMPERATURE:

Store Amplibiotherm DNA Polymerase below 0°C, preferably at -20° C, in a constant temperature freezer.

EXPIRY DATE:
1 year upon receipt.

10X REACTION BUFFER:
100mM Tris-HCl, 500mM KCl, pH 9.0 (25°C).

REACTION BUFFERS	
10X Reaction Buffer (contains 15mM $MgCl_2$; included)	Cat. No. **FS-B-006**
10X Reaction Buffer (without $MgCl_2$; plus 25 mM $MgCl_2$ separately)	Cat. No. **FS-B-007**

Protocol for routine *Taq* PCR reaction

On ice, prepare each of following master mixes, combine, and place in heated (to 95°C) thermal cycler:

For 50 µl PCR Reaction	Volume	Final Conc.
Amplibiotherm DNA Polymerase (5U/ul)	0.25 ul	1.25 U
10X PCR Buffer	5 ul	1 X
dNTP mix (2.5 mM each)	4 ul	200 uM each
Template	< 500 ng	< 500 ng
Forward Primer	5 ~ 50 pmol	0.1~1 uM
Reverse Primer	5 ~ 50 pmol	0.1~1 uM
Distilled water	up to 50 ul	

Gently mix the reaction and spin down in microcentrifuge.
If the termocycler does not have a heated cover, add one drop of mineral oil to the reaction tube to prevent evaporation.

Cycling conditions for a routine PCR reaction:

Step	Temp (°C)	Time	Cycle
Initial Denaturation	95	5 min.	1
Denature	95	10 ~ 30 sec.	25 ~40
Anneal	50~65	10 ~ 30 sec.	
Extend	72	10 ~ 60 sec.	
Final Extension	72	5 min.	1

Cycling conditions may need to be optimized, depending on different primer and template combinations. For example, raise the annealing temperature to prevent non-specific primer binding, increase extension time to generate longer PCR products.

GOTAQ FLEXI DNA POLYMERASE GREEN G2 FS-T-0531

Description

GoTaq Flexi DNA Polymerase Green G2 is new generation of Taq polymerase that gives robust amplification and high DNA yield in shorter PCR running time (15-30 s/kb extension). GoTaq Flexi DNA polymerase supports the robust and reliable amplification of a wide range of DNA templates up to 6 kb. GoTaq Flexi is provided with 5× Green Reaction Buffer allowing reactions to be loaded directly into gels without the extra adding of loading dye and with a 10x Colorless Buffer. GoTaq Flexi lacks 3' 5'exonuclease activity. Resulting PCR products have an A-overhang and suitable for cloning.

Description	FS-T-0531
GOTAQ Flexi DNA Polymerase Green G2	500 U
5X Green Reaction Buffer	2 x 2 ml
10 X Colorless Buffer	4X1 ml
50 mM MgCl2 Solution	2X1 ml

Storage Buffer

20mM Tris-HCl, 1mM dithiothreitol, 0.1mM EDTA, 100mM NaCl, Stabilizer, 50% glycerol, pH 8.0 (25°C).

5X Green Reaction Buffer

The 5X Green Reaction Buffer contains 2 dyes (blue & yellow) that separate during electrophoresis to monitor migration progress. The blue dye migrates at the same rate as a 3-5kb DNA fragment in a1% agarose gel,the yellow dye migrates a t a rate faster than primers (<50kb) in a 1% agarose gel.

10X Colorless Buffer

The Colorless Buffer is used when direct fluorescence or absorbance readings are required without prior purification of the amplified DNA from the PCR.
Proprietary formulation supplied at pH8.5 containing Tris-HCl, KCl and PCR enhancers and do not contain Mg.

Unit Definition

1 unit is defined as the amount of enzyme that will incorporate 10 nmol of dNTP into acid-insoluble material in 30 minutes at 74°C.

Storage Conditions

Store all components at -20ºC in a non-frost-free freezer.

Quality Control

No endonuclease activity, nicking activity, exonuclease activity, or priming activity has been detected.

Genomic DNA contamination:

The product must be free of any detectable DNA contamination as evaluated through PCR. Thus, it is suitable for the amplification of bacterial and fungal DNA based on 16S and 18S rRNA PCR assays.

Nuclease assays

0.2-0.3 µg of pNZY28 plasmid DNA are incubated with 5 U of GoFlexi Taq DNA polymerase, in 1× Reaction Buffer, for 14-16 hours at 37 °C. Following incubation, the DNA is visualised on a Green Gel Safe -stained agarose gel. There must be no visible nicking or cutting of the nucleic acid. Similar tests are performed with buffer and MgCl2 solution.

Functional assay

GoFlexi Taq DNA polymerase is extensively tested for performance in a polymerase chain reaction (PCR) of different-sized DNA fragments (1 and 2.5 kb) from human genomic DNA in the presence of 5× Green Reaction Buffer and MgCl2 solution. The resulting PCR products are visualized as single bands in a Green Gel Safe stained agarose gel

Recommended Protocol

On ice, prepare each of following master mixes, combine, and place in heated (to 95°C) thermal cycler:

PCR using Green reaction buffer

For 50µl PCR Reaction	Volume	Final Conc.
GOTAQ Flexi DNA Polymerase Green	0.25-1 µl	1.25 U
5X Green Reaction Buffer	10 µl	1 X
50 mM MgCl2 Solution	2.5mM	(1.5~4.0) mM
dNTP mix (2.5 mM each)	0.4 µl	(0.25-0.5)
Template DNA **(see below)**	5 pg-0.5ug	0.1-0.5 uM
Forward Primer	0.25 ul	0.1~0.5 µM
Nuclease free water	up to 50 µl	

PCR Reaction using Colorless buffer

For 50µl PCR Reaction	Volume	Final Conc.
GOTAQ Flexi DNA Polymerase G2	0.25-1 µl	1.25 U
10X Colorless Buffer	5 µl	1 X
50 mM MgCl2 Solution	2.5mM	(1.5~4.0) mM
dNTP mix (2.5 mM each)	0.4 µl	(0.25-0.5)
Template DNA **(see below)**	5 pg-0.5ug	0.1-0.5 uM
Forward Primer	0.25 ul	0.1~0.5 µM
Nuclease free water	up to 50 µl	

General Cycling Conditions:

Step	Temp (°C)	Time	Cycle
Initial Denaturation	95	3 min.	1
Denature	94	30 sec.(**)	25 ~ 35
Anneal	50~65(*)	30 sec.	
Extend	72	15 ~ 30 sec./kb**	
Final Extension	72	5 min.	1

* Annealing temperature should be optimized for each primer set based on the primer Tm; typically, it should be Tm- 5 °C. ** For DNA fragments higher than 3 kb to 6 kb in size, it may be beneficial to use 20 sec for denaturation and 30-60 sec/kb for extension

Primer Design

PCR primers generally range in length from 15–30 bases and are designed to flank the region of interest. Primers should contain 40–60% GC, and care should be taken to avoid sequences that might produce internal secondary structure. The 3'-ends of the primers should not be complementary to avoid the production of primer- dimers. Primer-dimers unnecessarily remove primers from the reaction and result in an unwanted polymerase reaction that competes with the desired reaction. Avoid three G or C nucleotides in a row near the 3'-end of the primer, as this may result in non- specific primer annealing. Ideally, both primers should have nearly identical melting temperatures (Tm), allowing their annealing with the denatured template DNA at roughly the same temp.

DNA template

The optimal amount of starting material may vary depending on its quality and complexity. In general, we recommend using 10 ng to 500 ng of genomic DNA templates, although the enzyme is sensitive enough to amplify fragments from as little as 5 pg of human gDNA,for example. Lower amounts of template may be used for amplification of less complex DNA (typically 1-20 ng). When using a cDNA synthesis reaction as template do not exceed 10% of the final PCR reaction volume.

Applications:

- Routine PCR
- Genotyping
- Library construction
- TA Cloning
- Primary Extension
- Colony PCR – Multiplex PCR

HOT - START DNA POLYMERASE (with GC-Buffer) FS-T-2131-1

Description

HOT START DNA polymerase is an ideal enzyme for high fidelity PCR with excellent processivity. It is a novel engineered enzyme with comparable performance to Pyrococcusiosus DNA polymerase. With unique structure HS DNA polymerase contains a recombinant synthesis enhancement domain to increase fidelity and extension speed. The antibody-mediated hot-start feature significantly inhibits non-specific amplifications at room temperature.
HOT START is one of the thermostable DNA polymerases with strong 3 ' -5 ' exonuclease activity which results in its extreme high fidelity, 10-15 times higher than Taq DNA polymerase and 6 times higher than Pyrococcus furiosus DNA polymerase.
The HS PCR Kit is supplied with a 5X HF Buffer and a 2.5X GC Buffer. The 5X HF Buffer is an optimized buffer for general high fidelity amplifications while the 2.5X GC Buffer is used in the amplifications of problematic or GC-rich templates.

Contents	FS-T-2131-200
Hot Start DNA Polymerase (2000U/mL)	200 RNXS
5X HF PCR Buffer	2x 1 mL
2.5X GC PCR Buffer	4X 1 mL
dNTPs (10 mM each)	200 µL

Thermal Inactivation: No
Product End: Blunt end

Standard Protocol

1. It is recommended to prepare all reaction components on ice, and then quickly transfer the reaction system to a thermocycler preheated to 98℃.It is recommended to prepare all reaction.
2. All components should be mixed and collected at the bottom of a tube with a quick spin before use. Add Hot Start 2X Master Mix with Dye at the end to prevent primer degradation by its strong 3´-5´ exonuclease activity.
Note: The Hot Start DNA polymerase requires special reaction conditions different from other polymerase protocols. Please refer to the recommended reaction conditions below for the better amplification yields.

Recommended Protocol

On ice, prepare each of following master mixes, combine, and place in heated (to 95°C) thermal cycler:

5X HF(High Fidelity) Buffer Reaction System

Component	25 µl reaction	50 µl reaction	Final Conc.
Hot Start DNA Polymerase (2000 U/mL)	0.5 ul	1 ul	2U/50ul
5X High Fidelity PCR Buffer	5 ul	10 ul	1 X
dNTP mix (10 mM each)	0.5 ul	1 ul	0.2mM
DNA Template	variable	variable	<300ng
Forward Primer (10µM)	0.5 ul	1 ul	0.2uM
Reverse Primer(10µM)	0.5 ul	1 ul	0.2uM
Distilled water	to 25 ul	to 50 ul	N/A

2.5 X GC BUFFER Reaction System

Component	25 µl reaction	50 µl reaction	Final Conc.
Hot Start DNA Polymerase (2000 U/mL)	0.5 ul	1 ul	2U/50ul
2,5X GC PCR Buffer	10 ul	20 ul	1x
dNTP mix (10 mM each)	0.5ul	1ul	0.2mM
DNA Template	Variable	variable	< 300 ng
Forward Primer (10µM)	0.5 ul	1 ul	0.2uM
Reverse Primer (10µM)	0.5 ul	1 ul	0.2uM
Distilled water	to 25 ul	to 50ul	N/A

General Cycling Conditions:

Step	Temp (°C)	Time	Cycle
Initial Denaturation	98	45 sec.	1
Denature	98	10 sec.	25 ~35
Anneal	55~65	20 ~ 30 sec.	
Extend	72	10 ~ 30 sec. **s/kb***	
Final Extension	72	5 min.	1
Hold	4-12		1

***Note:** Properly extending the extension time can improve the amplification yield. For complex amplification templates, such as genomic DNA, it is recommended to extend at a speed of 60 s/kb, and more recommended conditions please refer to the basic principles of PCR below**.**

PCR Principles

1. Template

High-quality purified DNA templates are important to high-fidelity PCR reactions. The recommended DNA template amounts with different complexity are listed below (For a 50 µL reaction):

DNA	IMPUT Amount
Plants, animals and human gDNA	**10 ng-300 ng**
E.coli, lambda gDNA	**10 ng-100 ng**
Plasmid DNA	**1 pg-10 ng**

Note: If the DNA template is obtained from a cDNA synthesis reaction, the template volume should be less than 10% of the total reaction volume. If long fragments are amplified, the amount of template input should be increased appropriately.

2. Primers

Oligonucleotide primers are typically 20-40 nucleotides in length with a GC content of 40-60%. Primers can be designed and analyzed using software such as Primer 3. The final concentration of each primer in the PCR reaction system should be in the range of 0.1-1 µM.

3. Enhancer

The Enhancer solution is an optional component to increase the amplification efficiency for problematic templates, such as GC-rich sequence or genes with strong secondary structure. Note:Since the enhancer is included in the 2.5X GC Buffer, additional enhancer is not recommended with the use of 2.5X GC Buffer. Excess amount of enhancer may be inhibitory.

4. Buffers The HS PCR Kit contains a 5X HF Buffer and a 2.5X GC Buffer.The 5X HF Buffer is designed for general high fidelity PCR amplification , and the 2.5X GC Buffer is optimized for the amplifications of GC-rich templates.

PFU DNA POLYMERASE FS-T-004

High Fidelity

Description

Pfu DNA polymerase, derived from the hyperthermophilic archae Pyrococcus furiosus, has superior thermostability and proofreading properties compared to other thermostable polymerase. Its molecular weight is 90 kDa. It can amplify DNA target up to 2 kb (simple template). The elongation velocity is 1kb/min(70~75°C). Pfu DNA polymerase possesses 3' to 5' exonuclease proofreading activity that enables the polymerase to correct nucleotide-misincorporation errors. This means that Pfu DNA polymerase-generated PCR fragments will have fewer errors than Taq-generated PCR inserts. Using Pfu DNA polymerase in your PCR reactions results in blunt-ended PCR products, which are ideal for cloning into blunt-ended vectors. Pfu DNA polymerase is superior for techniques that require high-fidelity DNA synthesis.

Description	FS-T-004
Pfu DNA Polymerase (5 U/ul)	**1000 U**
10X Pfu Buffer (MgCl2)	4x 1,25 ml
6x Loading Buffer	1ml

Activities detection conditions:

Unit Definition

1 unit of the enzyme catalyzes the incorporation of 10 nanomoles of deoxyribonucleotides into a polynucleotide fraction in 30 min at 70°C.

Storage Buffer

20mM Tris-HCl (pH 8.0), 100mM KCl, 3mM MgCl2, 1mM DTT, 0.1% NP-40, 0.1% Tween20, 0.2mg/ml BSA, 50% (V/V) glycerol.

10× Pfu Buffer with $MgCl_2$:

200mM Tris-HCl (pH 8.8), 100mM KCl, 100mM (NH4)2SO4, 20Mm MgSO4, 1% Triton X-100, 1mg/ml BSA.

Concentration:

5 u/ul

Quality control

Free of detectable, non-specific nucleases.

Storage Conditions

Store all components at -20ºC in a non-frost-free freezer.

Applications

- High-fidelity PCR and primer-extension reactions
- PCR cloning and blunt-end amplification product generation
- Site-directed mutagenesis
- Blunt-end PCR cloning

Recommended amount of template DNA:

Human genomic DNA	0.1µg-1µg
Plasmid DNA	0.5ng-5ng
Phage DNA	0.1ng-10ng
E.coli genomic DNA	10ng-100ng

Recommended Protocol

1. Add the following components to a sterile microcentrifuge tube placing on ice:

General PCR reaction mixture for 50 ul Reaction :

PCR Reaction	Volume (50 µl)	Concentration
Pfu DNA Polymerase	0.25-0.5 µl	1.25-2.5U/50 µl
10X Pfu Buffer	5 µl	1x
dNTP mix (10 mM each)	1 ul	0.2 mM each
Template DNA	variable	10 pg -1 ug
Forward Primer (10µM)	variable	04- 1µM
Reverse Primer (10 µM)	variable	04- 1µM
Distilled water	up to 50 ul	

2. Mix contents of tube. Cap tubes and centrifuge briefly to collect the contents to the bottom. When using a thermal cycler that does not contain a heated lid, overlay the reaction mixture with 25 µl mineral oil.

Recommended PCR Cycling Conditions:

Step	Temp (°C)	Time (min)	Cycle
Initial Denaturation	94	3	1
Denature	94	30 sec.	30
Anneal	55-68	30 sec.	
Extension*	72	1-3 min	
Final Extension	72	10	1

3.Incubate for an additional 10 min at 72°C and maintain the reaction at 4°C. The samples can be stored at -20°C until use.

4. Analyze the amplification products by agarose gel electrophoresis and visualize by nucleic acid dye staining. Use appropriate molecular weight standards.

Notice :

Cycling conditions may need to be optimized, depending on different primer and template combinations. For example, raise the annealing temperature to prevent non-specific primer binding, increase extension time to generate longer PCR products.

Quality Control Assay

Endodeoxyribonuclease Assay

No detectable conversion of covalently closed circular DNA to a nicked DNA was observed after incubation of 10U Pfu DNA Polymerase with 1µg pBR322 DNA for 4 hours at 37°C and 70°C.

Exodeoxyribonuclease Assay

No detectable degradation of lambda DNA-HindIII fragments was observed after incubation of 10U Pfu DNA Polymerase with 1µg digested DNA for 4 hours at 37°C and 70°C.

Ribonuclease Assay

0% of the total radioactivity was released into trichloroacetic acid-soluble fraction after incubation of 10U Pfu DNA Polymerase with 1µg E.coli [3H]-RNA (40000cpm/µg) for 4 hours at 37°C and 70°C.

Contamination Assay

Pfu DNA Polymerase was passed from quality control assay for contamination of bacterial host DNA using sequence-specific primer set from host bacterial genomic DNA.

Functional assay

Pfu DNA Polymerase was functionally tested for PCR amplifications using the various size from human genomic DNA

GOTAQ GREEN 2X MASTER G2 FS-T-5041

Cat No.	Size
FS-T-5041	500 reactions 1000 reactions

Description
GOTAQ Green 2x Master G2 is ready-to-use PCR pre-mixes are the innovation for convenience of your routine PCR. The PCR Green 2X Master is an optimized, ready-to-use PCR mixture of GOTAQ Green 2x Master G2, PCR buffer, $MgCl_2$ and dNTP's, except DNA template and primers. The mixture is suitable for amplification of most of the DNA templates and highly processive 5'→3' DNA polymerase that lacks 3'→5' exonuclease activity and lacks a 3'→5' proofreading function. PCR reactions contains two dyes (blue and yellow) can be directly loaded onto an agarose gel without the additional need of loading buffer and dyes.

Kit Contents

Contents	FS-T-5041
GOTAQ Green 2x Master G2	1 ml/ 100 reactions

Applications
GOTAQ Green 2x Master G2 is suitable and tested for amplification of genomic targets ranging from 100 bp to 4 kb and of episomal targets (lambda phage; plasmids) up to 10 kb under various reaction conditions.

- High through-put PCR
- Hot-start PCR
- Routine diagnostic PCR requiring high reproducibility
- DNA sequencing template preparation

Storage Conditions
Store all components at -20°C in a non-frost-free freezer.

Note
Do not contaminate the GOTAQ Green 2x Master G2 Master with primers and template DNA used in individual reactions. Thaw and mix all components thoroughly, spin down shortly and chill on ice.

Quality Control
No endonuclease activity, nicking activity, exonuclease activity, or priming activity has been detected.

**** Equivalent to GoTaq G2 Green Master Mix***

Recommended Protocol
This standard protocol applies to a single reaction where only template, primers, and water need to be added to the GOTAQ Green 2x Master G2 For multiple reactions, scale-up volume of reaction components proportionally. All reagents should be thawed on ice, gently mixed and briefly centrifuged before use.

1. Thaw reagents at room temperature. Mix thoroughly and then place on ice immediately after thawing.
2. Assemble reaction tubes on ice whenever possible to avoid premature, nonspecific polymerase activity.
3. The following table shows recommended component volumes:

Reaction Conditions

Component	20 µl reaction	Final Conc.
PCR Green 2X Master	10 µl	1X
10µM Forward Primer	0.2 ~ 2.0 µl	0.1~1.0 µM
10µM Reverse Primer	0.2 ~ 2.0 µl	0.1~1.0 µM
Template DNA	1 ~ 5 µl	< 250 ng
Water, RNase-Free	up to 20 µl	

NOTE: In general, use greater than 0.5 µM primers for sensitivity and less than 0.5 µM for specificity.
NOTE: Recommended amount of template per PCR reaction:
- < 50 ng plasmid or
- < 500~1000ng genomic DNA or
- 2µl of a 100µl single plaque eluate or
- one single bacterial colony

4. Ensure reactions are mixed thoroughly by pipetting or gentle vortexing followed by a brief spin in a microcentrifuge.
5. Optional-Overlay reactions with one-half volume PCR-grade mineral oil when not using heated lid on thermal cycler.
6. Transfer tubes on ice into a thermal cycler pre-warmed. The following table shows recommended cycling conditions:

PCR Conditions

Step	Temp (°C)	Time	Cycle
Initial Denaturation	95	2 min.	1
Denature	95	10 ~ 60 sec.	25 ~ 40
Anneal	50 ~ 65	10 ~ 60 sec.	
Extend	72	60 sec./kb	
Final Extension	72	5 min.	1

NOTE: Cycling conditions may need to be optimized, depending on different primer and template combinations. For example, raise the annealing temperature to prevent non-specific primer binding, increase extension time to generate longer PCR products.

7. After cycling, maintain the reactions at 4°C or store at -20°C until ready for analysis.

BLUE TAQ DNA 2X MASTER MIX FS-T-80602

Description
BlueTaq DNA 2X Master Mix is an optimized PCR premixed solution containing Taq DNA Polymerase, dNTPs, MgCl2, KCl, Bromophenol blue tracking dye, and other stabilizers. users only need to add template and primer to complete experiment. The amplified products were directly used for agarose gel electrophoresis.
It is ideally suited to routine PCR applications on various templates including pure DNA solutions, bacterial colony/ culture, and cDNA products. It can amplify 5 kb DNA from different sources genomic DNA. It is applicable to PCR reaction, colony PCR identification, rough sample amplification.

Description	FS-T-80602
BLUE TAQ DNA 2X Master Mix	5 ml/ 500 reactions

5´-3´exonuclease activity: Yes
3´-5´exonuclease activity: No
Product End: Single-base 3´ Overhangs

Storage Conditions
Store all components at -20˚C in a non-frost-free freezer.

Recommended protocol:
This standard protocol applies to a single reaction where only template, primers, and water need to be added to the Blue Taq DNA 2X Master Mix . For multiple reactions, scale-up volume of reaction components proportionally. All reagents should be thawed on ice, gently mixed and briefly centrifuged before use.

1. Thaw reagents at room temperature. Mix thoroughly and then place on ice immediately after thawing.
2. Assemble reaction tubes on ice whenever possible to avoid premature, nonspecific polymerase activity.
3. The following table shows recommended component volumes:

PCR Reactions

Component	20 µl reaction	Final Conc.
BLUE TAQ DNA 2X Master Mix	10 µl	1X
10µM Forward Primer	0.2 ~ 2.0 µl	0.1~1.0 µM
10µM Reverse Primer	0.2 ~ 2.0 µl	0.1~1.0 µM
Template DNA	1 ~ 5 µl	< 250 ng
Water, RNase-Free	up to 20 µl	

*** Note:** The optimal reaction concentration varies with different DNA templates
. Please refer to the basic principles of PCR condition below.

1. Template
Use of high quality, purified DNA templates greatly enhances the success of PCR. Recommended amounts of DNA template for a 50 µL reaction are as follows:

DNA	Imput Amount
Plants, animals and human gDNA	10 ng-100 ng
E.coli, lambda gDNA	100 pg-200 ng
Plasmid DNA	1 pg-10 ng

2. Primers
Oligonucleotide primers are generally 20–40 nucleotides in length and ideally have a GC content of 40–60%. Computer programs such as Primer3 can be used to design or analyze primers. The final concentration of each primer in a reaction may be 0.05–1 µM, typically 0.1–0.5 µM.

3. Mg2+ and Additives
In the BLUE TAQ DNA 2X Master Mix, the concentration of Mg2+ should be 4 mM, dNTP should be 300 µM. Amplification of some difficult targets, like GC-rich sequences, may be improved with additives, such as DMSO or formamide.

4. Denaturation
An initial denaturation of 3 minutes at 94℃ is sufficient for most amplicons from pure DNA templates. For difficult templates such as GC-rich sequences, a longer denaturation of 5 minutes to fully denature the template. With colony PCR, an initial 5 minutes denaturation at 94℃ is recommended to fully decompose the bacteria. During cycling a 10 seconds denaturation at 98℃ is recommended.

5. Extention
The recommended extension temperature is 65℃. The extension time is related to the length of the amplified fragment. Calculate the extension time at the speed of 1 kb/min. A final extension of 5 minutes at 65℃ is recommended.

6. Cycles
Generally, 25–35 cycles yields sufficient product. Up to 45 cycles may be required to detect low-copy-number targets.

PCR Conditions

Step	Temp (˚C)	Time	Cycle
Initial Denaturation	94	3 min.	1
Denature	98	5~ 10 sec.	30
Anneal	55~ 60	20 ~ 30 sec.	
Extend	65	1 kb/min	
Final Extension	65-68	5 min.	1
Hold	4-12° C	-	1

NOTE:The recommended pre-denaturation time for colony is :2-5 min

Quality Control
No endonuclease activity, nicking activity, exonuclease activity, or priming activity has been detected.

GC-PLATINUM POWER TAQ 2X MASTER MIX FS-T-1642-5

Description

GC-Platinum Power TAQ 2X Master MIX is a ready-to-use and complete system for rapid, consistent, and accurate amplification of long PCR products (>5~20 kb). This kit optimized for PCR amplification of genomic DNA templates up to 10 kb and lambda DNA up to 20 kb. With its enhanced processivity, yield, speed and excellent 3'→5' exonuclease and 3′→5′ proofreading activity, this enzyme is able to consistently deliver accurate and reliable amplification of long templates. This product is the ideal choice for long DNA templates unable to be amplified in conventional PCR, and is highly suitable for multiple downstream applications including complex cloning and genotyping experiments. The PCR product amplified with this mixture has one A added at 3'-end, so the product can be directly used for TA cloning.
The kit already contains blue loading dye, therefore the PCR reactions can be directly loaded onto an agarose gel without the additional need of loading buffer and dyes.

Kit Contents

Contents	FS-T-1642-5	FS-T-1642-25
GC-Platinum Power TAQ 2X Master Mix	5 ml	25 ml

1 ml= 100 Reactions (20 µl volume)

Applications

- Long range PCR
- High-fidelity PCR and primer-extension reactions
- Genotyping
- Library construction
- High GC amplification
- Next-generation DNA sequencing
- TA cloning

Storage Conditions

Store all components at -20ºC in a non-frost-free freezer.

Note

Do not contaminate the GC-Platinum Power TAQ 2X Master Mix with primers and template DNA used in individual reactions. Thaw and mix all components thoroughly, spin down shortly and chill on ice.

Quality Control

No endonuclease activity, nicking activity, exonuclease activity, or priming activity has been detected.

Template:

High-quality purified DNA templates are important to high-fidelity PCR reactions. The recommended DNA template amounts with different complexity are listed Below (For a 50µL reaction)

DNA	INPUT Amount
Plants, animals and human gDNA	10 ng~100 ng
E.coli, lambda gDNA	500 pg-200 ng
Plasmid DNA	1 pg~10 ng

Primers :

Oligonucleotide primers are typically 20-40 nucleotides in length with a GC content of 40-60%. Primers can be designed and analyzed using software such as Primer 3. The final concentration of each primer in the PCR reaction system should be in the range of 0.1-1 µM.

Recommended Protocol

This standard protocol applies to a single reaction where only template, primers, and water need to be added to the GC-Platinum Power TAQ 2X Master Mix. For multiple reactions, scale-up volume of reaction components proportionally. All reagents should be thawed on ice, gently mixed and briefly centrifuged before use.

1. Thaw reagents at room temperature. Mix thoroughly and then place on ice immediately after thawing.
2. Assemble reaction tubes on ice whenever possible to avoid premature, nonspecific polymerase activity.
3. The following table shows recommended component volumes:

Reaction Conditions

Component	20 µl	25 µl	50 µl	Final Conc
GC-Platinum POWER TAQ 2X Master Mix	10 µl	12,5 µl	25 µL	1X
10µM Forward Primer	0,4 µl	0.5 µL	1 µL	0.2 µM
10µM Reverse Primer	0,4 µl	0.5 µl	1 µL	0.2 µM
Template DNA*	Variable	Variable	Variable	>300ng
Water, RNase-Free	up to 20µl	up to 25µl	up to 50 µl	Not available

NOTE: In general, use greater than 0.5 µM primers for sensitivity and less than 0.5 µM for specificity.

4. Ensure reactions are mixed thoroughly by pipetting or gentle vortexing followed by a brief spin in a microcentrifuge.
5. Optional-Overlay reactions with one-half volume PCR-grade mineral oil when not using heated lid on thermal cycler.
6. Transfer tubes on ice into a thermal cycler pre-warmed. The following table shows recommended cycling conditions:

PCR Conditions

Step	Temp (°C)	Time	Cycles
Initial Denaturation	98	45s	1
Denaturation	98	10 sec.	30
Annealing	55 ~	30 sec.	
Extension	72	20-30 s/kb	
Final Extension	72	5 min.	1
Hold	4-12°C	∞	1

IMPORTANT: Annealing temperature should be 2-6ºC lower than the primer melting temperature. Elongation time should be ~1 min/1 kb.

NOTE: Cycling conditions may need to be optimized, depending on different primer and template combinations. For example, raise the annealing temperature to prevent non-specific primer binding, increase extension time to generate longer PCR products.

7. After cycling, maintain the reactions at 4°C or store at -20°C until ready for analysis.

GC-PLATINUM POWER TAQ 2X MASTER MIX (Blue Dye) FS-T-1641-5

Description

GC-Platinum POWER TAQ 2X Master MIX is a ready-to-use and complete system for rapid, consistent, and accurate amplification of long PCR products (>5~20 kb). This kit optimized for PCR amplification of genomic DNA templates up to 10 kb and lambda DNA up to 20 kb. With its enhanced processivity, yield, speed and excellent 3'→5' exonuclease and 3'→5' proofreading activity, this enzyme is able to consistently deliver accurate and reliable amplification of long templates. This product is the ideal choice for long DNA templates unable to be amplified in conventional PCR, and is highly suitable for multiple downstream applications including complex cloning and genotyping experiments. The PCR product amplified with this mixture has one A added at 3'-end, so the product can be directly used for TA cloning.

This Master Mix contains blue loading dye, therefore the PCR reactions can be directly loaded onto an agarose gel without the additional need of loading buffer and dyes.

Kit Contents

Contents	FS-T-1641-5	FS-T-1641-25
GC- Platinum Power TAQ 2X Master Mix (Blue dye)	5 ml	25 ml

1ml = 100 Reactions (20 µl volume)

Applications:

- Long range PCR
- High-fidelity PCR and primer-extension reactions
- Genotyping
- Library construction
- High GC amplification
- Next-generation DNA sequencing
- TA cloning

Source: The DNA Polymerase gene was induced and expressed in E.coli and obtained by separation and purification.

Thermal inactivation: No

5'-3' exonuclease activity: No

3-'5' exonuclease activity: Yes

Fast: The amplification speed for simple template is 5-10 s/kb, for complex template is 20-30 s/ kb

Note

Do not contaminate the GC-Platinum POWER TAQ 2X Master Mix with primers and template DNA used in individual reactions. Thaw and mix all components thoroughly, spin down shortly and chill on ice.

Recommended Protocol

This standard protocol applies to a single reaction where only template, primers, and water need to be added to the GC-Platinum Power TAQ 2X Master Mix. For multiple reactions, scale-up volume of reaction components proportionally. All reagents should be thawed on ice, gently mixed and briefly centrifuged before use.

1. Thaw reagents at room temperature. Mix thoroughly and then place on ice immediately after thawing.

2. Assemble reaction tubes on ice whenever possible to avoid premature, nonspecific polymerase activity.

Reaction Conditions

Component	20 µl	25 µl	50 µl	Final Conc.
GC-Platinum POWER TAQ 2X Master Mix **(Blue Dye)**	10 µl	12,5 µl	25 µL	1X
10µM Forward Primer	0,4 µl	0.5 µL	1 µL	0.2 µM
10µM Reverse Primer	0,4 µl	0.5 µl	1 µL	0.2 µM
Template DNA*	Variable	Variable	Variable	>300ng
Water, RNase-Free	up to 20µl	up to 25µl	up to 50 µl	Not available

*High-quality purified DNA templates are important to high-fidelity PCR reactions. The recommended DNA template amounts with different complexity are listed below.

Note: The optimal reaction concentration varies with different DNA templates. See table below.

Recommended PCR Program

Step	Temp (°C)	Time	Cycles
Initial Denaturation	98	45s	1
Denaturation	98	10 sec.	30
Annealing	55 ~65	30 sec.	
Extension	72	20-30 s/kb	
Final Extension	72	5 min.	1
Hold	4-12°C	∞	1

1. Template:

High-quality purified DNA templates are important to high-fidelity PCR reactions. The recommended DNA template amounts with different complexity are listed below

(For a 50µL reaction).

DNA TEMPLATE

DNA	INPUT Amount
Plants, animals and human gDNA	10 ng~100 ng
E.coli, lambda gDNA	500 pg-200 ng
Plasmid DNA	1 pg~10 ng

Note: If the DNA template is obtained from a cDNA synthesis reaction, the template volume should be less than 10% of the total reaction volume. If long fragments are amplified, the amount of template input should be increased appropriately

2. Primers :

Oligonucleotide primers are typically 20-40 nucleotides in length with a **GC content of 40-60%**. Primers can be designed and analyzed using software such as Primer 3 The final concentration of each primer in the PCR reaction system should be in the range of 0.1-1 µM.

3. Denaturation:

98°C pre-denaturation for 45 s can fully denature most DNA templates. In the case of high complexity DNA templates, the pre-denaturation time should be extended up to 3 minutes for fully denaturation.

Generally, the recommended denaturation condition for low-complexity DNA templates is 98℃, 5-10 s

4. Annealing:

The annealing temperature of GC Platinum Power TAQ 2x Master Mix is usually higher than other PCR polymerases. Generally, primers longer than 20 nt are annealed at (lower primer Tm+3)°C for 10-30 s;

When the primers are shorter than 20 nt, an annealing temperature equivalent to the lower primer Tm. When using a new primer set for PCR reaction, we recommend a gradient PCR to determine the optimal annealing temperature. In a two-step amplification protocol, the annealing temperature should be set to the extension temperature.

5. Extention:

The recommended extension temperature is 72°C. The extension time depends on the length and complexity of the amplicon. For the low-complexity amplicons (plasmid DNA), the extension condition is 10 s/kb. For high-complexity amplicons, it is recommended to increase the extension time to 20-30 s/kb. In some cases, the extension time for cDNA templates should be less than 1 min/kb

6. Cycles:

To obtain enough yield of PCR products, 25-35 cycles are recommended.

Quality Control

No endonuclease activity, nicking activity, exonuclease activity, or priming activity has been detected.

Storage Conditions

Store all components at -20°C in a non-frost-free freezer.

GOTAQ HOT START GREEN 2X MASTER MIX G2 FS-T-5141

Cat No.	Size
FS-T-5141	500 reactions 1000 reactions

Description

GO TAQ Hot Start Green Master Mix G2 is 2X Ready-to-Use Hot- start PCR pre-mixes are the innovation for convenience of your routine PCR.
The GO TAQ Hot Start Green 2X Master Mix G2 is an optimized, Ready-to-Use PCR mixture of GO TAQ Hot Start Green, PCR buffer, $MgCl_2$ and dNTP's , except DNA template and primers. The mixture is suitable for amplification of most of the DNA templates and highly processive 5'→3' DNA polymerase that lacks 3'→5' exonuclease activity and lacks a 3'→5' proofreading function. PCR reactions contains two dyes (blue and yellow) can be directly loaded onto an agarose gel without the additional need of loading buffer and dyes.

Contents	FS-T-5141
GO TAQ HS -PCR Green 2XMaster Mix G2	1 ml/100 reactions

Applications

GO TAQ Hot Start Green 2X Master Mix G2 is suitable and tested for amplification of genomic targets ranging from 100bp to 4kb and of episomal targets (lambda phage; plasmids) up to 10 kb under various reaction conditions.

- High through-put PCR
- Hot-start PCR
- Routine diagnostic PCR requiring high reproducibility
- DNA sequencing template preparation

Storage Conditions

Store all components at -20°C in a non-frost-free freezer.

Note

Do not contaminate the GOTAQ Hot Start Green 2X Master Mix G2 with primers and template DNA used in individual reactions. Thaw and mix all components thoroughly, spin down shortly and chill on ice.

Quality Control

No endonuclease activity, nicking activity, exonuclease activity, or priming activity has been detected.

**Equivalent to GoTaq G2 Hot Start Polymerase*

Recommended Protocol

This standard protocol applies to a single reaction where only template, primers, and water need to be added to the GOTAQ Hot Start Green 2X Master Mix G2. For multiple reactions, scale-up volume of reaction components proportionally. All reagents should be thawed on ice, gently mixed and briefly centrifuged before use.

1. Thaw reagents at room temperature. Mix thoroughly and then place on ice immediately after thawing.
2. Assemble reaction tubes on ice whenever possible to avoid premature, nonspecific polymerase activity.
3. The following table shows recommended component volumes:

Reaction Conditions

Component	20 µl reaction	Final Conc.
GO TAQ HS Green 2XMaster Mix G2	10 µl	1X
10µM Forward Primer	0.2 ~ 2.0 µl	0.1~1.0 µM
10µM Reverse Primer	0.2 ~ 2.0 µl	0.1~1.0 µM
Template DNA	1 ~ 5 µl	< 250 ng
Water, RNase-Free	up to 20 µl	

NOTE: In general, use greater than 0.5 µM primers for sensitivity and less than 0.5 µM for specificity.
NOTE: Recommended amount of template per PCR reaction:

- < 50 ng plasmid or
- < 500~1000 ng genomic DNA or
- 2 µl of a 100µl single plaque eluate or
- one single bacterial colony

4. Ensure reactions are mixed thoroughly by pipetting or gentle vortexing followed by a brief spin in a microcentrifuge.
5. Optional-Overlay reactions with one-half volume PCR-grade mineral oil when not using heated lid on thermal cycler.
6. Transfer tubes on ice into a thermal cycler pre-warmed. The following table shows recommended cycling conditions:

PCR Conditions

Step	Temp (°C)	Time	Cycle
Initial Denaturation	95	5 min.	1
Denature	95	10 ~ 60 sec.	25 ~ 40
Anneal	50 ~ 65	10 ~ 60 sec.	
Extend	72	60 sec./kb	
Final Extension	72	5 min.	1

NOTE: Cycling conditions may need to be optimized, depending on different primer and template combinations. For example, raise the annealing temperature to prevent non-specific primer binding, increase extension time to generate longer PCR products.

7. After cycling, maintain the reactions at 4°C or store at -20°C until ready for analysis.

HOT START PROOF 2X MASTER MIX FS-T-71702

Long Range PCR, High-Fidelity PCR, Fast PCR

Description

Host Start Proof DNA polymerase is an ideal enzyme for high fidelity PCR with excellent processivity. It is a novel engineered enzyme with unique structure.

Hot Start Proof 2X Master Mix DNA polymerase contains a recombinant synthesis enhancement domain to increase fidelity and extension speed. The antibody-mediated hot-start feature significantly inhibits non-specific amplifications at room temperature. Hot Start Proof is one of the thermostable DNA polymerases with strong 3'-5'exonuclease activity (**proofreading** activity), which results in its extreme **high fidelity.** The **Hot Start Proof** 2X Master Mix is an ideal product with good amplification efficiency for diversity templates including animals, plants, cDNA, etc.

Kit Contents

Contents	FS-T-71702-5
Hot Start Proof 2X Master Mix	5 ml

1 ml= 100 Reactions (20 μl volume)

Applications:

- Long range PCR
- High-fidelity PCR and primer-extension reactions
- Genotyping
- Library construction
- High GC amplification
- Next-generation DNA sequencing
- TA cloning

Thermal Inactivation: No
5´-3´exonuclease activity: No
3´-5´exonuclease activity: Yes
Product End: Blunt end

Standard Protocol：

-It is recommended to prepare all reaction components on ice, and then quickly transfer the reaction system to a thermocycler preheated to 98°C .

-All components should be mixed and collected at the bottom of a tube with a quick spin before use. Add Hot Start Proof DNA polymerase at the end to prevent primer degradation by its strong 3´-5´exonuclease activity.

-Note: The Hot Start Proof DNA polymerase requires special reaction conditions different from other polymerase protocols. Please refer to the recommended reaction conditions below for the better amplification yields

Note

Do not contaminate the Hot Start Proof 2X Master Mix with primers and template DNA used in individual reactions. Thaw and mix all components thoroughly, spin down shortly and chill on ice.

Components	20μL	Total Conc.
Hot Start Proof 2X Master Mix 2X Master Mix	10μL	1X
Forward Primer (10 μM)	0.4μL	0.2 μM
Reverse Primer (10 μM)	0.4μL	0.2 μM
DNA Template*	Variable	<300 ng
Nuclease-free Water	to 20μL	N/A

***Note**：The optimal reaction concentration varies with different DNA templates. Please refer to the basic principles of PCR below

Recommended PCR Program

Step	Temp.	Time	Cycles
Initial Denaturation	98°C	45s	1
Denaturation	98°C	10 sec.	25-35
Annealing	55 ~65°C	20-30 sec.	
Extension	72°C	10-30 sec.	
Final Extension	72°C	1-5 min.	1
Hold	4-12°C	∞	1

PCR Principle

1. Template:

High-quality purified DNA templates are important to high-fidelity PCR reactions. The recommended DNA template amounts with different complexity are listed below
(For a 50μL reaction).

***DNA TEMPLATE**

DNA	20 μL reaction
Plants, animals and human gDNA	4 ng - 40 ng
E.coli, lambda gDNA	200 pg-75 ng
Plasmid DNA	0,4 pg-4 ng

Note: If the DNA template is obtained from a cDNA synthesis reaction, the template volume should be less than 10% of the total reaction volume. If long fragments are amplified, the amount of template input should be increased appropriately

2. Primers：

Oligonucleotide primers are typically 20-40 nucleotides in length with a **GC content of 40-60%**. Primers can be designed and analyzed using software such as Primer 3 The final oncentration of each primer in the PCR reaction system should be in the range of 0.1-1 μM.

3. Denaturation:

98°C pre-denaturation for 45 seconds can fully denature most DNA templates. In the case of high complexity DNA templates, the pre-denaturation time should be extended up to 3 minutes for fully denaturation.
Generally, the recommended denaturation condition for low-complexity DNA templates is 98℃, 5-10 s

4. Annealing:

The annealing temperature of **Hot Start Proof TAQ 2x Master Mix** is usually higher than other PCR polymerases.
Generally, primers longer than 20 nt are annealed at (lower primer Tm+3)°C for 10-30 seconds;
When the primers are shorter than 20 nt, an annealing temperature equivalent to the lower primer Tm.
When using a new primer set for PCR reaction, we recommend a gradient PCR to determine the optimal annealing temperature. In a two-step amplification protocol, the annealing temperature should be set to the extension temperature.

5. Extention:

The recommended extension temperature is 72°C. The extension time depends on the length and complexity of the amplicon. For the low-complexity amplicons (plasmid DNA), the extension conditions is 10 s/kb. For high-complexity amplicons, it is recommended to increase the extension time to 20-30 s/kb. In some cases, the extension time for cDNA templates should be less than 1 min/kb

6. Cycles:

To obtain enough yield of PCR products, 25-35 cycles are recommended.

Store at: -20°C – avoid freezing and thawing cycles.

HOT START PROOF 2X MASTER MIX with Red Dye FS-T-82702

Long Range PCR, High-Fidelity PCR, Fast PCR

Description

Host Start Proof DNA polymerase is an ideal enzyme for high fidelity PCR with excellent processivity. It is a novel engineered enzyme with unique structure.

Hot Start Proof 2X Master Mix DNA polymerase contains a recombinant synthesis enhancement domain to increase fidelity and extension speed. The antibody-mediated hot-start feature significantly inhibits non-specific amplifications at room temperature. Hot Start Proof is one of the thermostable DNA polymerases with strong 3'-5'exonuclease activity (**proofreading** activity), which results in its extreme **high fidelity.** The **Hot Start Proof** 2X Master Mix with Red Dye is an ideal product with good amplification efficiency for diversity templates including animals, plants, cDNA, etc.

Kit Contents

Contents	FS-T-82702-5
Hot Start Proof 2X Master Mix with Red Dye	5 ml

1 ml= 100 Reactions (20 μl volume)

Applications:

- Long range PCR
- High-fidelity PCR and primer-extension reactions
- Genotyping
- Library construction
- High GC amplification
- Next-generation DNA sequencing
- TA cloning

Thermal Inactivation: No
5´-3´exonuclease activity: No
3´-5´exonuclease activity: Yes
Product End: Blunt end

Standard Protocol :

-It is recommended to prepare all reaction components on ice, and then quickly transfer the reaction system to a thermocycler preheated to 98°C .

-All components should be mixed and collected at the bottom of a tube with a quick spin before use. Add Hot Start Proof DNA polymerase at the end to prevent primer degradation by its strong 3´-5´exonuclease activity.

-Note: The Hot Start Proof DNA polymerase requires special reaction conditions different from other polymerase protocols. Please refer to the recommended reaction conditions below for the better amplification yields

Note

Do not contaminate the Hot Start Proof 2X Master Mix with primers and template DNA used in individual reactions. Thaw and mix all components thoroughly, spin down shortly and chill on ice.

Components	20μL	25μL	50μL	Total Conc.
Hot Start Proof 2X Master Mix with Red Dye	10μL	12,5 μL	25 μL	1X
Forward Primer (10 μM)	0.4μL	0.5μL	1 μL	0.2 μM
Reverse Primer (10 μM)	0.4μL	0.5μL	μL	0.2 μM
DNA Template*	Variable	Variable	Variable	<300 ng
Nuclease-free Water	to 20μL	To25μl	to50μl	N/A

***Note** : The optimal reaction concentration varies with different DNA templates. Please refer to the basic principles of PCR below

Recommended PCR Program

Step	Temp.	Time	Cycles
Initial Denaturation	98°C	3 min	1
Denaturation	98°C	10 sec.	25-35
Annealing	55 ~65°C	20-30 sec.	
Extension	72°C	30-60 sec./kb*	
Final Extension	72°C	1-5 min.	1
Hold	4-12°C	∞	1

PCR Principle

1. Template:

High-quality purified DNA templates are important to high-fidelity PCR reactions. The recommended DNA template amounts with different complexity are listed below (For a 20μL reaction).

***DNA TEMPLATE**

DNA	20 μL reaction
Plants, animals and human gDNA	4 ng - 40 ng
E.coli, lambda gDNA	200 pg-75 ng
Plasmid DNA	0,4 pg-4 ng

Note: If the DNA template is obtained from a cDNA synthesis reaction, the template volume should be less than 10% of the total reaction volume. If long fragments are amplified, the amount of template input should be increased appropriately

2. Primers :

Oligonucleotide primers are typically 20-40 nucleotides in length with a **GC content of 40-60%**. Primers can be designed and analyzed using software such as Primer 3 The final oncentration of each primer in the PCR reaction system should be in the range of 0.1-1 μM.

3. Denaturation:

The DNA templates can fully denaturation when the initial denaturation is set to 3 min.Generally, the recommended denaturation condition for low- complexity DNA templates is 98℃, 5-10 s

4. Annealing:

The annealing temperature of **Hot Start Proof TAQ 2x Master Mix** is usually higher than other PCR polymerases.
Generally, primers longer than 20 nt are annealed at (lower primer Tm+3)°C for 10-30 seconds; when the primers are shorter than 20 nt, an annealing temperature equivalent to the lower primer Tm.When using a new primer set for PCR reaction, we recommend a gradient PCR to determine the optimal annealing temperature. In a two-step amplification protocol, the annealing temperature should be set to the extension temperature.

5. Extention:

The recommended extension temperature is 72°C. The extension time depends on the length and complexity of the amplicon. For the low-complexity amplicons (plasmid DNA), the extension conditions is 30 s/kb. For high-complexity amplicons, it is recommended to increase the extension time to 60 s/kb. In some cases, the extension time for cDNA templates should be less than 1 min/kb

6. Cycles:

To obtain enough yield of PCR products, 25-35 cycles are recommended.

Store at: -20°C – avoid freezing and thawing cycles.

HIFI DNA MULTIPLE ASSEMBLY 2X MASTER MIX FS-02012

Description:

Hi Fidelity DNA Multiple Assembly Master Mix allows for seamless assembly of multiple DNA fragments, regardless of fragment length or end compatibility. It has utility for the synthetic biology community, as well as those interested in one-step cloning of multiple fragments due to its ease of use, flexibility and simple master-mix format.

Hi fi DNA Multiple Assembly 2X Master Mix is a simple, fast, and efficient seamless cloning reagent. It enables targeted cloning of inserts to any site in any vector.
Up to 5 inserts can be assembled sequentially at a time, regardless of the digestion site carried by the insert itself. To perform seamless assembly cloning, user needs to linearize the vector, design gene-specific F/R primers with 15-25 homologous bases to vector ends, so that the PCR products 5' and 3 'have the same sequence (15-25 bp) as the two ends of the linearized vector, respectively. Hi fi DNA Multiple Assembly 2X Master Mix works on the inserted fragment(s) and vector DNA, and then they are incubated at 50°C for 15-60 minutes. During incubation, specific enzymatic reactions facilitate a fully assembled DNA construct.

Hi fi DNA Multiple Assembly 2X Master Mix is the optimization form that significantly improves fragment assembly efficiency and tolerance to impurities. The final product is a fully enclosed, double-stranded DNA that can be directly used for further PCR, RCA, or other molecular biology manipulations ***(e.g., transformation into competent cells).***

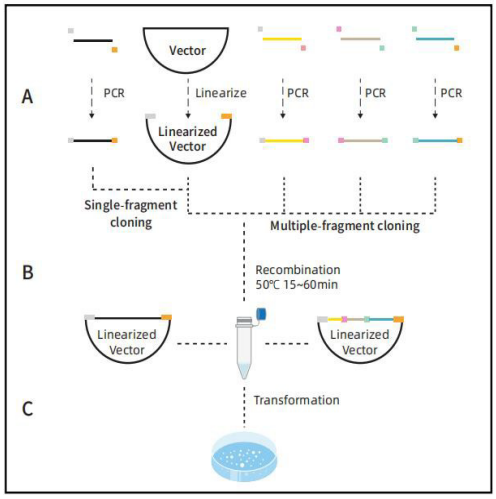
CAT.#	Description	Size
FS-02012-100	**Hifi DNA Multiple Assembly 2X Master Mix**	100μl(10 rxns)
FS-02012-500		500 μL(50 rxns)

Note: Before use, fully thaw the reagents and mix thoroughly. Keep on ice to avoid repeating freeze-thaw cycles.

Additional Materials Required, But Not Supplied

- ✧ Templates for fragment amplification, primers, and linearized vectors.
- ✧ High-fidelity DNA polymerase (to amplify fragments): HOT START PROOF PCR 2X MASTER MIX x or other equivalent products.
- ✧ DpnI (Used to remove methylated DNA templates from PCR products)
- ✧ Competent cells: For the plasmid size ≤10 kb, we recommend DH5α chemically competent cells in the transformation. For the plasmid size > 10 kb, XL10 chemically competent cells are recommended.
- ✧ Nuclease-free water, PCR tubes, Thermocyclers, LB plates with selective resistances.
- ✧

Figure 1. Hifi DNA Multiple Assembly 2X Master Mix Principle



Applications

- ✧ Rapid clone: One vector to one insert cloning
- ✧ Multiple clone: Assembly of multiple DNA fragments at same time
- ✧ Site-direct mutagenesis
- ✧ High-throughput library construction

Summary

Design primers to amplify DNA fragments (or vector) with appropriate overlapping ends. Use high-fidelity DNA polymerase in PCR amplification.
Linearize a vector by inverse PCR amplification or restriction enzyme digestion.
Quantify DNA fragments by agarose gel electrophoresis, NanodropTM or other methods.
Add the appropriate amount of DNA f ragments t o t he Hifi DNA Multiple Assembly 2X Master Mix and incubate the reaction mixture at 50 ℃ f or 15 t o 60 minutes. The i ncubation t ime depends on t he number of DNA fragments.
Transform the final product into E. coli competent cells.

Real Time qPCR Master Mix

2X ADVANCED SYBR GREEN SYBR qPCR Super Mix

2X SYBR GREEN FAST qPCR Master Mix

2X UNIVERSAL SYBR GREEN FAST qPCR Master Mix

2X UNIVERSAL POWER PLUS SYBR GREEN qPCR Master Mix UDG

2X TAQMAN PROBE qPCR Super Mix UDG V5

2X TAQMAN PROBE qPCR GENOTYPING Master Mix (UDG)

2X TAQMAN UNIVERSAL MULTIPLEX qPCR Master Mix

2X FAST EVA GREEN qPCR Super Mix

2X ADVANCED SYBR GREEN qPCR SUPER MIX FS-T-71310

Description

Advanced Sybr qPCR SuperMix is a 2X Ready-to-Use Hot-start mix containing dNTPs, Hot Start DNA polymerase, $MgCL_2$, Sybr Green dye, except primers & templates for real time quantitative PCR. The Mixture has been optimized for excellent performance and provide higher processivity. Sybr Green® dye binds directly to dsDNA generated during amplification, which permits saturation dye concentration in qPCR without PCR inhibition this dye is ideal for qPCR.

Kit Contents

Cat.n.	Description	Size
FS-T-71310 -5	2X Ad. Sybr Green qPCR Super Mix (No ROX)	5 x 1 ml
FS-T-71345-5	2X Ad. Sybr Green qPCR Super Mix (Low ROX)	5 x 1 ml
FS-T-71346-5	2X Ad. Sybr Green qPCR Super Mix (High ROX)	5 x 1 ml

1 ml = 100 reactions (20µl volume)

PCR Machines requiring ROX dye

- **High Rox Dye:**

ABI 7000, 7300, 7700, 7900HT and 7900HT Fast, StepOne, StepOne Plus:

- **Low ROX Dye:**

ABI 7500, 7500 Fast, Viia 7, QuantStudio; Roche LightCycler; Stratagene Mx3000, Mx3005P and Mx4000 :

PCR Machines requiring no ROX Dye

BioRad: iCycler, MyiQ, MiQ 2, iQ5, CFX-96, CFX-384, MJ Opticon, Option2, Chromo4, MiniOpticon

Qiagen: Roto-Gene Q, Roto-Gene3000, Roto-Gene 6000 -
Roche:LightCycler 480,LightCycler 2.0

Additional Material Required but not Supplied

1. Optical-grade qPCR tubes, plates, sealing films, and aerosol-resistant pipette tips
2. qPCR primers and probes DNA or cDNA templates

Applications

- Real-time PCR
- Gene expression profiling
- Gene knockdown verification
- Array validation

Note

Do not contaminate 2X Advanced Sybr Green qPCR Super Mix with primers, probe and template DNA used in individual reactions. Thaw and mix all components thoroughly, spin down shortly and chill on ice.

Precautions

1) Ensure that the reagent is fully thawed before use. Gently mix before use to avoid creating bubbles, then thoroughly and centrifuge before use. After use, immediately return it to -20°C storage.

2) This product contains polymerase. During use, keep it on ice. If multiple uses are required within a short period, it can be temporarily stored at 4°C. Repeated freeze-thaw cycles should be avoided as much as possible.

3) Choose an appropriate reference dye based on the qPCR machine model you are using.

4) When preparing and dispensing the reaction mixture, use sterile pipette tips, preferably those with filters.

Important Steps before reaction

General Considerations

• Primer design and amplicon length: For optimal results, use appropriate software to design primers with melting temperatures (Tm) of approximately 60°C that amplify products of 60-200 bp. For longer amplicons, extension times may need to be extended.

• Gel electrophoresis analysis of PCR products: After PCR with Sybr Green Dye, PCR products need not be stained with another DNA gel stain. Simply add DNA loading buffer to your PCR reaction solution, load on a gel, and conduct electrophoresis as usual. Gel visualization can be carried out using a 254 nm UV box, or a blue LED imager using a SYBR® Green filter. Alternatively, the gel may be imaged using a 488 nm laser-based gel scanner.

Protocol

1- The following table shows recommended component volumes:

Reaction Conditions

Reagents	10 µl	20 µl	Final Conc.
Advanced Sybr Green qPCR 2x Super Mix	5 µl	10 µl	1X
ROX Dye (10X)	(optional)	(optional)	
10 um Forward Primer*	variable	variable	0.1~0.5 uM
10 um Reverse Primer*	variable	variable	0.1~0.5 uM
Template**	50 pg to 50 ng	50 pg to 50 ng	NA
Water RNase Free	Up to 10µl	Up to 20µl	

***Please note "Use of the ROX Reference Dye only if required**

1.Ensure reactions are mixed thoroughly by pipetting or gentle vortexing followed by a brief spin in a microcentrifuge.

2.Transfer tubes on ice into a thermal cycler pre-warmed. The following table shows recommended cycling conditions

3. To increase the success rate of the reaction, it is recommended to use high-quality DNA templates. If you need to pre-mix primers and probes for stability testing at different environmental temperatures, the final primer concentration can be adjusted between 0.1-0.5 µM.

Cycling protocols

Depends on your instrument.

A. Two-step fast cycling protocol

This cycling protocol should be applicable to most amplifications where the Melting temperature of the primers is designed to be 60℃.

Cycling Step	Temp	Holding Time	Number of cycles
Enzyme activation	95℃	2 min	1
Denaturation	95°C	2-5 sec	40
Annealing	55-65°C	10 sec	
Extension / data acquisition	72°C	10-20 sec	

B. Three-step fast cycling protocol

Use this protocol when optimal primer annealing and extension temperatures are desired.

Cycling Step	Temp	Holding Time	Number of cycles
Enzyme activation	95℃	2 min	1
Denaturation	95°C	2-5 sec	40
Annealing	55-65°C	10 sec	
Extension / data acquisition	72oC	10-20 sec	

2X SYBR GREEN FAST qPCR MASTER MIX FS-T-50212

Description

Real-time Quantitative PCR (qPCR) is a powerful technique in detecting initial DNA input in a PCR reaction by fluorescence signal accumulation. The DNA double strand bonded dye, SYBR® Green I is the most commonly used dye in qPCR. 2X SYBR Green Fast qPCR Mix is provided in 3 versions: No Rox, Low ROX, High Rox and they are optimized for Real Time machines with no Rox, High Rox and Low ROX mode. It contains all required components in qPCR except primers and template. It is convenient for experiment and suitable for multiple species. The above features make it as an ideal experiment tool for gene quantitative research.

Kit Contents

Cat.n.	Description	Size
FS-T-50212-5	**2X SYBR Green Fast qPCR Mix (No ROX)**	5 X 1 mL
FS-T-50213-5	**2X SYBR Green Fast qPCR Mix (Low ROX)**	5 X 1 mL
FS-T-50214-5	**2X SYBR Green Fast qPCR Mix (High ROX)**	5 X 1 mL

1 ml =100 reactions

Compatible Instruments

Following table is helpful for choosing the right product formats

No Rox Reference Dye I	Bio-Rad iCycler serious, Roche Light Cycler serious Qiagen/Corbett serious and others
Low Rox	ABI 7500, ABI ViiATM7, ABI QuantaStudio serious, Stratagene serious, Corbett Rotor Gene 3000 and others
High Rox Reference Dye	ABI 7000/7300/7700/7900, ABI StepOne/StepOnePlus, Eppendorf and others

Materials Required

1. EP tubes, PCR tubes and other related materials.
2. qPCR specific primers and templates.
3. qPCR plates and seal membrane.

Usage Notes

1. Before using 2X SYBR Green Fast qPCR Mix, please make sure that the mix is thawed completely and then placed it on ice for use.
2. Mix other qPCR components with Mix thoroughly and gently by pipetting or vortexing. After usage, place the Mix back to -20°C for long time storage or 4°C for short period storage.
3. 2X SYBR Green Fast qPCR Mix (Low ROX) contains Hot Start *Taq* polymerase, all operation should be performed on ice.
4. 2X SYBR Green Fast qPCR Mix (Low ROX) contains low ROX dye, suits for qPCR instruments that required Low ROX mode.* See table below.
5. To avoid contamination, pipette tips with filters is suggested.
6. To guarantee better qPCR results, DNA template in good quality is suggested.

Before Use

1. Specificity of primers should be checked and a final concentration of 0.2 μM is suitable for most of primers.
2. The length of amplification products is usually range from 70 bp to 200 bp.
3. Dilute the template in gradient.
4. Add 1 pg-50 ng DNA as PCR templates and a "No Template Control sample" is suggested.
5. To ensure the confidence of experiment, at least 2 repeats of each sample is suggested.

Procedure

1. Prepare the following reaction systems on ice for a 20 ul

Component	20 ul Reaction
2X SYBR Green Fast qPCR Mix	10 μL
Forward Primer (10 μM)	0,4 μL
Reverse Primer (10 μM)	0,4 μL
gDNA or cDNA (<50 ng)	2 μL
RNase free ddH_20	up to 20 μL

2. Dissolve 2X SYBR Green Fast qPCR Mix (No Rox,Low ROX, High ROX) at room temperature then placed it on ice for further usage. Before using, agitate the Mix thoroughly and centrifuge to collect all solution.
3. Calculate the amount of mix need, generally a 10% extra amount is suggested.
4. Dispense solution in sterile PCR or EP tubes in case of any contamination.
5. Add all components listed in above table, agitate the tubes gently to mix thoroughly (avoid bubbles) and centrifuge it.
6. Dispense the reaction solution into qPCR plates and seal the plates with optical membrane.
7. 2500 rpm centrifuge the qPCR plates to collect all solution.

2. Program qPCR reaction as follows:

Stage 1	Denaturation	Reps:1	95°C	3 minutes
Stage 2	Cycles	Reps: 40-45	95°C	5 seconds
			60°C	30-34 seconds
Stage 3	Melt Curve	Reps: 1	Default	

**Confirm there is a signal collection step after each extending step. The extending time is varied according to different machines: 30s for StepOne Plus, 31s for 7300 and 34s for 7500*

Data Analysis :

1. Draw a standard curve according to Ct values of endogenous gene. The value of R^2 should be more than 0.98 and the slope of curve should be in the range of -3 to -3.5 which means the PCR amplification efficiency is in the range of 90% to 120%.
2. The standard deviation (STD) of Ct values should be less than 0.2 and the variation of Ct value for different experiment should be less than 0.5 (the threshold value of different experiments should be same when comparing Ct values).
3. The single melt curve indicate the no non-specific amplification products or primer dimmers, and theTm value in melt curve is usually in the range of 80 to 95°c.

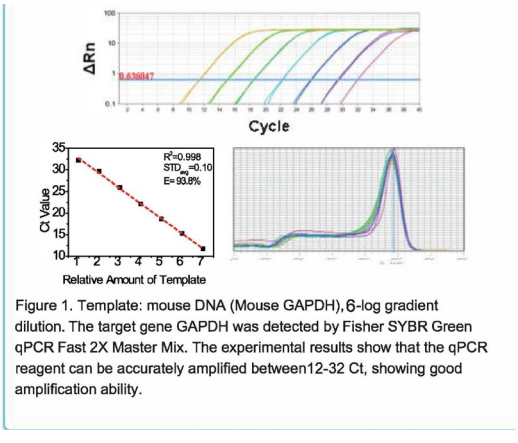


Figure 1. Template: mouse DNA (Mouse GAPDH),6-log gradient dilution. The target gene GAPDH was detected by Fisher SYBR Green qPCR Fast 2X Master Mix. The experimental results show that the qPCR reagent can be accurately amplified between12-32 Ct, showing good amplification ability.

2X UNIVERSAL SYBR GREEN FAST qPCR MASTER MIX FS-T-50215

Real-time Quantitative PCR (qPCR) is a powerful technique in detecting initial DNA input in a PCR reactionby fluorescence signal accumulation. The DNA double strand bonded dye, SYBR® Green is the most commonly used dye in qPCR.
It contains Hot Start Taq, to avoid unexpected amplification Results. It is an optimized qPCR Mix you need to add primers and template. It isconvenient for experiment and suitable for multiple species. The above features make it as an ideal experiment tool for gene quantitative research.

Kit Contents

Cat.n.	Description	Size
FS-T-50215	2 X Universal SYBR Green Fast qPCR Mix	5 X 1 mL

Compatibility:

2X Universal SYBR Green Fast qPCR Mix contains the novel designed universal reference dye, which can realize higher signal resolution and suits for all qPCR Instruments (including High ROX mode, Low ROX mode and No Rox mode).

Materials Required

EP tubes, PCR tubes and other related materials.
qPCR specific primers and templates.
qPCR plates and seal membrane.

Usage Notes

1.Before using 2X Universal SYBR Green Fast qPCR Mix,please make sure that the mix is thawed completely and then placed it on ice for use.
2.Mix other qPCR components with Mix thoroughly and gently by pipetting or vortexing. After usage, place the Mix back to -20°C for long time storage.
3.2X Universal SYBR Green Fast qPCR Mix (No ROX) contains Hot Start *Taq* polymerase, all operation should be performed on ice.
4. 2X Universal SYBR Green Fast qPCR Mix contains specially reference dye, which suits for all qPCR instruments. No Rox is required.
5.To avoid contamination, pipette tips with filters is suggested.
6.To guarantee better qPCR results, DNA template in good quality is suggested.

Before Use

Experimental Preparation
1. It is recommended to choose the amplification product length within the range of 70-200 bp.
2. It is recommended to take a reaction volume of 20 μL, add 1 pg-50 ng of DNA as a template, and set NTC (No Template Control).
3. To ensure experimental accuracy, perform triplicate technical replicates for all samples and controls.

Procedure:

Prepare the following reaction systems on ice

Components	20 ul Reaction
2X Universal SYBR Green Fast qPCR Mix	10 μL
Forward Primer (10 μM)	0,4 μL
Reverse Primer (10 μM)	0,4 μL
gDNA or cDNA (<50 ng)	2 μL
ddH20	to 20 μL

* Using 10 pg-10 ng genomic DNA or 10 pg-100 ng cDNA as the template reference quantity, gradient dilution can be performed on the template to determine the optimal template usage due to the different copy numbers of the target genes contained in the templates of different species. In addition, when using cDNA (RT reaction solution) from the two-step RT qPCR reaction as a template, the addition amount should not exceed 10% of the qPCR reaction system.
** Typically, the final concentration of the primer is 0.2 μM, and good results can be obtained , and the final concentration of 0.1-1.0 μM can be used as a reference for setting the range. In the case of low amplification efficiency, the concentration of primers can be increased; When non-specific reactions occur, the concentration of primers can be reduced and the reaction system can be optimized.

2. Program qPCR reaction as follows:

Pre-Denaturation	95°C	3 minutes	Cycles 1
Cycling	95°C	5 seconds	
	60°C	30-34 seconds**	40-45
Melt Curve	Default		

**Confirm there is a signal collection step after each extending step. The extending time is varied according to different machines: 30s for StepOne Plus, 31s for 7300 and 34s for 7500.*

Data Analysis：
1.Plot the standard curve based on Ct values and sample input quantity. The correlation coefficient (R^2) of the standard curve should be >0.98, with a slope between-3 and-3.5. The PCR amplification efficiency (E) generally falls between 90-120%. 2.
2.The standard deviation (STD) of Ct values between replicate wells should be <0.2. The STD of Ct values for the same experiment across different batches should be <0.5 (when comparing the same experiment across batches, ensure the threshold setting is essentially consistent).
3.The melting curve of the amplification product shows no significant non-specific amplification products (non-specific peaks) or primer-dimer peaks (confirm by agarose gel electrophoresis if necessary). Furthermore, the Tm value of the melting curve is typically between 80-95°C.
4. Validation of Valid Ct: A valid amplification Ct value must be less than the Ct value of the no-template control curve, while its melting curve must show no non-specific peaks.

2X UNIVERSAL POWER PLUS SYBR GREEN qPCR MASTER MIX UDG FS-T-50216

Description	FS-T-50216-5	FS-T-50216-25
2X Universal Power Plus SYBR Green qPCR Master Mix with UDG	5 X 1 mL	25 X 1mL

1 mL develops: 100 reactions (20μl)

HIGHLIGHTS:

Specific—minimize primer-dimer and non-specific amplification
• Reproducible and sensitive—consistent amplification across a wide dynamic range
• Bright—contains SYBR Green for maximum brightness
• Carry-over contamination control—contains heat-labile UDG
• Compatible with all quantitative PCR instruments

Product Description

2X Universal Power Plus SYBR Green qPCR Master Mix contains all the components needed for your real-time PCR reaction, except the template and primers, in a convenient 2X concentration premix designed to be compatible with all types of fluorescence quantitative PCR instruments on the market, including High ROX, Low ROX, and No ROX required instruments.
It utilizes a specially designed reference dye (ROX) for improved sensitivity and resolution. The reagent also incorporates a dUTP/UDG anti-contamination system, which includes UDG to degrade contaminants containing U at room temperature. UDG quickly deactivates when pre-denatured at 95℃ without affecting the efficiency and sensitivity of qPCR. Hot start Taq DNA polymerase is used for amplification, which enhances the specificity of the product while ensuring an efficient amplification effect. Overall, this product provides a reliable and versatile solution for SYBR Green-based qPCR experiments.

Storage

This product should be stored at -20℃ for long-term storage and should be protected from light.

Materials Required

1. EP tubes, PCR tubes and other related materials.
2. qPCR specific primers and templates.
3. qPCR plates and seal membrane.

Instruments

No additional reference dye is required. Universal Power Plus SYBR Green qPCR Mix with UDG is suited for all currently used qPCR instruments (including high ROX mode, low ROX mode and No ROX mode required machine).

qPCR machine Compatibility:

7500 Fast System, 7500 System, QuantStudio™ 12k Flex, QuantStudio™ 3, QuantStudio™ 5, QuantStudio™ 6 Flex, QuantStudio™ 7, StepOne™, Fast Mode, StepOne™, Standard Mode, StepOnePlus™, Fast Mode, StepOnePlus™, Standard Mode, ViiA™ 7 System, AB StepOnePlus™, Fast Mode, AB StepOne™, Standard Mode, AB 7500, Fast Mode, AB 7500, Standard Mode, AB StepOne™, Fast Mode, AB StepOnePlus™, Standard Mode

Operating instructions

Preparation before experiment

1. It is recommended to choose the amplification product length within the range of 70-200 bp.
2. It is recommended to take a reaction volume of 20 μL, add 1 pg-50 ng of DNA as a template, and set NTC (no template control).

To ensure the accuracy of the experimental results, it is recommended to weigh each sample and control group three times.

Experimental methods

Configure qPCR reaction system.
It is recommended to prepare a reaction system on ice and quickly transfer the system to a qPCR instrument preheated at 95 ℃.

Recommended Reaction 20 μL qPCR Reaction

Components	Input
2x Universal Power Plus SYBR Green qPCR Mix with UDG*	10 μL
DNA template *	2 μL
Forward primer（10 μM）**	0.4 μL
Reverse primer（10 μM）**	0.4 μL
ddH2O	To 20 μL

* Note: Using 10 pg-10 ng genomic DNA or 10 pg-100 ng cDNA as the template reference quantity, gradient dilution can be performed on the template to determine the optimal template usage due to the different copy numbers of the target genes contained in the templates of different species. In addition, when using cDNA (RT reaction solution) from the two-step RT qPCR reaction as a template, the addition amount should not exceed 10% of the qPCR reaction system.

**Note: Typically, the final concentration of the primer is 0.2 μM,and good results can be obtained , and the final concentration of 0.1-1.0 μM can be used as a reference for setting the range. In the case of low amplification efficiency, the concentration of primers can be increased; When non-specific reactions occur, the concentration of primers can be reduced and the reaction system can be optimized.

Recommended PCR Program

Steps	Temp	Time	Cycles
UDG Reaction	37℃	2 min	1
Pre Denaturation	95℃	3 min	1
Cycles	95℃	5 sec	40
	60℃	30-34 sec	
Melt Curve	Instrument automatic setting		

*Confirm there is a signal collection step after each extending step. The extending time is varied according to different machines: 30s for StepOne Plus, 31s for 7300 and 34s for 7500.

2X TAQMAN PROBE qPCR SUPER MIX UDG V5 FS-T-72225

Description

TaqMan Probe 2X qPCR Super Mix UDG V5 ready-to-use reagent ideal for most quantitative Real-time PCR applications. containing all components except primers, probes and templates. This Super Mix includes Hot start Taq DNA polymerase modified both chemically and by antibody to inhibit non- specific amplification, which can guarantee high efficiency, high sensitivity and also high reproducibility in qPCR amplification. Is is optimized by UDG anti- pollution system. It is bacterial DNA free for human and veterinary samples. This Super Mix has additional capabilities for your gene expression analysis

For viral detection and more complex templates use 10xqPCR Enhancer

Kit Contents

Contents	CAT. N°	Size
TaqMan Probe 2X qPCR Super Mix UDG V5	FS-T-72225	*500 RX
10 X qPCR Enhancer		1 vial
50x Rox Dye I (High Rox)		1 vial
50x Rox Dye II (Low Rox)		1 vail

PCR Machines requiring ROX dye

- **High Rox Dye:**
  ABI 7000, 7300, 7700, 7900HT and 7900HT Fast, StepOne, StepOne Plus:
- **Low ROX Dye:**
  ABI 7500, 7500 Fast, Viia 7, QuantStudio; Roche LightCycler; Stratagene Mx3000, Mx3005P and Mx4000 :

PCR Machines requiring no ROX Dye

BioRad: iCycler, MyiQ, MiQ 2, iQ5, CFX-96, CFX-384, MJ Opticon, Option2, Chromo4, MiniOpticon

Qiagen: Roto-Gene Q, Roto-Gene3000, Roto-Gene 6000 -
Roche:LightCycler 480,LightCycler 2.0

Additional Material Required but not Supplied

1. Optical-grade qPCR tubes, plates, sealing films, and aerosol-resistant pipette tips
2. qPCR primers and probes DNA or cDNA templates

Applications

- Real-time PCR
- Gene expression profiling
- Gene knockdown verification
- Array validation

Note

Do not contaminate the TaqMan Probe 2x qPCR Super Mix with primers, probe and template DNA used in individual reactions. Thaw and mix all components thoroughly, spin down shortly and chill on ice.

Precautions

1) Ensure that the reagent is fully thawed before use. Gently mix before use to avoid creating bubbles, then thoroughly and centrifuge before use. After use, immediately return it to -20°C storage.

2) This product contains polymerase. During use, keep it on ice. If multiple uses are required within a short period, it can be temporarily stored at 4°C. Repeated freeze-thaw cycles should be avoided as much as possible.

3) Choose an appropriate reference dye based on the qPCR machine model you are using.

4) When preparing and dispensing the reaction mixture, use sterile pipette tips, preferably those with filters.

Important Steps before reaction

(1) Ensure the correctness and specificity of primer design. Generally, a final primer concentration of 0.2 μM yields good results. If amplification efficiency is suboptimal, the primer concentration can be adjusted within a range of 0.1-1.0 μM.

(2) It is recommended that the length of the amplification product be in the range of 70-200 bp.

(3) Perform gradient dilution of the template and successively establish a standard curve.

(4) In a 25 μL reaction system, it is recommended to add 1 pg-50 ng of DNA as a template and design a NTC.

(5) To ensure the accuracy of the experimental results, it is recommended to perform each sample and control group in triplicate.

(6) **For viral detection and more complex templates**, you can add 250 μL of 10X qPCR Enhancer to 1.25 mL of 2X qPCR Taqman Probe Super Mix UDG V5, mix thoroughly by shaking, or calculate and add according to the actual usage amount.

Protocol

1- The following table shows recommended component volumes:

Reaction Conditions

	20 μl	25 μl	50μl
TaqMan Probe 2X qPCR Super Mix UDG V5	10 μl	12,5 μl	25 μl
ROX Dye (50X) *(optional)	0.4μl	0.5-0.6 μl	1μl
10um Forward Primer	0.4 μl	0.5~0.6 μl	1μl
10 um Reverse Primer	0,4 μl	0.5~0.6 μl	1μl
Probe (10μm)	0,4 μl	0.5~0.6 μl	1μl
Template	4 μl <50ng	5 μl <50ng	10 μl
Water RNase Free	up to 20ul	up to 25 ul	up to 50 ul

***Please note "Use of the ROX Reference Dye"**

2. Ensure reactions are mixed thoroughly by pipetting or gentle vortexing followed by a brief spin in a microcentrifuge.
3. Transfer tubes on ice into a thermal cycler pre-warmed. The following table shows recommended cycling conditions
4. To increase the success rate of the reaction, it is recommended to use high-quality DNA templates. If you need to pre-mix primers and probes for stability testing at different environmental temperatures, the final primer concentration can be adjusted between 0.4-1 μM.

Reaction Conditions

	20 μl	25 μl	50μl
TaqMan Probe 2X qPCR Super Mix UDG V5	10 μl	12,5 μl	25 μl
10X qPCR Enhancer**	2,0 μl	2,5 μl	5 μl
ROX Dye (50X) *(optional)	0.4μl	0.5-0.6 μl	1μl
10um Forward Primer	0.4 μl	0.5~0.6 μl	1μl
10 um Reverse Primer	0,4 μl	0.5~0.6 μl	1μl
Probe (10μm)	0,4 μl	0.5~0.6 μl	1μl
Template	4 μl <50ng	5 μl <50ng	10 μl
Water RNase Free	up to 20ul	up to 25 ul	up to 50 ul

****use the 10X qPCR Enhancer** **For viral detection and more complex templates**

2X TAQMAN PROBE qPCR 2X GENOTYPING MASTER MIX (UDG) FS-T-72212

Description

TaqMan Probe qPCR 2X Genotyping Master Mix with UDG is a ready-to-use reagent for SNP genotyping probe-based for multiplexing qPCR reactions, containing all components except primers, probes and templates. This master mix includes Hot start Taq DNA polymerase modified both chemically and by antibody to inhibit non- specific amplification, which can guarantee high efficiency, high sensitivity and also high reproducibility in qPCR amplification.

It contains dUTP/UDG anti-contamination system, where thermolabile UDG rapidly degrades uracil-containing DNA at room temperature. During the 95°C pre-denaturation step, the thermolabile UDG is quickly inactivated, preventing carryover contamination and ensuring the accuracy of genotyping. This product offers advantages such as accurate genotyping of low-concentration templates and excellent reproducibility.

Kit Contents

Contents	CAT. N°	Size
TaqMan Probe qPCR 2X Genotyping Master Mix with UDG*	**FS-T-72212**	**5ml/500 RX**
Rox Dye I (high Rox) 50X		**1 Vial**
Rox Dye II (low Rox) 50X		**1 Vial**

1ml = 100 Reactions

**Contain hot-start Taq DNA Polymerase,UDG Mg2+, dNTPs et. al.*

ROX dye - Real Time Machines:

High Rox Dye: ABI 7000, 7300, 7700, 7900HT and 7900HT Fast, StepOne,StepOne Plus:

Low ROX Dye: ABI 7500, 7500 Fast, Viia 7, QuantStudio; Qiagen: Roto-Gene Q, Roto-Gene3000, Roto-Gene 6000- Stratagene Real Time PCR System

No ROX Dye – Real Time Machines

BioRad: iCycler, MyiQ, MiQ 2, iQ5, CFX-96, CFX-384, MJ Opticon, 4, MiniOpticon Roche:LightCycler 480, LightCycler 2.0, Quiagen/Corbett

Applications

- Real-time PCR/Genotyping / PCR samples from human or animal sources

Note

Do not contaminate the TaqMan Probe 2X qPCR Genotyping Master Mix with primers, probe and template DNA used in individual reactions. Thaw and mix all components thoroughly, spin down shortly and chill on ice.

Additional Material Required but not Supplied

-Optical-grade qPCR tubes, plates, sealing films, and aerosol-resistant pipette tips

-qPCR primers and probes

-DNA or cDNA templates

Storage: Upon receipt, store all components at –20°C.

Precautions

1. Fully thaw TaqMan Probe 2X qPCR Genotyping Master Mix with UDG before use.

2. The TaqMan Probe 2X qPCR Genotyping Master Mix with UDG should be gently mixed before use to avoid generating bubbles. Mix and briefly centrifuge prior to reaction setup. After use, return immediately to the -20°C storage.

3. A Hot-start version of Taq polymerase is included in the master mix, allowing reaction. After first thaw, the master mix is stable at 4 °C for 1 week

4. Use the ROX reference dye according to the requirement of qPCR instrument to be used.

5. If applicable, use aerosol-resistant pipette tips to minimize contamination.

Experimental Preparation:

1. EP tubes, PCR tubes, pipettes, pipette tips, and ice boxes.
2. PCR probes, primers, and templates.
3. Tubes or plates specifically for quantitative PCR, along with sealing consumables
4. Reagents to be prepared by the user: DNA templates, primers, probes, and nuclease-free H_2O.

Set up: Prepare the reaction mix.

1. Fully thaw the TaqMan Probe 2X qPCR Probe Master Mix with UDG at room temperature, and gently mix well without creating bubbles. Spin down briefly in a microcentrifuge to collect all contents at the bottom.

Reaction Conditions

Reagents	20 µl reaction
TaqMan Probe qPCR 2X Genotyping Master Mix with UDG	10 µl
(10 µm) Forward Primer	0.25-0.5µl
(10 µm) Reverse Primer	0.25-0.5µl
Fluorescence Probe(10 µm)	0.25-0.5µl
Rox Dye (50X) optional*	0.2 µl
DNA Template**	100pg/100 ng
Water RNase Free	Up to 20µl

***Please note "Use of the ROX Dye on Real Time Machines"**

2. Calculate the required volume of each components based on the number of reactions to be set up and add extra 10% volume of each component to compensate pipette errors

3. Add all the common reaction components (primers and probes) in a master mix and mix thoroughly.

4. Transfer the appropriate volumes of reaction mix into qPCR plates, and carefully seal it with an optical sealing film, avoiding bubbles formation and preventing the liquid from contacting the sealing surface

5. Add templates or NTC into wells containing the qPCR reaction mix.

6. Centrifuge the qPCR plates (tubes) at 500 rpm before loading it into the instrument.

PCR Conditions

Step	Tem p (°C)	Time	Cycle
Predenaturation	95°	5min.	1
Cycling	95° 60°	5 -15 s 15-30 s	40-45
Final signal acquisition	60°	1 min.	

Notes:

* UDG enzyme is heat-sensitive and active at room temperature. It begins acting before the PCR program starts and is irreversibly inactivated during the pre-denaturation step.

* Generally, the pre-denaturation time is recommended to be no shorter than 3 minutes and no longer than 10 minutes. During cycling, the denaturation time should be no shorter than 5 seconds and no longer than 15 seconds, while the extension time should be no shorter than 10 seconds and can be adjusted according to the requirements of the primers, probes, and signal collection.

* If abnormal genotyping results occur, fluorescence collection can be collected during the annealing and extension steps to monitor signals changes throughout amplification.

2X TAQMAN UNIVERSAL MULTIPLEX qPCR MASTER MIX FS-T-70222

Description

TaqMan Universal Multiplex PCR Master Mix is a 2X ready to use master solution for multiple 5' nuclease DNA applications. The mix contains Hot Start DNA Polymerase, Dnase inhibitor, MgCl2, a passive internal reference based on proprietary ROX dye. It contains buffer enhancements to guarantee performance and reliability in all including multiplex fluorescence quantitative experiments.

This Master Mix support 3-4 target gene in a single reaction and can achieve 50 PCR cycles.

This master mix can be used for DNA detection: Genomic DNA, cDNA, plasmid DNA, and Viral Sequences.

This Master Mix can be used for gene typing and gene multiplex quantitative analysis.

Description	FS-T-70222
2X TaqMan Universal Multiplex qPCR Universal Master Mix	2 x 1.25 mL
50x Rox Dye I	100 ul
50x Rox Dye II	100 ul

- **200 rxs (25 ul)**

- Rox Dye are supplied as a separate vials.

Highlights

The Master Mix mix can provide reliable results up to 50 real-time PCR cycles.
The master mix ensure the possibility of having up to three primer pairs in the same PCR reaction, with the respective 3 probes equipped with three different fluorophores.(for example:Taqman, MGB, LNA, Molecular Beacons, Scorpions)

Compatible instruments

No Rox	Bio-Rad iCyclers/ CFX96/ CFX 384, Roche Light Cyclers® QIAGEN/Corbett Systems Eppendorf Mastercyclers®, Opticon 2
Rox Dye I	Applied Biosystems 7000/7300/7700/7900, 7900HT, Applied Biosystems Step One Plus
Rox Dye II	Applied Biosystems 7500/ViiA7, QuantStudio 7 Flex ABI, Stratagene Real-time PCR Systems, Rotor-gene3000

Additional Material Required but not Supplied

1. Optical-grade qPCR tubes, plates, sealing films, and aerosol-resistant pipette tips
2. qPCR primers and probes
3. DNA or cDNA templates

Precautions Important points before reaction setup

1.Fully thaw TaqMan Universal Multiplex qPCR Master Mix before use.
2.The TaqMan Universal Multiplex qPCR Master Mix contains glycerin. Mix gently before use without generating air bubbles. Spin briefly to collect all the contents at the bottom. After use, return it to -20°C immediately.
3.A Hot-start version of Taq polymerase is included in the master mix, allowing reaction setup at room temperature. After first thaw, the master mix is stable at 4 °C for 1 week.
4.Use the ROX reference dye according to the requirement of qPCR instrument to be used.
5.If applicable, use aerosol-resistant pipette tips to minimize contamination.
6.High quality DNA templates are recommended for optimal results.

Operation Description

Important points before reaction setup:
(1) A final primer concentration of 0.2 μM is recommended for most reactions. However, to optimize individual reaction, a primer titration from 0.1 μM to 1.0 μM can be performed.
(2) The length of amplified PCR products should ideally be in the range of 70-200 bp.
(3) Prepare a serial dilution of the template to access standard curve and test primer efficiency.
(4) Use 1 pg-50 ng of DNA template in a 20 μL reaction. The volume of template should not exceed 10% of the final PCR reaction volume.
(5) Always include a no template control (NTC) reaction.
(6) Triplicates are recommended as technical replicates in real-time PCR reactions.

Recommended Reaction

Components	25 μL
TaqMan Probe 2X Multiplex qPCR Mater Mix	12,5 μL
Forward Primer (10 μM)	0.5 μL
Reverse Primer (10 μM)	0.5 μL
Probe (10 μM)	0.5 μL
50X ROX Dye (optional)	0.5 μL
DNA Template	2,5 ul
Nuclease-free Water	Up to 25 μL

(1) Fully thaw the TaqMan Universal Multiplex PCR Master Mix at room temperature, and gently mix well without creating bubbles. Spin down briefly in a microcentrifuge to collect all contents at the bottom.
(2) Calculate the required volume of each components based on the number of reactions to be set up and add extra 10% volume of each component to compensate pipette errors.
(3) Add all the common reaction components (primers and probes) in a master mix and mix thoroughly.
(4) Dispense appropriate volumes of reaction mix into qPCR plates, and carefully seal it with an optical sealing film.
(5) Add templates or NTC into wells containing the qPCR reaction mix.
(6) Centrifuge the qPCR plates (tubes) at 2500 rpm to collect all the contents at the bottom of wells. The samples are ready for thermocycling.

Step	Temp	Time	Cycles
Predenaturation	95°C	3 min	1
Denaturation	95°C	15 s	40-50
Annealing and extension	55°C	30 s	

2X FAST EVA GREEN® qPCR SUPER MIX FS-T-41310

Description

Fast EvaGreen qPCR SuperMix is a 2X Hot Start mix containing dNTPs, HotStart Taq,$MgCl_2$, EvaGreen dye.
The Master Mix was formulated to for Fast Cycling PCR, and can be used for regular cycling protocols.
The optional **40x template buffer** helps to track where DNA templates have been added to the reaction mixes. It offers high sensitivity, robust fluorescence signal amplification.
EvaGreen® dye binds directly to dsDNA generated during amplification, which permits saturation dye concentration in qPCR without PCR inhibition this dye is ideal for both qPCR and HRM High Resolution Melting analysis
The absorption and fluorescence emission spectra of DNA-bound EvaGreen® Dye are very similar to those of SYBR® Green I or FAM with Ex/Em at 500/530 nm with DNA.

Kit Components

Cat.n.	Fast Eva Green qPCR Super Mix	Size
FS-T-41310-5	2X Fast Eva Green qPCR Super Mix (No ROX)	5 x 1 ml
	40x Template Buffer	2 x 1 ml
FS-T-41315-5	2X Fast Eva Green qPCR Super Mix (Low ROX)	5 x 1 ml
FS-T-41316-5	2X Fast Eva Green qPCR Super Mix (High ROX)	5 x 1 ml

1 ml = 100 reactions (20µl volume)

Additional Material Required but not Supplied

1. Optical-grade qPCR tubes, plates, sealing films, and aerosol-resistant pipette tips
2. qPCR primers and probes DNA or cDNA templates

Applications

- Real-time PCR
- Gene expression profiling
- Gene knockdown verification
- Array validation

Note

Do not contaminate 2X Fast EVA Green qPCR Super Mix with primers, probe and template DNA used in individual reactions. Thaw and mix all components thoroughly, spin down shortly and chill on ice.

Precautions

1) Ensure that the reagent is fully thawed before use. Gently mix before use to avoid creating bubbles, then thoroughly and centrifuge before use. After use, immediately return it to -20°C

2) This product contains polymerase. During use, keep it on ice. If multiple uses are required within a short period, it can be temporarily stored at 4°C. Repeated freeze-thaw cycles should be avoided as much as possible.

3) Choose an appropriate reference dye based on the qPCR machine model you are using.

4) When preparing and dispensing the reaction mixture, use sterile pipette tips, preferably those with filters.

General Considerations

• Primer design and amplicon length: For optimal results, use appropriate software to design primers with melting temperatures (Tm) of approximately 60°C that amplify products of 60-200 bp. For longer amplicons, extension times
may need to be extended.
• Gel electrophoresis analysis of PCR products: After PCR with EVA Green Dye, PCR products need not be stained with another DNA gel stain. Simply add DNA loading buffer to your PCR reaction solution, load on a gel, and conduct electrophoresis as usual. Gel visualization can be carried out using a 254 nm UV box, or a blue LED imager using a SYBR® Green filter. Alternatively, the gel may be imaged using a 488 nm laser-based gel scanner.

Protocol

1- The following table shows recommended component volumes:

Reaction Conditions

Reagents	10 µl	20 µl	Final Conc.
2x Fast EVA Green qPCR Super Mix	5 µl	10 µl	1X
40x Template Buffer (optional)***	(optional)	(optional)	
ROX Dye (10X)	(optional)	(optional)	
10 um Forward Primer*	variable	variable	0.1~0.5 uM
10 um Reverse Primer*	variable	variable	0.1~0.5 uM
Template**	50 pg to 50 ng	50 pg to 50 ng	NA
Water RNase Free	Up to 10µl	Up to 20µl	

***Please note "Use of the ROX Reference Dye only if required**

***The use of Template Buffer** is optional, but all reactions in a given experiment should contain the same amount for accurate comparisons. Template Buffer should be at 1X in the final reaction. If 1 uL of DNA is to be added to each 20 uL reaction, mix 40X Template Buffer with DNA at a ratio of 0.5 uL Template Buffer per 1 uL DNA, then add 1.5 uL of the mix to each reaction. If 5 uL of DNA is to be added to each reaction, mix at a ratio of 0.5 uL 40X Template Buffer per 5 uL DNA, and then add 5.5 uL of the mix per reaction. When using small volumes of template it may be convenient to dilute 40X Template Buffer with PCR grade water prior to use. For example, you could mix 1 uL of 20X Template Buffer per 1 uL DNA, then add 2 uL of the mix to each reaction. Note: **Template Buffer quenches ROX fluorescence**. Refer to Table 1 for the recommended ROX concentrations when Template Buffer is used.

Template concentration: The optimal amount of template DNA varies by application. We recommend 50 pg to 50 ng genomic DNA per reaction. For two-step RT-PCR: the A260 measurement of a reverse transcription reaction does not accurately quantify cDNA. Add undiluted or diluted cDNA from a RT reaction (generated from < 1 ug RNA), but the RT reaction volume must not exceed 10% of the final PCR volume.

Cycling protocols

Depends on your instrument.

A. Two-step fast cycling protocol

This cycling protocol should be applicable to most amplifications where the Melting temperature of the primers is designed to be 60℃.

Cycling Step	Temp	Holding Time	Number of cycles
Enzyme activation	95℃	2 min	1
Denaturation	95°C	2-5 sec	40
Annealing	55-65°C	10 sec	
Extension / data acquisition	72°C	10-20 sec	

B. Three-step fast cycling protocol

Use this protocol when optimal primer annealing and extension temperatures are desired.

Cycling Step	Temp	Holding Time	Number of cycles
Enzyme activation	95℃	2 min	1
Denaturation	95°C	2-5 sec	40
Annealing	55-65°C	10 sec	
Extension / data acquisition	72oC	10-20 sec	

Reverse Transcription Enzymes & Kits

qRT-PCR Master Mix

mi-RNA FIRST STRAND-SYNTHESIS KIT

cDNA First Strand Synthesis Kit

MMLV R-Transcriptase (RNase H+)

RNase Inhibitor (40 U/µl)

Oligo d(T)23VN (50uM)

Random Primer Mix (60uM)

SCRIPT-III ONE STEP RT- qPCR TAQMAN PROBE (UDG)

2X One Step Sybr Green RT-PCR Mix

VET PLUS ONE STEP TAQMAN PRPBE RT-qPCR MULTIPLEX

mi-RNA cDNA FIRST STRAND-SYNTHESIS KIT FS-RT-1034

Introduction

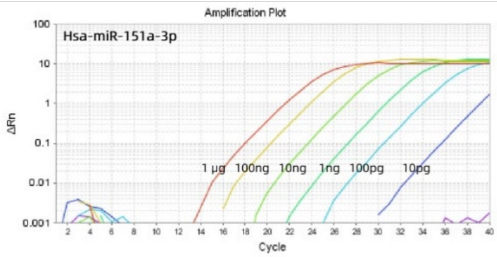
This kit is suitable for cDNA first strand synthesis using microRNA as template through the tail addition method, where the Poly (A) tail addition reaction and reverse transcription reaction at the 3 'end of miRNA can be efficiently carried out simultaneously.

miRNA-A Enzyme Mix contains Poly (A) Polymerase (PAP) and reverse transcriptase. PAP is mainly used to add Poly (A) tails at the 3 'end of RNA molecules, and can also specifically recognize single stranded RNA, effectively avoiding RT reactions of pre-miRNA with double stranded or stem-loop structures.

The modified reverse transcriptase lacks of RNase H activity and increases its affinity with RNA, resulting in a significant improvement in the efficiency and sensitivity of miRNA reverse transcription. The obtained cDNA can be directly used for qPCR detection using either SYBR Green dye-base or Taqman probe-base reagent.

Kit Components

Components	FS-RT-1034
miRNA-A Enzyme Mix (20X)	20 µl
miRNA-A Reaction Buffer (2X)	200 µl
Universal RT Primer	60 µl
Universal miRNA-A qPCR Primer R (10 µM)*	200 µL
U6 qPCR Primer F (10 µM)**	100 µL
Nuclease-free ddH2O	1 mL



***Universal miRNA-A qPCR Primer R (10µM) can be used together with designed qPCR forward primers for qPCR detection.**
****U6 qPCR Primer F, a universal reference forward primer for human, mouse and rat U6, can be used together with Universal miRNA-A qPCR Primer R for qPCR detection.**

HIGHLIGHTS

- . High specificity: The kit only performs Poly (A) tail addition reaction and reverse transcription reaction on single-stranded miRNAs, avoiding interference from pre-miRNAs with secondary structure;
- Convenient and fast: Poly (A) tail addition reaction and reverse transcription reaction can be completed in one preparation.
- High sensitivity: Total RNA as low as 10 pg can be detected

Ordering Information

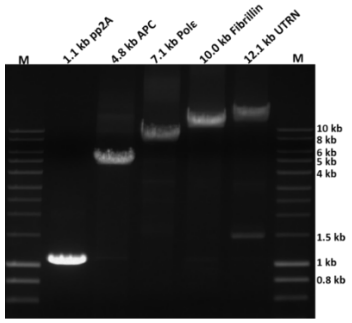
Cat.#	Description	Size
FS-RT-1034	miRNA First Strand cDNA Synthesis Kit	20 Reactions (20 µl)

cDNA FIRST STRAND SYNTHESIS KIT FS-RT-4020

Introduction

cDNA First Strand Synthesis Kit features two optimized mixes: cDNA ***Enzyme Mix*** and cDNA ***Reaction Mix.*** The enzyme mix combines Reverse Transcriptase and RNase Inhibitor, and the reaction mix contains an optimized buffer. The Reverse Transcriptase is a recombinant M-MuLV reverse transcriptase with reduced RNase H activity and increased thermostability.
It can be used to synthesize first strand cDNA at higher temperatures than the wild type M-MuLV.
The enzyme is active up to 48°C, which provides higher specificity and higher yield of cDNA.
The kit also provides two optimized primers for reverse transcription and nuclease-free water.
An anchored Oligo-dT primer [$d(T)_{23}VN$] forces the primer to anneal to the beginning of the poly A tail.
The optimized Random Primer Mix provides random and consistent priming sites covering the entire RNA templates including both mRNAs and non-polyadenylated RNAs. The first strand cDNA product generated is up to 10 kb.

Kit components	FS-RT-4020 100 RXN	FS-RT-4021 200 RXN
cDNA Enzyme Mix (10X)	200 µL	2X 200 µL
cDNA Reaction Mix (2X)	1 ml	2X 1 ml
Oligo d(T)23VN * (50 µM) **	200 µL	2x200 µL
Random Primer Mix (60 µM)**	200 µL	2x200 µL
dNTPs (10 mM each)	100 µL	2x 100 µL
Nuclease-free H2O	1,25 ml	2x 1,25 ml



Using 1µg of HeLa cell total RNA as template, reverse transcription was performed with cDNA First-Strand Synthesis Kit, and ***1µL of cDNA*** was used as template to obtain clear and specific electrophoresis bands for genes of different length

* V = A, G or C; N = A, G, C or T.
** Contains 1 mM dNTP.

Highlights

- The low activity of RNase H is beneficial to the synthesis of long fragment cDNA, which can synthesize cDNA up to 10 KB.
- The optimum temperature of the enzyme was 42℃ and the enzyme still has high activity at 48℃.
- The specific cDNA yield was higher than that of ordinary M-MLV reverse transcriptase (37℃).
- Suitable for coronavirus detection;
- With a variety of RT primers, both mRNA and ncRNA can be used as reverse transcription templates.

Quality control

The performance of cDNA First Strand Synthesis Kit is tested in an RT reaction using Jurkat total RNA with primer $d(T)_{23}VN$. The length of cDNA achieved is verified by detection of a 9.2 kb amplicon of fibrillin gene.

First Strand cDNA Synthesis Reaction

1. Denaturation of RNA and primer at 65 –70° C for 5 minutes can remove secondary structures that may impede long cDNA synthesis. However, this step can be omitted in many cases (unpublished results).

2. We recommend incubation at 42 ° C for one hour for maximum yield and length. However, many targets can be detected after a much shorter incubation time. For example, 10 minutes incubation can be used for up to 5 kb cDNA synthesis.

M-MLV REVERSE TRANSCRIPTASE FS-RT-1032

M-MLV Reverse Transcriptase is a recombinant M-MuLV reverse transcriptase with reduced RNase H activity and increased thermostability. It can be used to synthesize first-strand cDNA at higher temperatures than the wild type M-MuLV. The enzyme is active up to 48°C, providing higher specificity, higher yield of cDNA and more full-length cDNA product up to 12 kb.

Concentration: 200U/ul

Cat N°	Size	Storage/Shelf life
FS-RT-1032	10,000 U	-20°C/one year

Kit Components:

FS-RT-1033	Size
M-MLV Reverse Transcriptase (200U/ul)	50 ul
5X First-Strand Buffer	500 ul
100 mM DTT (10X)	200 ul

Reaction Conditions:
1X First-Strand Reaction Buffer, 10 mM DTT, 200 units M-MLV Reverse Transcriptase, supplemented with 0.5 mM dNTPs (not included) and 5 µM dT23VN (not included). Incubate at 42°C for 50 minutes. If random primers are used, a 10-minute incubation at room temperature is recommended before transferring to 42°C.

RNase INHIBITOR (40 U/ul) FS-RT-1152-1

RNase Inhibitor is a recombinant RNase inhibitor expressed in soluble form in Escherichia coli. It has the same application effect as a specific ribonuclease inhibitor present in human placenta. Its essence is a protein with a molecular weightof 51,000 Da, etc. The pH of the electrical point is 4.7.

RNase Inhibitor can specifically bind RNase A, B, and C with a non-covalent bond toform a 1:1 complex to inactivate RNase, and has a broad spectrum of RNase inhibitory activity. RNasin is active in buffers of 0-0.5 M NaCl, pH 5-8, and has the highest activity at pH 7.8. RNasin protects the integrity of mRNA and improves theefficiency of transcription and translation, while avoiding the possible effects of using organic compound inhibitors.

Description	FS-RT-1152-1	FS-RT-1152-5
RNase Ihibitor 40U/µl	1,000 units	5,000 units

Applications: First-strand cDNA synthesis, isolation of polysomes, in vitro translation, in vitro cell-free system transcription, in vitro transcription of SP6 or T7 RNA polymerase.

Oligo d(T)23VN (50 µM) FS-ODT-50

Oligo d(T)23VN is used for the priming and sequencing of mRNA adjacent to the 3'-poly A tail.
V = A or G or C
N = A or G or C or T Reconstitute with water.
Applications: RT-qPCR, RT-PCR and cDNA Synthesis, PCR.

Description	CAT.#	Size
Oligo d(T)23VN (50 µM)	FS-OTD-50	200 ul

Random Primer Mix (60 µM) FS-RH-50

High-quality DNA hexamers of randomized sequence Random Primers are random hexadeoxynucleotides that can be used for first-strand cDNA synthesis and cloning. They are also available as components of the Reverse Transcription System

Description	CAT.#	Size
Random Primer Mix (60 µM)	FS-RH-50	200 ul

SCRIPT-III ONE STEP RT- qPCR TAQMAN PROBE (UDG) FS-RT-21402

Description

Script-III One Step RT-qPCR Taqman Probe Kit with UDG a ready-to-use kit allowing reverse transcription and subsequent probe-based qPCR in a single tube. It contains all components for RT-qPCR except primers, probes and RNA templates. The one-step format significantly improves sensitivity and effectively prevent contamination. The heat-liable UDG in this product could degrade U-contained contamination in room temperature, and inactivated in 50°C, which could prevent false positive results without affect the efficiency and sensitivity. The Script Reverse Transcriptase in the kit provides reliable reverse transcription to a wide range of RNA template amount. After reverse transcription, the Hot-start version of Taq polymerase is activated at 95 ° C and the Script Reverse Transcriptase is inactivated simultaneously. In the sequential PCR reaction,the 5'-3' exonuclease activity of Taq polymerase cleaves the hybridized probe, separating the reporter from the quencher and releasing fluorescent signal. The Script-III One Step RT-qPCR Probe Kit is an ideal product for high-speed

Kit Contents

Contents	Cat.#	Size
2xOne Step RT-qPCR Probe Buffer IV*	FS-RT-21402	1.25ml x 2
One Step Probe Enzyme Mix IV**		500µl
50X ROX Dye I (High Rox)***		100µl
50X ROX Dye II (Low Rox)***		100µl
Nuclease Free Water H_2O		1.25ml x 2

* Containing dNTP/dUTP Mix, prevent false positive caused by cross contamination with UDG.

** the Taq polymerase is blocked by antibody, containing RNase Inhibitor, Heat-labile UDG

*** Passive reference dye to normalize the fluorescence signals

Applications

- Real-time PCR
- Detection and quantification of DNA and cDNA targets
- Gene expression profiling
- Microbial detection
- Viral load determination
- Array validation
- SNP genotyping

Storage Conditions

Upon receipt, store all components at –20°C.

Use of the ROX Reference Dye:

-50x Rox Dye I (High Rox)

Applied Biosystems 7000/7300/7700/7900, Applied Biosystems StepOne™/StepOnePlus™.

-50x ROX Dye II (Low Rox)

Applied Biosystems 7500/ViiA7TM, QuantStudio™, Stratagene Real-time PCR Systems, Rotor-gene™ 3000

-NO ROX Dye

Bio-Rad iCyclers/ CFX96/ CFX 384, Roche Light Cyclers®, QIAGEN/Corbett Systems, Eppendor Mastercyclers

Recommended Protocol

1.Fully thaw the 2X One Step RT-qPCR Probe Buffer IV before use. Mix the buffer well and avoid directly sunlight. Determine the total number of reactions required and prepare master mix. Triple replicates for each reaction are recommended.

2.The One Step Probe Enzyme Mix IV contain high concentration of glycerin. Mix gently before use without generating air bubbles. Spin briefly to collect all the contents at the bottom. After use, return it to -20°C immediately.

3.If applicable, use aerosol-resistant pipette tips and microtubes to minimize contamination.

4.High quality RNA templates are recommended for optimal results

5.Only gene specific primers are recommended. Random primers and Oligo dT primers are NOT recommended in the reverse transcription reaction.

6.The optimal length of amplicon is between 70 and 200 bp for general cycling condition.

Prepare materials before reaction setup:

Pipette, aerosol-resistant pipette tip, cold blocks and ice.
Gene expression primers and probes.
RNA templates.
1.5 mL RNase-free EP tubes, Real-time PCR tubes and plates.

1. Prepare the reaction mix :

Set up the reaction on ice by adding the following components for the number of reactions required. :

Reaction Conditions

Component	20 µL reaction	25 µL reaction	50 µL reaction
2xOne Step RT-qPCR Probe Buffer IV	10 µl	12,50µl	25µl
One Step Probe Enzyme Mix IV	2 µl	2,5 µl	5µl
10uM Forward Primer*	0.4 µl	0.5-0.6 µl	1µl
10uM Reverse Primer*	0.4 µl	0.5-0.6 µl	1µl
TaqMan Probe (10µM)***	0.4 µl	0.5-0.6 µl	1µl
50X Rox Dye (optional)	0.4 µl	0.5-0.6 µl	1µl
Totatal RNA **	2 µL	2,5 µL	5µl
Water, RNase-Free	Up to 20µl	up to 25 µl	up to 50 µl

* A final primer concentration of 0.2 µM is recommended for most reactions. However, to optimize individual reaction, a primer titration from 0.1 µM to 1.0 µM can be performed. The length of amplified PCR products should ideally be in the range of 70 - 200bp.

** Use 10 pg~100 ng of RNA template in a 20 µL reaction.

*** A Probe concentration of 50-250 nM is recommended.

Optimized One Step RT-qPCR Conditions

Step	Temp (°C)	Time	Cycle
UDG Reaction	25°C	5 min.	1
Reverse Transcription	50°C	5 min.	1
Polymerase Activation	95°C	3 min.	1
Denaturation Annealing, and Extention	95° C	5-15 sec.	45
	60° C	30-34 sec.	

The extension time should be adjusted to the minimum time required for data acquisition according to qPCR instrument guidelines used. (30 s for Applied Biosystems StepOnePlusTM, 31 s for Applied Biosystems 7300, and 34 s for Applied Biosystems 7500)

2X ONE STEP SYBR GREEN RT-PCR Mix FS-RT-007

Description

One Step SYBR Green RT-qPCR Kit is a special kit for one-step RT-qPCR reaction by using SYBR Green I chimeric fluorescence. The kit takes RNA as the template, uses gene-specific primers, and reverse transcription and PCR reaction can be carried out continuously in the same tube without additional pipe opening and pipetting operations, greatly improving the detection flux and effectively preventing contamination. This reaction system can detect the amplification products in real time, greatly improving the detection sensitivity, and omitting the electrophoresis step after PCR reaction, which is very suitable for the detection of RNA virus and other trace RNA.

This product integrates the superiority of Reverse Transcriptase and Taq DNA Polymerase, cooperate with optimized buffer system, with high amplification efficiency and high amplification specificity, one-step RT-qPCR reaction can be stable. In addition, all the enzymes used in the reaction are made into enzyme Mix, which is easier and more convenient to operate.

Storage: -20℃, protect from light

Kit Contents: 500 Reactions (20ul reaction volume)

Contents	Cat.#	Size 500 rxns
2xOne Step Sybr Green RT-qPCR Buffer*	FS-RT-007	4 X 1.25ml
One Step Enzyme Mix**		2x 200ul
50X ROX Dye Reference Dye ***		2 x 100ul
50X ROX Dye Reference II (Low Rox)***		2 x 100ul
RNase Free ddH_2O		4x 1.25ml

** contain dNTPs, Mg2+, SYBR Green, etc.*
*** contain Reverse Transcriptase，RNase Inhibitor ,Taq DNA Polymerase.*
**** Use to correct the error of fluorescence signal between holes.*

Instruments:

ROX Types	qPCR Machines
No Rox	Bio-Rad iCycler serious, Roche Light Cycler serious, Qiagen/Corbett serious and others
ROX Reference Dye I	ABI 7000/7300/7700/7900, ABI StepOne/StepOnePlus, Eppendorf and others
ROX Reference Dye II	ABI 7500, ABI ViiATM7, ABI QuantaStudio serious, Stratagene serious, Corbett Rotor Gene 3000 and others

Usage Notes

1. When using 2X One Step SYBR Green RT-qPCR Buffer, please fully melt it, mix it well and then use it. Avoid direct sunlight and keep it away from light. If multiple One Step RT-qPCR reactions need to be prepared at the same time, it is recommended to allocate all groups except primers and templates to make premix, and then separate it into each reaction tube to reduce the loss of reagents.
2. The One Step Enzyme Mix in the kit contains high concentration of glycerol, before using the Enzyme Mix, pleasemix gently and avoid foaming; Please use immediately after centrifugal. After usage please put it back -20℃ refrigerator.
3. The configuration and split charging of the reaction liquid must use non-polluting spear and Microtube to avoid contamination as far as possible.
4. To ensure the success of the reaction, it is recommended to use high-quality RNA templates.
5. The kit can only use specific primers, and can't use random primers or Oligo dT primers for reverse transcription reaction.
6. When the one-step RT-qPCR experiment is designed for amplification of primers, the recommended product length of 70-200 bp has the best effect.

Protocol

Provision of Experiment

1. 1.5 mL RNase-free EP tubes, RNase-free PCR tubes, pipettors and spears, ice or ice box.
2. PCR specific primers and templates.
3. Real-time PCR special tubes or plates.

Method of Experiment

Prepare reagents：RNA templates、primers. Please follow the instructions of different brands of fluorescence quantitative PCR instrument for experimental operation.

1. Preparation of One Step RT-qPCR reaction system
Prepare the following reaction system on the ice.
Take the 20 ul reaction system as an example:

Components	Volume	Volume
2X One Step SYBR Green RT-qPCR Buffer	10 ul	25 ul
One Step Enzyme Mix	0.8ul	2ul
Forward primer (10 μM) *	0.4 ul	1 ul
Reverse primer (10 μM) *	0.4 ul	1 ul
ROX Reference Dye I (50X)	0.4 ul	1 ul
Total RNA **	2 ul	5 ul
RNase-free H2O	To 20 ul	To 50 ul

* The final concentration of primer is usually 0.2um which can get better results. When the reaction performance is poor, the primer concentration can be adjusted in the range of 0.1-1.0 uM. The length of the amplification product is recommended to be within the range of 70-200 bp.
** It is recommended to input 50 pg~300 ng Total RNA as template in 20 ul system.

2. One Step RT-qPCR reaction procedure

Step	Temperature	Time	Cycles
Reverse Transcription	42℃	5 min	1
Pre-denaturation	95℃ 95℃	1 min 5 sec	1
Circular reaction	60°C	30~34 s *	40

Melt Curve (automatic instrument setting)

** Please adjust the extension time according to the minimum time limit for data collection required by your Real Time PCR device: set it to 30 sec. when using Step One Plus;

Please set it to 31 s when using 7300; Set it to 34 s when using 7500.
Confirm the amplification curve and melting curve after the reaction, make the standard curve.

VET PLUS FAST ONE STEP TAQMAN PROBE RT-qPCR MULTIPLEX FS-RT-72413

Description

VET PLUS FAST One Step TAQMAN PROBE RT-qPCR Master Mix is a ready-to-use kit Multiplexing, allowing Fast-Cycle Amplification,One Step reverse transcription qPCR in a single tube on Animal sourced samples.It is supplied with a separate vial of ROX Dye, and contains all components for RT-qPCR except primers, probes RNA templates. The one-step format significantly improves sensitivity and effectively prevent contamination. The heat-liable dUTP/UDG prevents contamination and degrades uracyl-contaminants at RT, to avoid false positive results. At 50°C during reverse transcription the heat-labile UDG quickly deactivates ensuring the efficiency and sensitivity of RT-qPCR. The Script Reverse Transcriptase in the kit provides reliable reverse transcription to a wide range of RNA template amount.

Kit Contents

Contents	Cat.#	Size
Vet Plus Fast One Step Taqman Probe RT-qPCR Buffer *	FS-RT-72413	1 ml x 2 (500 tests)
Vet Plus Fast One Step Taqman Probe RT-qPCR Enzyme Mix **		500µl
50X ROX Dye I (High Rox)***		100µl
50X ROX Dye II (Low Rox)***		100µl

1 ml (100 reactions)

* Containing dNTP/dUTP Mix, prevent false positive caused by cross contamination with UDG.

** the Master Mix is blocked by antibody, containing RNase Inhibitor, Heat-labile UDG

*** Passive reference dye to normalize the fluorescence signals

Applications

- Real-time qPCR – Multiplexing Fast Cycles
- Detection and quantification of DNA and RNA targets

Storage Conditions

Upon receipt, store all components at –20°C.

Use of the ROX Reference Dye:

-50x Rox Dye I (High Rox)

Applied Biosystems 7000/7300/7700/7900, Applied Biosystems StepOne™/StepOnePlus™.

-50x ROX Dye II (Low Rox)

Applied Biosystems 7500/ViiA7TM, QuantStudio™, Stratagene Real-time PCR Systems, Rotor-gene™ 3000

-NO ROX Dye

Bio-Rad iCyclers/ CFX96/ CFX 384, Roche Light Cyclers®, QIAGEN/Corbett Systems, Eppendor Mastercyclers

Recommended Protocol

1. Fully thaw the Vet Plus Fast One Step Taqman Probe RT-qPCR Buffer, and the Master Mix before use.

2. The Vet Plus One Step Taqman Probe RT-qPCR Enzyme Mix contains high concentration of glycerin. Mix gently before use without generating air bubbles. Spin briefly to collect all the contents at the bottom. When preparing multiplexing reactions simultaneously, prepare a mixture of all the reagents and aliquote into each reaction tube, to avoid loss. After use, return it to -20°C immediately.

3. If applicable, use aerosol-resistant pipette tips and microtubes to minimize contamination.

4. High quality RNA templates are recommended for optimal results

5. Only gene specific primers are recommended.
Random primers and Oligo dT primers are NOT recommended in the reverse transcription reaction.

6. The optimal length of amplicon is between 70 and 200 bp for general cycling conditions.

Prepare materials before reaction setup:

Pipette, aerosol-resistant pipette tip, cold blocks and ice.

Gene expression primers and probes.

RNA templates.

1.5 mL RNase-free EP tubes, Real-time PCR tubes

and plates.

1. **Prepare the reaction mix** :

Set up the reaction on ice by adding the following components for the number of reactions required. :

Reaction Conditions

Component	20 µL reaction
Vet Plus Fast One Step Taqman Probe RT-qPCR Buffer	3,2 µl
Vet Plus Fast One Step Taqman Probe RT-qPCR Enzyme Mix	0.8 µl
10uM Forward Primer*	X
10uM Reverse Primer*	X
Probe (10µM)***	X
50X Rox Dye (optional)	0.4 µl
Total RNA **	2 µL
Water, RNase-Free	Up to 20µl

* A final primer concentration of 0.2 µM is recommended for most reactions. However, to optimize individual reaction, a primer titration from 0.1 µM to 1.0 µM can be performed. The length of amplified PCR products should ideally be in the range of 70 - 200bp.

** Use 10 pg~100 ng of RNA template in a 20 µL reaction.

*** A Probe concentration of 50-250 nM is recommended.

***Note: 1)** The amount of primers/probe needs to be titrated for the desired concentration for fast programs, which may differ from standard reactions .

The optimal concentration range for primers and probes in the FAM channel is 0.16-0.32 µM, and for the VIC/ROX/TAMARA channels, the concentration range is 0.32-0.48 µM.

T7 HIGH YIELD RNA SYNTHESIS KIT FS-RT-30220

T7 High Yield RNA Synthesis Kit is a flexible kit for in vitro transcription of RNA using T7 RNA polymerase. Enables the substitution of NTPs for labeling and incorporation of modified bases.

The kit is suitable for synthesis of high yield RNA transcripts and for incorporation of modified nucleotides to obtain biotin labeled, dye labeled or capped RNA. The kit is also capable of synthesizing high specific activity radiolabeled RNA probes.

RNA synthesized from the kit is suitable for many applications including RNA structure and function studies, ribozyme biochemistry, probes for RNase protection assays and hybridization based blots, anti-sense RNA and RNAi experiments, microarray analysis, microinjection, and in vitro translation and RNA vaccines.

The T7 Enzyme Mix in the kit contains RNase inhibitors and inorganic pyrophosphatase, enabling a yield of at least **150 µg of RNA per reaction** from 1 µg of DNA template.

Kit Components (50 reactions)	Size
T7 RNA Enzyme Mix	100 µL
T7 Transcription Buffer (10x)	100 µL
ATP (100 mM)	100 µL
UTP (100 mM)	100 µL
GTP (100 mM)	100 µL
CTP (100 mM)	100 µL
Control Template (0.5 µg/µL)	10 µL
DNase I, RNase-free	100 µL

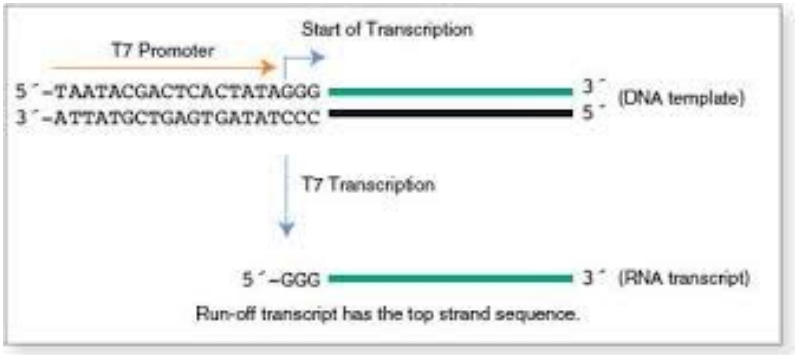
The kit contains sufficient reagents for 50 Reactions of 20 µl each

Note:

1.Prevent RNase contamination: When using this kit, wear a lab coat, disposable latex gloves, disposable masks, and use RNase-free consumables.

2.Template selection: It is recommended to purify the template before in vitro transcription to prevent contamination from RNase, protein, RNA, and salts.

3.Capped or modified RNA synthesis: If capped RNA is needed, prepare Cap Analog separately. If synthesizing labeled RNA (e.g., biotin, digoxigenin, FITC) or RNA with special modifications, the corresponding modified NTP needs to be prepared additionally.



1. Figure 1 illustrates the minimal T7 promoter sequence and the start of transcription as well as a run-off transcript after T7 transcription

NUCLEIC ACID STAINS, Nucleotides & DNA Ladders

Green Gel Safe Nucleic Acid Stain

Clearsight Nucleic Acid Stain

Green Gel Plus Nucleic Acid Stain

Sybr Safe Acid Stain

Eurosafe Green

Midori Green

Ethidium Bromide Destroyer

RNase Removal Spray Solution

dNTP SET (High Concentration)

dNTP SET MIX 10

dNTP SET MIX 20

1 Kb DNA Ladder (RTU)

100bp DNA Ladder (RTU)

50 bp DNA Ladder (RTU)

NUCLEIC ACID STAINS

Our Nucleic Acid Stains are ultra sensitive, extremely stable and environmentally safe fluorescent nucleic acid dye designed to replace the highly toxic ethidium bromide (EB) for staining dsDNA, ssDNA or RNA in agarose gels or polyacrylamide gels.
Our Nucleic Acid Stains are is far more sensitive than EB without requiring a destaining step.

Features:

- **Safer than EB**: Shown by the Ames test and other tests to be non-mutagenic and noncytotoxic
- **Easy disposal**: Passed environmental safety tests for direct disposal down the drain or in regular trash
- **Ultra-sensitive**: Much more sensitive than EtBr
- **Extremely stable**: Available in water, stable at room temperature for long-term storage and microwavable
- **Simple to use**: Very simple procedures for precast or post-electrophoresis gel staining
- **Compatible with a standard UV transilluminator**: Replaces EtBr with no optical setting change
- **Compatible with downstream applications**: Gel purification, restriction digest, sequencing and cloning

Cat.#	Description	Size
FS-02	**GREEN GEL SAFE** Nucleic Acid Stain **conc. 10,000X**	1 X 0,5 ML
FS-005	**CLEARSIGHT** Nucleic Acid Stain **conc. 20,000X**	1 X 1 ML
FS-GEL01	**GREEN GEL PLUS** Nucleic Acid Stain **High conc. 50,000X**	2 X 0,5 ML
FS-33102	**SYBR SAFE** Nucleic Acid Stain **conc. 10,000X**	1 X 400 µL
FS-GEL02	**EUROSAFE GREEN**	1 X 1 ML
FS-31	**MIDORI GREEN**	1 X 0,5 ML

ETHIDIUM BROMIDE DESTROYER

Fisher Molecular Biology Eth Br Destroyer is a specifically designed reagent effectively degrade and destroy the Ethidium Bromide and result in non-fluorescence and non-mutagenic remain. And also it has been demonstrated that its effectiveness of destructing the SYBR dyes. The FMB EtBr Destroyer Sprayer is for the treatment of solid Ethidium Bromide contaminant. The Sprayer can be used for the treatment of solid contaminant waste including electrophoresis gels, glassware, paper towels, gloves, laboratory equipment, bench surface etc

Features:

- **Effective**: EtBr destroyer can destroy EtBr and other SYBR Dyes. This effect can be monitored and confirmed by UV light exposure. Once destroyed, the fluorescence will disappear.
- **Safe**: The blocking of mutagenic effect of EtBr Destroyer has been demonstrated by Ames Test
- **Fast**: For general protection of uncontaminated area, spray the EtBr Destroyer on the entire working area, leave for about 5 minutes, then wipe it dry with paper towel.

Cat.#	Description	Size
EDB-30	Ethidium Bromide Destroyer Sprayer (2 X 200 ML) 1 sprayer contains 0.4 ml -1 sprayer can do 600 T.	2 Sprayers/box

RNase REMOVAL SPRAY SOLUTION

RNase Removal Spray Solution is ready to use and has more safely than traditional chemicals such as DEPC, a known carcinogen. RNase Removal Spray Solution can be used to remove RNase and DNA contamination from bench, instruments, experiment tools, glass and plastic consumables. Ready to use right out of the bottle, these solutions leave no residue on work surfaces when used as directed. It is good for lab-ware and surfaces that can't be autoclaved.

Features:

- **Easy Application**: Convenient spray bottle for direct application on surfaces.
- **Effective Decontamination**: Chemically inactivates and removes RNases (enzymes that degrade RNA).
- **Versatile**: Works on various surfaces like glass, plasticware, pipettors, and benchtops

Cat.#	Description	Size
FS-5371	RNase Removal Spray Solution	500 ml spray

dNTP Mix - NUCLEOTIDE SET (High concentration) 100 mM FS-013-1

100 mM dNTP Mix is a mixture of 4 deoxynucleotides (dATP, dCTP, dGTP, dTTP) in purified water. Each nucleotide is at a concentration of 100 mM.

100 mM dNTP mix is suitable for use in polymerase chain reaction (PCR), sequencing, fill-in reactions, nick translation, cDNA synthesis, and TdT-tailing reactions, qPCR , RT_qPCR.

Features:
• Chemically synthesized
• pH 7.5
• Free from qPCR, PCR, reverse transcription inhibitors
• Free of DNases and RNases
• Free of human and E. coli DNA

Size	Description	Size
FS-0131-1	**dNTP Mix - NUCLEOTIDE SET (High concentration)**	4 x 250 µl

dNTP Mix – Conc. 10 mM FS-013-2

dNTP Mix contains premixed aqueous solutions of dATP, dCTP, dGTP and dTTP, each at a final concentration of 10 mM. The nucleotides have greater than 99% purity, are free of nuclease activities, human and E. coli DNA. Mixes offer the possibility to reduce the number of pipetting steps and the risk of reaction set up errors. They are designed for many different molecular biology applications. Standard PCR, High-fidelity PCR, RT-PCR, Real Time PCR (qPCR)

Highlights

• Greater than 99% purity confirmed by HPLC
• Free of human and E. coli DNA
• Highly stable

Size	Description	Size
FS-0131-2	**dNTP Mix - 10mM**	500 µl

dNTP Mix – Conc. 20 mM FS-013-4

dNTP Mix contains premixed aqueous solutions of dATP, dCTP, dGTP and dTTP, each at a final concentration of 20 mM.

Applications:PCR, real-time PCR, high fidelity and long PCR, LAMP-PCR, cDNA synthesis, RT-PCR, RDA, MDA, DNA labeling, and DNA sequencing.

Size	Description	Size
FS-0131-4	**dNTP Mix- 20mM**	500 µl

READY-TO-USE DNA LADDERS 1 kb

1 kb DNA Ladder

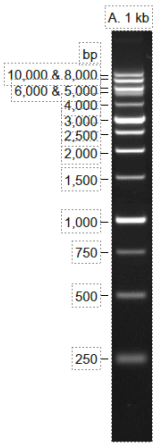
Product Description

The 1 KB DNA Ladder is suitable for sizing linear double-stranded DNA fragments from 250 bp to 10 kb. The 1 kb and 3 kb bands have increased intensity to provide internal orientation.
The ladders are generated from PCR and restriction enzyme digestion of double stranded DNA. The DNA is purified by phenol extraction, and equilibrated to 10 mM Tris-HCl (pH 8.0) and 1 mM EDTA. Approximate amounts of DNA per band per 100 ng ladder are listed in Figure 1 for reference, and are not intended for quantification of unknown DNA samples.

Protocol

For agarose gel electrophoresis the ladders can be loaded directly on a gel, 5-10 uL per well gives the optimal loading for a GelRed precast gel (100-200 ng/lane).

Storage: Store at 4°C for 6 months or at -20°C for 24 months.



ng/100 ng 1 kb Ladder

bp	ng
10,000	5.6
8,000	5.6
6,000	5.6
5,000	5.6
4,000	3.6
3,000	18.4
2,500	6.8
2,000	6.8
1,500	4
1,000	18.4
750	4.6
500	6
250	9

Figure 1. 100 ng of 1 kb DNA Ladder (A) or 100 bp DNA Ladder (B) were run on a 1% agarose/TBE/1X GelRed gel in 1X TBE at 100 volts for 90 minutes. Gels were imaged using a UVP GelDoc-It imaging system with ethidium bromide filter and 1 second exposure time. Fragment sizes in base pairs (bp) are shown next to each band. Approximate mass per band is shown for 100 ng DNA ladder in tables at right.

CAT.#	Description	Components	Size
FS-MW-500RT	**1 kb DNA Ladder**	1 KB DNA ladder in 1 X DNA Loading Buffer	500 ul

READY-TO-USE DNA LADDERS 100 bp

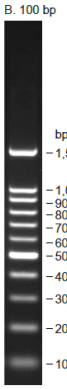
100 bp DNA Ladder

Product Description

100 bp DNA Ladder is suitable for sizing linear double-stranded DNA fragments from 100 bp to 1500 bp. The ladder consists of 11 bands that are generated from PCR and restriction enzyme digestion of double-stranded DNA. The DNA is purified by phenol extraction, and equilibrated to 10 mM Tris-HCl (pH 8.0) and 1 mM EDTA. The 500 bp and 1,500 bp bands have increased intensity to provide internal orientation.

Protocol

The Ready-to-Use DNA Ladders are supplied in a ready-to-load format at an optimal concentration for use on GelRed® precast gels. There is no need to mix with 6X loading buffer prior to loading onto a gel. For agarose gel electrophoresis, load 100-200 ng of DNA ladder (5-10 uL) per 5 mm lane.



ng/100 ng 100 bp Ladder

bp	ng
1,500	14.5
1,000	10
900	8
800	8
700	5.5
600	6
500	17
400	8
300	7
200	8
100	8

Storage: Store at 4°C for 6 months or at -20°C for 24 months.

Figure 1. 100 ng of 1 kb DNA Ladder (A) or 100 bp DNA Ladder (B) were run on a 1% agarose/TBE/1X GelRed gel in 1X TBE at 100 volts for 90 minutes. Gels were imaged using a UVP GelDoc-It imaging system with ethidium bromide filter and 1 second exposure time. Fragment sizes in base pairs (bp) are shown next to each band. Approximate mass per band is shown for 100 ng DNA ladder in tables at right.

CAT.#	Description	Components	Size
FS-MW-600RT	**100 bp DNA Ladder**	100 bp DNA ladder in 1 X DNA Loading Buffer	500 ul

READY-TO-USE DNA LADDER 50 bp

50 bp DNA Ladder

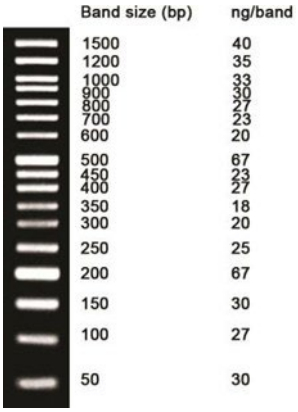
Fisher Molecular Biology Molecular 50 bp DNA Ladder is a molecular weight marker specially designed for easy quantification and size determination of small DNA fragments in agarose gels. Gels need to contain at least a 2% (w/v) concentration of agarose to allow the clear identification of each band. For best results using our ladder range we recommend using our agaroses

Size range: 50 bp to 1500 bp

Concentration:100ng/µL

Number of bands: 17

Size of bands: 50 bp, 100 bp, 150 bp, 200 bp, 250 bp, 300 bp, 350 bp, 400 bp, 450 bp, 500 bp, 600 bp, 700 bp, 800 bp, 900 bp, 1000 bp, 1200 bp,1500 bp

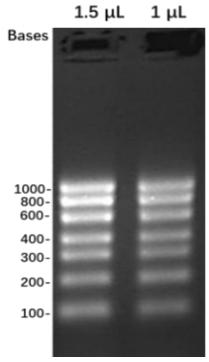


CAT.#	Description	Size
FS-MW-011	50 bp DNA Ladder RTU	500 ul

RNA MARKER 1000 FS-RMW-1000

Cat.n.	Product Name	Spec.
FS-RMW-1000	RNA Marker 1000	25 µL

Product Description/Introduction
RNA marker 1000 is a mixture of seven high-purified single-stranded RNA transcripts (in bases): 1000, 800, 600, 400, 300, 200 and 100 bases, obtaining by in vitro transcription.
The RNA concentration of the product is about 700 ng/µL suitable for routine gel electrophoresis of RNA.
Storage and Shipping Conditions
Ship with dry ice; store at -80℃ (short-term stored at -20℃), valid for 12 months.



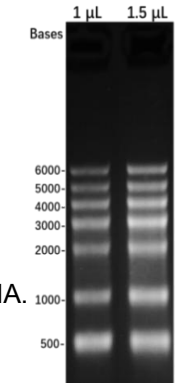
3 % Agrose Electrophoresis

RNA MARKER 6000 FS-RMW-6000

Product Information

Cat.n.	Product Name	Spec.
FS-RMW-6000	RNA Marker 6000	25 µL

Product Description/Introduction
RNA marker 6000 is a mixture of seven high-purified single-stranded RNA transcripts (in bases): 6000, 5000, 4000, 3000, 2000, 1000 and 500 bases, obtaining by in vitro transcription.
The RNA concentration of the product is about 700 ng/µL suitable for routine gel electrophoresis of RNA.



2% Agrose Electrophoresis

Molecular Biology Reagents & Buffers

Agarose D1-LE Standard

Agarose MS-8 Metaphor

Agarose (PFGE)

Agarose NuSieve 3:1

Agarose Low Melting

Dithiothreitol

EDTA

GLYCINE

GLYCEROL

HEPES

IPTG

MOPS

PROTEINASE K

Sodium Dodecyl Sulfate

SUCROSE

TEMED

TRIS Base Ultrapure

UREA Ultrapure

Acrylamide Solutions “Ready To Use”

30% Acry-BisAcrylamide ratio 29:1

30% Acry-BisAcrylamide ratio 37.5:1

Biological Buffers

TAE, TBE, TBS, PBS,Dulbecco’s PBS,

TE,PBS Tablets

AGAROSE D1-LE MOLECULAR BIOLOGY STANDARD AS-101

D-1 LE: with Low EEO.
High electrophoresis mobility ideal for DNA and RNA fragments as well as PCR products, for preparation of plasmids, for screening, cloning and blotting techniques.

- Nucleic acid analytical and preparative electrophoresis.
- Blotting
- Protein electrophoresis such as radial immunodiffusion.

Size: 500 gr

AGAROSE AS-101	
Moisture	4.62%
Ash	≤ 0.43%
EEO * (pH8.4)	0.12
Sulfate	≤ 0,097%
Clarity 1.5% (NTU)	3.89
Gel Strength 1% (g/cm2)	≤1.180
Gel Strength 1.5% (g/cm3)	≤2.920
Gelling temperature 1.5% (°C)	36.7
Melting Temperature 1.5% (°C)	88.2
DNase/RNase activity	None detected
DNA Resolution 1000 bp	Finely Resolved
Gel Background	Very low

AGAROSE MS-8 METAPHOR AS-109

An agarose for molecular screening that improves resolution of small DNA fragments and PCR products. Recommended for analytical gels for DNA ≤1,200 bp.

Functional Tests:

- DNA resolution: bands appear sharp and finely resolved.
- DNAse/RNAse activity: none detected.
- Gel background: very low after EtBr staining.
- DNA binding: very low

Size: 100 gr

AGAROSE METAPHOR	1,5%	3%
Moisture	=4.36%	
Ash	=0,26%	
EEO*	=0,11	
Sulfate	≤ 0,075%	
Clarity (NTU)	3,83	
Gel Strength (g/cm2)	1,965	3,810
Gelling Temperature (°C)		33,5
Melting Temperature (°C)		73,3

* EEO (electroendosmosis)

Ranges of separation:

1.8% 400-1200 bp	**3.0%** 150-800 bp	**4.5%** 15-400 bp

AGAROSE (PULSED FIELD ELECTROPHORESIS) AS-108

Agarose Pulsed Field Gel Electrophoresis is a linear polymer with a very high molecular weight, giving gel structures unlike those of traditional agaroses. This characteristic, added to the very low sulfate content, produces a strong intercatenary interaction, yielding a gel with very high gel strength and higher exclusion limit.

- Pulsed Field Gel Electrophoresis: because of its higher exclusion limit,larger molecules can be separated
  Separation Range: from ≥1 Kb up to 40 Kb.
- Blotting.
- Agarose Beads preparation.
- Cell and enzyme immobilization

Size: 100 gr

AGAROSE PULSED FIELD (Gel Electrophoresis)	
Moisture	≤ 5,12%
Ash	≤ 0.22 %
EEO*	≤ 0.11
Sulfate	≤0.083%
Clarity (NTU)	≤ 4
Gel Strength (g/cm2)	≥ 1,910
Gel Strength 1.5% (g/cm2)	≥ 3,900
Gelling Temperature (°C)	36,3
Melting Temperature (°C)	88,5
DNAse/RNAse activity	None Detected
DNA resolution	≥ 1 Kb- Up to 40 Kb
Gel background	Very Low

AGAROSE NUSIEVE 3:1 AS-110

NuSieve 3:1, a standard gelling/melting temperature agarose, is designed for analytical electrophoresis wherehigh resolving capacity is required. Recommended for DNA analytical gels at 2% concentrations, it can separate 30 -1,500 bp fragments The viscosity is low, so it is easy to make gels at high concentrations which have a very high resolving capacity.
Solutions of 4% or higher are feasible because of this low viscosity.
Because of the high gel strength, gels can also be prepared at lower concentrations, 1.0-1.5%. Gels are strong, flexible and very easy to handle.These features make NuSieve 3:1 gels compatible with blotting of small fragments.
Size: 100 gr

AGAROSE NuSieve 3:1	
Moisture	10%
Ash	0.4%
EEO*	0.13
Sulfate	0.15%
Clarity 4% (NTU)	4
Gel strength 4% (g/cm2)	1400
Gelling temperature 4% (°C)	32.5-38
Melting temperature 4% (°C)	90
DNase/RNase activity	None detected
Ranges of separation	2%:500 – 1500 bp 4%:150 – 600 bp

AGAROSE LOW MELTING AS-107

The low melting temperature allows for the recovery of undamaged nucleic acids below the denaturation
temperature. The low gelling temperature ensures that the agarose will be in a liquid state at a temperature range where In-Gel manipulations can be performed without prior extraction of the DNA from the gel slice.
Applications:
LM (Low Melting): with the highest gelling/melting temperatures and gel strength.

- Electrophoresis of DNA fragments ≥ 1000 bp
- Tissue and cell culture.
- Viral plaque assays

Size: 25 gr Size 50gr

AGAROSE LOW MELTING	
Moisture	≤ 7%
Ash	≤ 0.4%
EEO *	≤ 0.12
Sulfate	≤ 0,10%
Clarity 1.5% (NTU)	≤ 4
Gel Strength 1% (g/cm2)	≤ 250
Gelling temperature 1.5% (°C)	26
Melting Temperature 1.5% (°C)	≤ 65.5
DNase/RNase activity	None detected
Separation Range	≤1 bp
Inhibitors	none

DITHOTHREITOL (DTT) FS-0912

Formula : C4H10O2S2
Formula weight : 154.24 CAS #27565-41 -9
Product Specifications:
Form : White crystalline powder
Assay (S-H) : 99.5%
Melting point : 40 - 43°C A(280nm,=.1M, 1 cm) :≤0.06
A(260nm,=.1M, 1 cm) :≤0.40
Oxidized DTT : ≤0.2%
Storage : -20°C
Size: 10 g

EDTA FS-03620
Ethylenediaminetetraacetic Acid

Formula: C10H Assay: ≤ 98.0%
Water:<1.0%
Heavy Metals (as Pb):<0.001% Sizes: 100g , 500g, 1 Kg

GLYCINE FS-5037G

Assay by titration 99+% anhydrous Purity (by TLC) one spot Water (by Karl Fisher) ≤ 1.0%
pH (1 .0M)= 6.2± 0.3
A280 < 0.05 (1 .0M in H2O)
A260 < 0.05 (1 .0M in H2O)
IR: Conforms to known reference
Sizes: 1Kg - 5 Kg

GLYCEROL FS-7009

Formula: C3H803
MW: 92.09
Purity: 99.5+%
Cas# 56-81-5
DNase – Rnase – none detected

Glycerol does not freeze at -20°C Size: 500 Ml

HEPES FS-3071

Cas No 7365-45-9
(4-(Hydroxyethyl)piperazine-1-ethanesulphonic acid)
HEPES may be used as an alternative to PBS. It is the most generally used zwitterionic buffer which improves pH control between pH 6.7 and 8.4 and is obtained when 20-50 mM HEPES is incorporated into culture media. TBS and PBS may be used as washing buffers for alkaline and peroxidase conjugates in Western blotting as well as in various Cell Biology applications.
Sizes: 100g - 500g - 1Kg

IPTG FS-0481
Isopropyl-b-D (thiogalactopyranoside)

Presented as a white crystalline powder
Application: A gratuitous inducer of the E.coli lac + colonies or cells in a colorimetric assay
Size: 10gr

MOPS FS-2071

White powder, MW 209.3
Assay (by tritation); 99.5+%
Water (by Frank Fisher): ≤ 1,0%
Forms a clear, colorless solution in water (10%) ph(1%)=4.0
Size: 1 Kg

PROTEINASE K (Powder) FS-M-112

Cas No: 39450-01-6
Grade: High purity grade, for Molecular Biology
Purity: 99%
Specific activity: 35 units/mg of protein
DNase – none detected RNase – none detected Endonuclease (nickase) - none detected
Store: at -20°C
Size: 100 mg

SODIUM DODECYL SULFATE FS-0109
Ultrapure

Formula : C12H25Na04S
CAS # 151-21-3
Moisture : ≤1 %
Insolubles : ≤0.003%
Chloride (CI) : ≤0.01
Phosphate (PO4) : ≤1 ppm
Copper (Cu) : ≤5 ppm
Iron (Fe) : ≤1 ppm
Lead (Pb) : ≤5 ppm

Formula weight : 288.38
Form : White crystalline flakes
A(280nm,3%1cm):≤0.1
A(230nm,3%1cm):≤0.2
Assay : ≥99% (titrimetric)
Assay(C1 2) : ≥98% (GC)
Storage : Ambient
Sizes: 100g , 500g

SUCROSE FS-5393

Formula: C12H22O11 Formula weight: 342.30
CAS # 57-50-1
Product Specifications:
Form: White crystalline powder Identity: IR
Purify : ≥99.5% Storage : +20°C Sizes: 1 kg - 5 kg

TEMED FS-3009T
N,N,N',N' Tetramethylethyldiamine

Form: clear colorless liquid MW 116.2 Assay (by titration) 97+%

Forms a clear solution in water (20%)
pH(0.5%)=10.5±0.5
A400 ≤0.05(20% IN H2O)
IR: Conforms to known reference
Size : 100 ml

TRIS BASE ULTRAPURE FS-1503
equivalent to TRIZMA BASE (Sigma)

M.W. (Tris base /tris HCl) 121.1 / 157.6
Purity: >99.8%
Moisture: <1.0%
A280(1.0M,water) <0.05%
Insolubles: < 0.005%
Arsenic: <0.0005%
Copper: <0.0001%
Iron: <0.0001%

Magnesium:< 0.0001%
Heavy Metals:<0.0001%
DNase, RNase, protease: none detected
Storage : Rt
Sizes: 1 kg , 5 kg

UREA ULTRAPURE FS-0114

Formula : NH2-CO-NH2
Formula weight : 60.06
CAS #57-1 3-6
Form:White crystalline powder
Identity : By IR
Assay : ≥99.5%
Melting point : 132 - 135°C
Insolubles : Negligible
Turbidity : ≤2NTU
A(260nm,6M, 1 cm) : ≤0.055
A(280nm,6M, 1 cm) : ≤0.044

Copper (Cu) : ≤0.5 ppm
Iron (Fe) : ≤0.5 ppm
Lead (Pb) :≤0.5 ppm
Chloride (CI) :≤0.0005%
Cyanate : None detected
Conductivity : ≤1 5 µmho/cm
DNase (endo) : None detected
Rnase : None detected
Protease : Non detected
Storage : RT
Size: 1 kg/5 kg

ACRYLAMIDE SOLUTIONS – "READY TO USE"

Fisher Molecular Biology's liquid Acrylamide Solutions are made from highest quality pure material to exact standards.This ensure crystal clear electrophoresis gels which give realiable and reproducible results for separation of DNA and Protein Biomolecules.

CAT. N°	Description	Size	Applications
FS-2600	30% Acrylamide, Acrylamide to Bis-Acrylamide ratio 29:1	500 ml 1,000 ml	Separation of small acrylamide to bis-dsDNA fragments acrylamide (<1 kbps) + proteins
FS-2100	30% Acrylamide, Acrylamide to Bis-Acrylamide ratio 37.5:1	500 ml 1,000 ml	Preparation of protein gels

BIOLOGICAL BUFFERS

Fisher Molecular Biology provides a range of pre-filtered formulated buffer concentrates for a range of Molecular and Cell Biology application. Made from Ultrapure Reagents of molecular Biology Grade. Each lot is tested for DNase , RNase and protease Activity.

CAT. N°	Descritipion	Size
FSB-6002-10	TAE Buffer (10X)	1 L
FSB-6000-10	TBE Buffer (10X)	1 L
FSB-7301-10	TBS Buffer (10X)	1 L
FSB-74-10	PBS Buffer (1X)	500 ml
FSB-7415D	Dulbecco's PBS Buffer (1X)	500ml
FSB-6201	TE Buffer (1X)	1 L
FSB-2052-100	PBS Buffer Tablets (200 ml/each)	100 tablets

DNA Extraction & Purification

GEL Extraction & PCR Clean UP Kit

MicroElute GEL Extraction and PCR Clean UP Kit

GEL EXTRACTION / PCR CLEAN UP KIT DE-001

Description

The DE-001 Gel Extraction & PCR Clean Up Kit is designed to recover or concentrate DNA Fragment (50bp- 10Kb) from agarose gel, PCR or other enzymatic reaction. The unique dual purpose application and high yield DNA column make this kit exceptional value.

Features

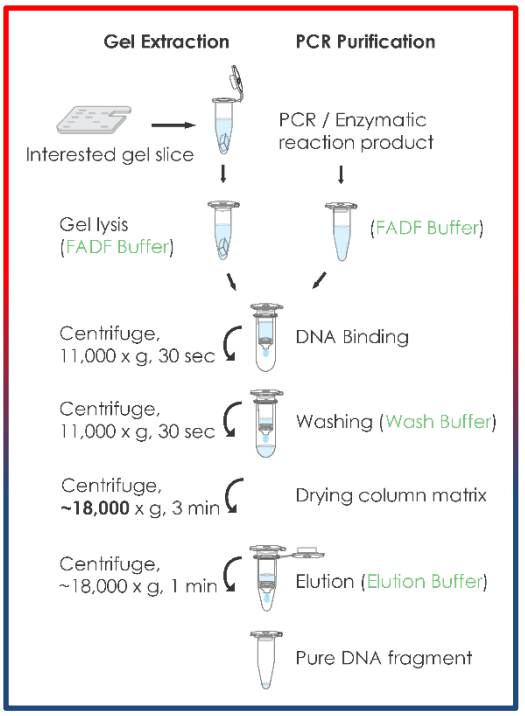
- With simple steps, quick and easy to use.
- Highly pure DNA (suitable for PCR).
- No phenol/chloroform extraction and ethanol
- precipitation required.

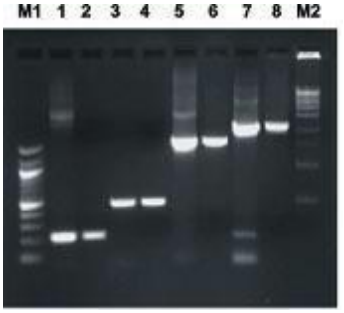
Applications

- PCR
- Fluorescent or Radioactive Sequencing
- Restriction Digestion
- DNA Labeling
- Ligation and Transformation

Specification:

Principle: spin column (silica matrix)
DNA Binding capacity of spin column: 20 µg
Sample size: up to 300 mg of agarose gel
up to 100 µl of reaction solution
Recovery: 70% ~ 85% for Gel extraction
90% ~ 95% for PCR clean-up
Operation time: 10 ~ 20 min
Elution volume: 40 µl





The Quality of DNA After Purification
DNA fragments before and after extraction with the GEL Extraction and PCR Clean Up Mini Kit
Lane 1, 3, 5, 7 before extraction: 200bp, 500bp, 2Kb, 3Kb.
Lane 2, 4, 6, 8 after extraction: 200bp, 500bp, 2Kb, 3Kb
M1: 1 00bp DNA Ladder
M2: 1 Kb DNA Ladder

Procedure: The method uses a chaotropic salt, guanidine thiocyanante to dissolve the agarose gel and denature enzymes. The DNA fragment in the chaotropic salt is bond to the glass fiber matrix of the spin column. After washing off the contaminants, the purified DNA fragments are eluted by a low salt elution buffer or water. Salts, enzymes and unincorporated nucleotides can be effectively removed from reaction mixture without phenol extraction and alcohol precipitation.

Cat. N.	Product Name	Size	Kit Components	Store at
DE-001	GEL Extraction & PCR Clean Up Kit	100 preps	FSDF Buffer Wash Buffer (Conc.) Elution Buffer FSDF columns 2 ml Collection tubes	Store at room temperature for 1 year.
DE-002		300 preps		

MICROELUTE GEL EXTRACTION / PCR CLEAN UP KIT DE-020

Description

The MicroElute Gel Extraction/PCR Clean UP Kit allows isolation and concentration of DNA fragments, 70bp~4Kb, from agarose gel, PCR reaction or enzymatic reactions. This kit eliminates impurities and salt efficiently from the sample matrix. The purified DNA fragments can be used diectly fro downstream applications and the end elution volume can be as low as 10 µl to obtain high concentration of DNA.

Specifications:

Principle: spin column (silica matrix)

DNA Binding capacity of spin column: 5 µg

Sample size:up to 200 mg of agarose gel

up to 100 µl of reaction solution

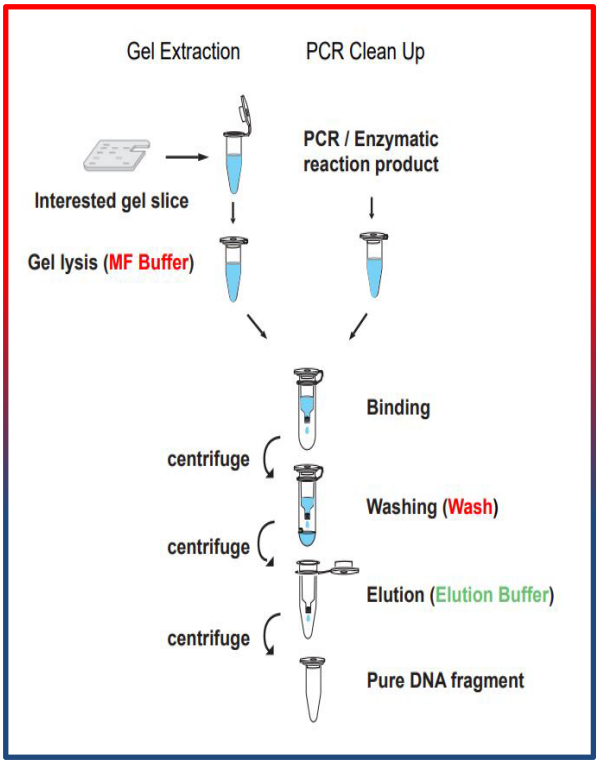
DNA size: 65 bp ~ 10 kbp

Recovery: 70% ~ 85% for Gel extraction

85% ~ 95% for PCR clean-up

Operation time: 10 ~ 20 min

Elution volume: 10 ~12 µl



Applications

Purified DNA is ready for downstream applicatons such as sequencing, ligation, labeling, amplication and enzymatic digestion.

Procedure

The DNA fragments in the chaotropic salt, are bonded to the glass-fiber matrix of the spin column. After washing off the contaminants, the purified DNA fragments are eluted by low-salt elution buffer or ddH2O. Salt, enzymes and unin-coporated nucleotides can be effectively removed from the reaction mixture without phenol/ chloroform extraction and alcohol precipitation.

Storage Conditions

Stable for 1 year at room temperature.

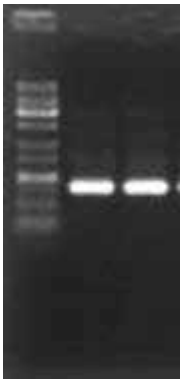


Fig.1: Agarose gel analysis of PCR product before and after purification with MicoElute Gel/PCR Purification.M: 1Kb DNA Ladder.
Lane 1: Before purification
Lane 2: after purification

Ordering Information

CAT.N°	Product Name	Size	Kit Components	Storage
DE-020	MicroElute Gel Extraction/ PCR Clean UP Kit	100 preps	GEL Lysis Buffer PCR Binding Buffer Wash Buffer conc. Elution Buffer FAPC-2 Columns 2 ml Collection Tube	Store at RT for 1 year

Blood & Tissue DNA Extraction & Purification

Blood & Tissue DNA Extraction kit

Tissue Genomic DNA Extraction Kit

Mouse Tail Direct PCR Kit (Genotyping)

Mouse Tissue Direct PCR Kit

Mouse Tails Direct PCR Lysis Buffer

BLOOD & TISSUE GENOMIC DNA EXCTRATION KIT DE-047 & DE-049

Description

The Blood & Tissue Genomic DNA Extraction Kit is designed for rapid extraction of pure, small-scale genomic & mitochondrial DNA from **Whole Blood (not frozen), dried blood spots, buffy coat and several types of tissues (fresh or frozen), fixed tissues (Formalin, Paraffin) , Bacteria , Yeast and Amniotic Fluid , sea urchins, marine mollusks and octopus.**

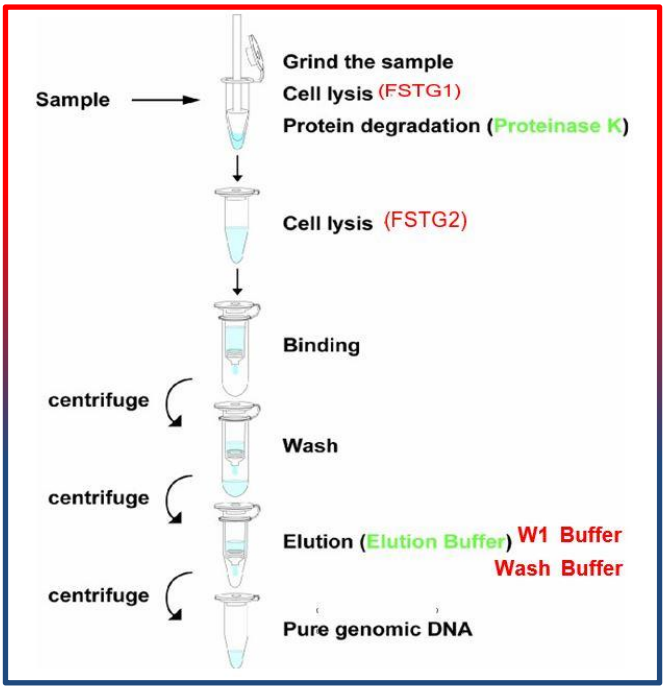
Features

-Rapid isolation cellular DNA from animal tissue (e.g. mouse tails), bacteria, yeast etc.
-Purified DNA is ready for downstream application such as PCR, Southern blotting.
-Centrifugation-based method.
-Efficiently remove cellular debair and inhibitors
-No phenol/chloroform extraction and ethanol precipitation.

Applications

- PCR
- Southern Blotting
- Forensic Analysis

Principle: spin column (silica membrane)
Operation time: 30 - 60 min
Binding capacity: up to 60 µg/ column
Minimum elution volume: 50 µl
Sample Sizes:
Up to 200 µl whole blood, serum, plasma, body fluids
< 25 mg animal tissue
1.2 cm mouse tail
< 10^7 cultured cells
DNA Yield: 4~8 µg/ 200 µl (whole blood)
DNA Yield: 15 ~35 µg/ prep (tissues)



Cat.n.	Description	Size	Kit Contents	Storage
DE-047	Blood & Tissue DNA Extraction	100 preps	Proteinase K (powder) FSTG1 Buffer FSTG2 Buffer W1 Buffer Wash Buffer (concentrated) FSTG Columns 2 mL Collection Tubes 1.5 Elution tubes Micropestles	Store at room temperature. Except Proteinase K, store at +4°C.
DE-049-200		200 preps		
DE-049-400		400 preps		

TISSUE GENOMIC DNA EXTRACTION KIT DE-012 -DE-013

Description

The Tissue Genomic DNA Extraction Kit is designed for rapid extraction of pure, small-scale genomic & mitochondrial DNA from several types of tissues: **fixed tissues (Fresh , Frozen ,Formalin, Paraffin) whole Blood , buffy coat , bacteria, yeast, fungi and ,saliva dry blood spots, viral, hair, bone tissue, dental tissue, insects, amniotic fluid, sea urchins, marine mollusks, octopus and insects.**

Sampling

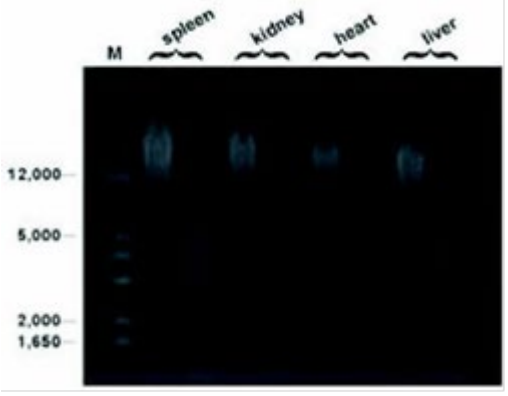
- **Principle:** mini spin column (silica matrix)
- **Operation time:** 30 ~ 60 minutes
- **Binding capacity:** up to 60 ug DNA/column
- **Typical yield:** 15 ~35 ug/ prep
- **Column applicability:** centrifugation and vacuum
- **Minimum elution volume:** 50 ul
- **Sample size**: < 25 mg animal tissue
  - 1.2 cm mouse tail
  - < 10^7 cultured cells

Rapid isolation cellular DNA from animal tissue (e.g. mouse tails), bacteria, yeast etc.
Purified DNA is ready for downstream application such as PCR, Southern blotting.
Centrifugation-based method.
Efficiently remove cellular inhibitors
No phenol/chloroform extraction and ethanol precipitation.

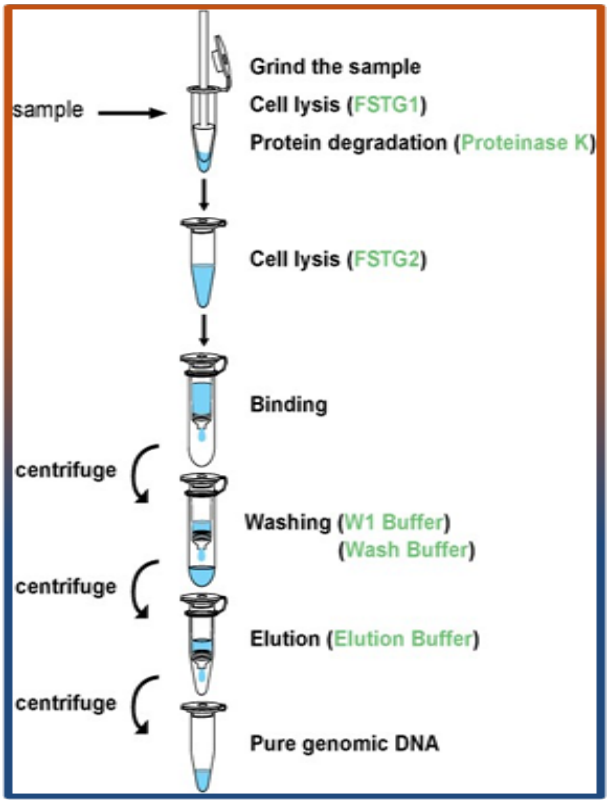
Applications

PCR
Southern Blotting
Forensic Analysis

The Quality of DNA after Purification



Genomic DNA Extracted from the indicated mouse tissue by the Tissue Genomic DNA Extraction Kit
For each tissue, the amount of undigested (left) and EcoRI digested (right) are equivalent M1: 1Kb DNA Ladder (100bp-12,000bp)



Cat. N.	Product Name	Size	Store at
DE-012	Tissue genomic DNA Extraction Mini Kit	100 preps	Store at room temperature for 1 year. (Except Proteinase K:store at -20°C)
DE-013		200 preps	

MOUSE DIRECT PCR KIT

DE-070 - DE-071

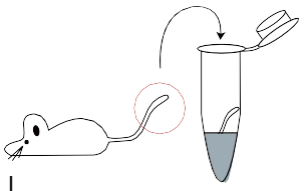
Description:

The Mouse Direct PCR Kit provides a fast preparation and PCR amplification that is specifically designed for mouse genotyping. The Buffer L and Protease Plus rapidly digest mouse **tail**, **ear** and **toe** to release intact genomic DNA that can be used directly as the template for PCR amplification. By using this kit, the digestion process only takes **15 min**. In addition, the 2x PCR Master Mix (which includes an optimized Taq Polymerase) ensures high amplification efficiency of target DNA.

Storage

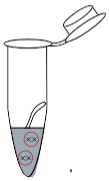
Buffer L should be stored at 4°C. Other reagents should be stored at -20°C. All reagents can be stored for 2 years.

Protocol



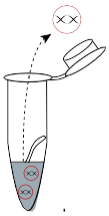
Add protease mixture to the tissue tub

Tissue Digestion



Incubate at 55°C for 15 min to release genomic DNA

Protease Inactivation



After digestion, incubate at 95°C for 5 min to inactivate the proteases

PCR

After centrifuge, 12,000 rpm for 10 min, the supernatant of digested solution can be used as the template for PCR

Components

Contents	DE-070 (200 rxns)	DE-071 (500 rxns)
Buffer L	20 mL	50 mL
Protease Plus	0.4 mL	1 mL
2 x PCR Master Mix[a]	2 mL	5 mL

a. 2x PCR Master Mix includes more powerful DNA polymerase, dNTPs, Mg^{2+}, and DNA Loading Dye.

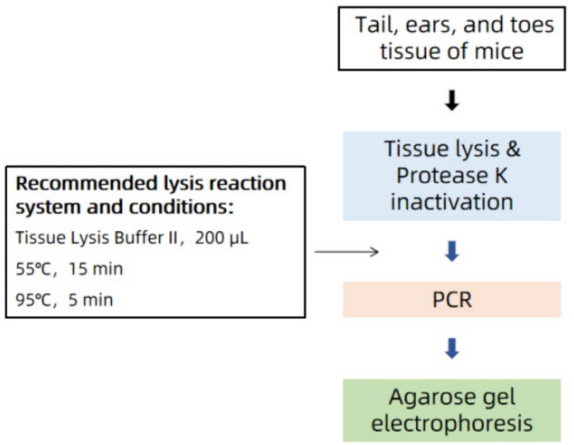
MOUSE TISSUE DIRECT PCR KIT DE-736
(Genotyping)

The Mouse Tissue Direct PCR Kit is specially designed for the rapid genotyping of mouse, which contains Tissue Lysis Buffer II and HS 2X PCR Mix for Mouse Genotyping for DNA release and PCR amplification. This kit can be used for the rapid extraction of genomic DNA from mouse tails, ears, toes, and other tissues. The extracted genomic DNA can be used directly as template for PCR amplification, which greatly shortens the experimental time. Tissue Lysis Buffer II in this kit already contains proteinase K, which does not require additional preparation.
Hot Start 2X PCR Mix for Mouse Genotyping in this kit only needs to add primers and templates to perform amplification, and the reagent contains loading buffer, PCR products can be directly loaded for electrophoresis.

Tissue Lysis Buffer II in this kit already contains proteinase K, which does not require additional preparation, and Tissue Lysis Buffer II has been tested to be stable storage for 4 weeks at room temperature or 4°C, thus avoiding repeated freeze-thaw cycles.
HS 2X PCR Mix for Mouse Genotyping in this kit contains high-performance DNA Polymerase, dNTP,and an optimized buffer system. It only needs to add primers and templates to perform amplification, thereby reducing pipetting operations, significantly controlling cross-contamination among samples, and improving detection throughput and reproducibility of results.The amplification system contains protective agents that keep HS 2X PCR Mix for Mouse Genotyping stable in activity after repeated freezing and thawing. HS 2X PCR Mix for Mouse Genotyping contains loading buffer, PCR products can be directly loaded for electrophoresis after the reaction.

PCR Product End: blunt end

Experiment workflow



Operation Description Standard Protocol

1. Take a clean 1.5 mL EP tube and add mouse tail to the EP tube (1-2 mm or 3-5 mg is recommended for mouse tail tip and toes, and 1-5 mm 2 or 3-5 mg for mouse ears).

2. Add 200 μL of Tissue Lysis Buffer II to the EP tube to completely submerge the tissue. Note: Tissue Lysis Buffer II must be fully dissolved till becoming clear before use.

The recommended lysis reaction conditions are as follows:

Step	Temp	Time
Digestion	55℃	15 min
Inactivation	95℃	5 min

Note:
To ensure the efficiency of DNA release, be sure to immerse all tissues in the lysis buffer. After the incubation, the tissue block may not be completely digested, which is normal and does not affect the use.

Components	DE-736-100	DE-736-500
Hot Start 2xPCR Mix For Mouse Genotyping	100 RXN (25 μL/Rxn)	500 RXN (25 μl/Rxn)
Tissue Lysis Buffer II	20 ml	100 ml

***Note**：Tissue Lysis Buffer II contains proteinase K, which can be stored at 4°C/Room temperature for less than 4 weeks, -20°C is recommended if stored for more than 4 weeks.

MOUSE TAILS DIRECT PCR LYSIS BUFFER FLB-1001T

Direct PCR Lysis Buffer was especially developed for the lysis of mouse tail tissue, and other tissues.
After a brief heat treatment, the crude lysates are directly used for PCR without time-consuming genomic DNA isolation.

Using Fisher Molecular Biology Mouse Tails-Direct PCR Lysis Buffer, DNA extracts can be easily obtained directly from

- Mouse Tails
- Mouse Ears
- Yolk Sac
- Culture cells

No purification of DNA is required

The DNA extracts will be suitable for one-step PCR genotyping and PCR amplifications.

Fisher Molecular Biology Direct PCR Lysis Buffer are single-tube systems for rapid, convenient, and reliable preparation of DNA from mouse tails, ears, yolk sacs, and culture cells.
The innovative system developed by **Fisher Molecular Biology** allows the resulting DNA crude extracts to be ready for genomic PCR for genotyping in less time and less hands-on involvement.
Crude extracts of biological samples are not compatible with many molecular biology-grade reactions such as polymerase chain reaction (PCR), in part due to inhibitors contained in crude extracts.

The **Direct PCR Lysis Buffer** not only mediate the ***rapid lysis of biological samples*** but also contain inhibitors that effectively suppress the inhibitory activities of crude lysates for PCR amplification, while maximally ***maintaining the integrity of released genomic DNA***. Our lysis reagents completely eliminate any solution transfer or tube-opening steps, providing you with substantial extra time and less risk.

Brief procedure:

1. **Lyse tails in Direct PCR Lysis Buffer**
2. **Incubate for 45 min at 85°C.**
3. **PCR genotyping with 1 µl lysates.**

Detailed protocols: Tails, Ears, Yolk Sac, and Cultured cells.

The Direct PCR Lysis system offers advantages and savings over conventional protocols that include:
- **Time:** Virtually no hands-on time. Crude tail lysates for PCR.
- **Safety:** No organic reagents.
- **Environmental:** Less waste (organic reagents, tubes, tips, etc...)
- **Reliability:** Virtually 100% success rate with high yields.

Direct PCR Lysis Reagents

Cat #	Description
FLB-1001T	Direct Lysis Buffer for mouse tails (100 ml) (500 tails)
FLB-1002E	Direct Lysis Buffer for mouse ears (100 ml) (1000 ears)
FLB-1003Y	Direct Lysis Buffer for Yolk sac (100 ml)
FLB-1004C	Direct Lysis Buffer for Cultured cell (100 ml)

Plasmid DNA Extraction and Purification

Plasmid DNA Extraction Mini Prep Kit

Plasmid DNA Extraction Extraction Midi Prep Kit (Endotoxin Free)

Plasmid DNA Extraction Extraction Maxi Filter Prep Kit (Endotoxin Free)

PLASMID DNA EXTRACTION MINI PREP KIT DE-034

The Plasmid DNA Extraction Kit provides a rapid, phenol-free method for the extraction of high-purity plasmid DNA from bacterial cultures such as E. coli , which bacteria is pellet, lysed, and then neutralized. The extracted DNA can be used in a variety of applications such as PCR, cloning, sequencing, in vitro transcription, and labeling. Also, as a column-type tube is utilized in the purification process, extraction is carried out in three simple steps of binding / wa- shing / elution. Once bound, the DNA is washed and then eluted from the column, ready for use.

Features

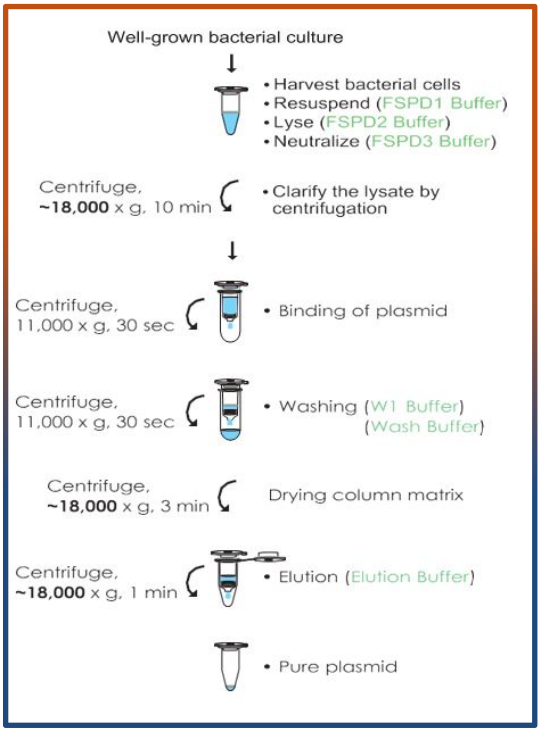
- For high yields of plasmid DNA-up to 30µg from 1~5ml overnight cultures.
- Effective purification of DNA fragments ranging from 100bp to <15kb.
- No need for messy resin slurries, extracting with phenol, or concentrating via alcohol precipitation. Superior
- purity-DNA yields quality sequence data using automated or manual methods.
- Optimized buffers are included for maximum DNA purity and yield.
- Versatile protocol-works with all neutral gel buffers and both conventional and low-melting agarose gel.

Format

Spin Columns

Specifications

Principle: mini spin column (silica matrix)
Sample size: 1 ~ 5 ml
Size of plasmid or construct:< 15 kb
Operation time: < 25 minutesTypical
Yield: 20 ~ 30 µg of high copy plasmid 3
~ 10 µg of low copy plasmid
Binding capacity: 60µg/column
Column applicability: centrifugation and vaccum



Applications

Fluorescent or radioactive Sequencing

- Ligation
- Restriction enzyme digestion
- Ligation and Transformation
- Library screening

Cat. N.	Product Name	Size	Store at
DE-034	Plasmid DNA Extraction Mini Prep	100 preps.	Store at RT for 1 year Store FSPD1 Buffer with RNase A included at +4°C. Store the RNAse A vial at -20°C for 1 year.
DE-035	Plasmid DNA Extraction Mini Prep	300 preps.	

PLASMID DNA EXTRACTION MIDI PREP KIT DE-051EF

(ENDOTOXIN FREE)

The Plasmid DNA Extraction Midi Kit “Endotoxin Free” is designed for efficient extraction of high quality plasmid DNA from bacterial culture. This kit provide the alkaline lysis reagents and the columns packed with anion-exchanger resin. After the cells lysis , the plasmid DNA is bound to the resin inside the column by a gravity-flow procedure, and the contaminants can be remove with wash buffer. After using this convenient kit, the purified plasmid DNA is suitable for downstream application such as: transfection, in vitro transcription and translation, and all enzymatic modification.

Specifications: bacterial culture
Sample Size: up to 60 ml of high-copy number plasmid
up to 120 ml of low-copy number plasmid

Plasmid or costruct Range: 3kbp-150 kbp
Binding Capacity: 650 µg / Midi column
Features:

- **Time saving:** Complete the process in less than 2 hours.

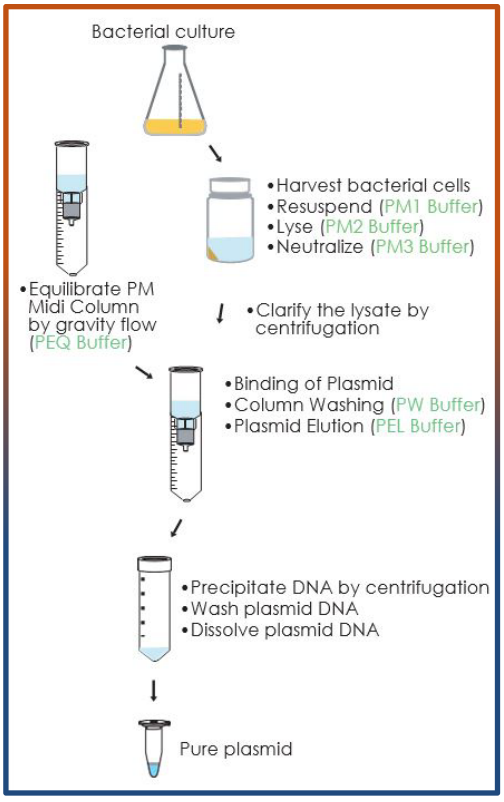
Applications:
The purified plasmid DNA can be immediately used in any downstream molecular biology application.

- **Purity:** equity to that obtained by 2x CsCl-gradient centrifugation .

Safe: Eliminates the use of phenol, chloroform, ethidium bromide.

- Transfection
- Microinjection
- Sequencing
- PCR
- Restriction enzyme digestion

Operation Time :Less than 2 hours
Operation Format: Gravity-Flow



Ordering Information

Cat. N°	Product Name	Size	Store at:
DE-051EF	Plasmid Extraction Midi Prep Kit Endotoxin Free	25 preps	At Room Temperature RNase A at -20°C

PLASMID DNA EXTRACTION MAXI FILTER KIT DE-055-EF
(ENDOTOXIN FREE)

The Plasmid DNA Extraction Maxi Filter "Endotoxin Free" Kit is designed for rapid and efficient extraction of high quality endotoxin-free purified plasmid DNA with anion-exchange technology. With provided filter cartridges the bacteria lysates will be removed without centrifugation. Following a gravity-flow procedure, the plasmid DNA is bound to the resin, and the contaminants can be remove with wash buffer.PTR Buffer washes away the endotoxins in just one step. This kit is designed for the convenient, easy, and efficient extraction of pure plasmid DNA and makes the endotoxins less than 0.05 EU/µg DNA that is suitable for the transfection of cultured cells.

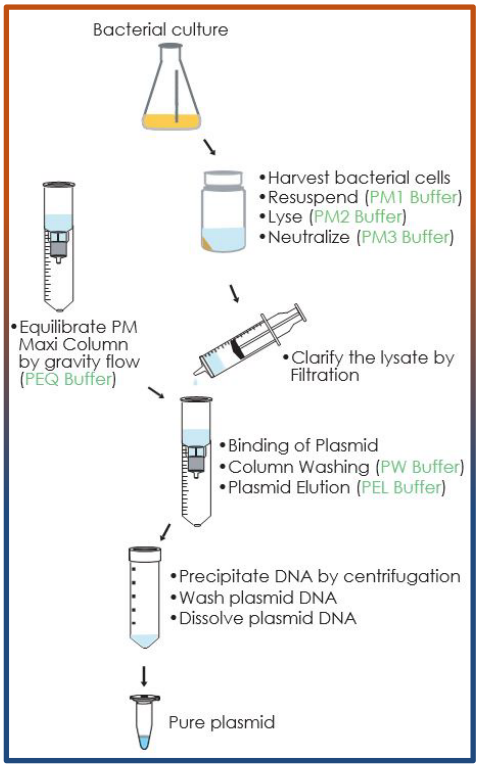
After using this convenient kit, the purified plasmid DNA is suitable for downstream application such as transfection, in vitro transcription and translation, and all enzymatic modification.

Specifications: bacterial culture

Sample Size:

- 120- 240 ml of bacterial for high copy plasmid or low copy plasmid
- Plasmid constructs' range: 3kbp-150 kbp

Binding Capacity: up to 1.5 mg /Maxi Column



Additional Requirements:

1. 50 ml tubes
2. Refrigerated centrifuge capable of ≥5,000 xg and the centrifuge tube suitable for the centrifuge rotor
3. Isopropanol
4. 70% ethanol
5. TE buffer or ddH2O

Features

Time saving: 1,5 hour

DNA Yield : up to 1,500µg/column

- High Purity: Equal to that obtained by 2x CsCl gradient centrifugation.
- Safe: Eliminates the use of phenol, chloroform, ethidium bromide, and cesium chloride, minimizing exposure to and disposal of hazardous materials.

Applications

The purified plasmid DNA can be used immediately in downstream application.

- Transfection (non-endotoxin sensitive)
- Microinjection
- In Vitro transcription
- Restriction Enzyme digestion

Procedure

In the process, after the modified alkaline lysis, the filter cartridge is used to remove bacteria lysates to obtain cleared sample matrix. Then the plasmid DNA will bind to the onion exchange resin inside the Maxi Column when the sample matrix flowing through. The contaminants can be removed by wash buffer. Finally, the purified plasmid DNA is luted using high-salt buffer and then precipitated with iospropanol for desalting.

Ordering Information

Cat. N°	Product Name	Size	Store at:
DE-055EF	Plasmid Extraction Maxi Filter Endotoxin Free Kit	10 preps	At Room Temperature RNase A: at-20°C

RNA Extraction & Purification

Tissue Total RNA Etraction Mini Kit

TRIZOL Ultrapure

RNA-ZOL Direct Clean Up-Plus Kit

RNA Stabilization Reagent

Viral Nucleic acid (DNA& RNA) Extraction Kit

TISSUE & CELLS TOTAL RNA EXTRACTION MINI-KIT RE-006

The Tissue & Cells Total RNA Purification Mini Kit is designed for purification of total RNA from: Animal Tissue (fresh, frozen, paraffin) colture Cells, Bacteria,Yeast, Fungi, it makes RNA clean-up, using the chaotropic salt- lysis method without the use of hazardous solvents such as phenol. The Kit can quickly purify total RNA from up to 10mg of tissues within 30 minutes.
The purified RNA is suitable for direct use in RT-PCR,Northern blotting, primer extension and cDNA library construction.

Features:
Operation time: 30 ~ 60 minutes
Binding capacity: up to 100 µg total RNA/ column
Column applicability: centrifugation and vaccum
Minimum elution volume: 40 µl

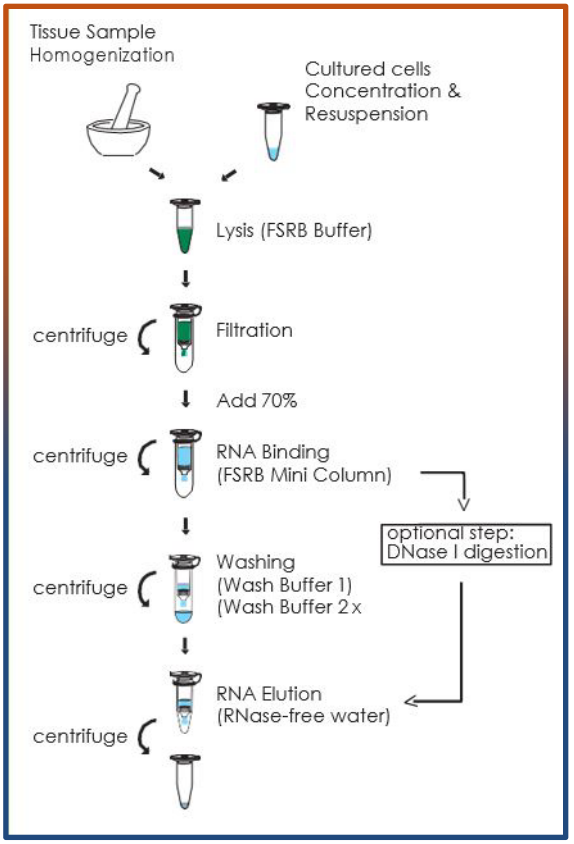
Applications
- Nothern blotting hybridizations
- Primer extension
- RT-PCR
- RNase protection assays
- Differential display
- As starting material for purification of mRNA for cDNA synthesis

Sample Size: Animal cells: from 1 x 10^6 up to 5 x 10^6 cells
Animal Tissues: (Mouse/Rat) from 10 mg up to 30 mg
Bacteria: 1ml or up to 1x10^9 cells
Yeast: (up to 5 x 10^7)

Yield of Purification

Sample	Recommended amount of sample used		Yield (ug)
Animal Cells (up to 5 x 10^6 Cells)	NIH/3T3 HeLa COS-7 LMH	1 X 10^6 Cells	10 15 30 12
Animal Tissue (mouse/rat) (Up to 30 mg)	Embryo Heart Brain Kidney Liver Spleen Lung Thymus	10 mg	25 10 10 30 50 35 15 45
Bacteria	E.coli B. subtilis	1 x 10^9 Cells	60 40
Yeast (up to 5 x10^7 cells)	S. cerevisiae	1 x 10^7 Cells	25

Brief procedure:



Procedure
The method uses detergents and a chaotropic salt to lysis cell and inactivate RNase, then RNA in chaotropic salt is bonded to the glass fiber matrix of column. After washing off the contaminants, the purified RNA is eluted by RNase-free water. The entire procedure can be completed about 30~60 minutes.

Storage Conditions
Tissue Total RNA Mini Kit can be stored at room temperature (15-25°C). Stable for 1 year at room temperature at 15-25°C.

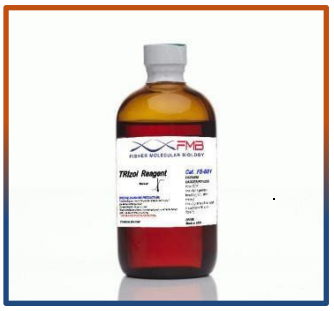
Cat. No.	Product Name	Size	Store at
RE-006	Tissue & Cells Total RNA Mini Kit	100 preps	At Room Temperature at 15-25°C for 1 year

TRIZOL ULTRAPURE-RNA / DNA / PROTEIN ISOLATION REAGENT FS-881

TRIzol Reagent is a ready-to-use reagent, designed to isolate high quality total RNA (as well as DNA and proteins) from cell and tissue samples of human, animal, plant, yeast, or bacterial origin, within one hour. TRIzol Reagent is a monophasic solution of phenol, guanidine isothiocyanate, and other proprietary components which facilitate the isolation of a variety of RNA species of large or mall molecular size. TRIzol Reagent maintains the integrity of the RNA due to highly effective inhibition of RNase activity while disrupting cells and dissolving cell components during sample homogenization. TRIzol Reagent allows for simultaneous processing of a large number of samples, and is an improvement to the single-step RNA isolation method developed by Chomcynski and Sacchi (Chomczynski & Sacchi, 1987).

- Isolated DNA can be used in PCR, Restriction Enzyme digestion, and Southern Blots.
- Isolated protein can be used for Western Blots, recovery of some enzymatic activity, and some immunoprecipitation.
- Ready-to-use reagent for the isolation of total RNA or the simultaneous isolation of RNA, DNA and proteins from samples of human, animal, plant, yeast, bacterial and viral origin.
- Single-step method of total RNA isolation
- Performs well with small and large quantities of tissues or cultured cells and allows simultaneous processing of a large number of samples.
- Combines phenol and guanidine thiocyanate in a mono-phase solution to facilitate the immediate and most effective inhibition of RNase activity.
- RNA remains exclusively in the aqueous phase, DNA in the interphase and proteins in the organic phase.

Cat. N.	Product Name	Size
FS-881	TRIZOL DNA/RNA Protein Isolation Reagent	100 ml
		200 ml



RNA-ZOL DIRECT CLEAN-UP PLUS KIT RE-040

The RNA-Zol Direct Clean Up Plus kit provides a streamlined method for the purification of up to 100 µg (per column) of high-quality RNA directly from samples in TRIzol®, TRI Reagent® or similar
Total RNA including small RNAs (17-200 nt) is isolated from a variety of sample sources (cells, tissues, serum, plasma, blood, etc.). Simply add ethanol to a TRI Reagent® sample, bind directly to the Column, wash, and elute RNA. No phase separation, precipitation, or post-purification steps are necessary. RNA is high-quality and ready for Next-Gen Sequencing, RT-qPCR, transcription profiling, hybridization, etc.

Features

RNA clean up plus can be operated directly after the chloroform extraction without isopropanol precipitation.

- Sample Size: Up to 100 µl of RNA sample or enzymatic reaction mixture.
- High purity: OD260/280: 1.9~21.
- Binding Capacity: Up to 100 ug
- Handling Time: Within 10 minutes
- Expected Recovery: 85~95%
- Format: Spin Column

Applications

- Real-Time PCR
- Northern blotting hybridization
- Primer extension
- Differential display
- RNase protection assays
- As starting material for the purification of mRNA for cDNA synthesis

Storage Conditions

Stable for 1 year at room temperature

Cat. N.	Product Name	Size	Store at
RE-040	RNA-Zol Direct Clean-Up Plus Kit (Tri-Zol included)	50 preps	Store at 15°C for 1 years
RE-041		200 preps	
RE-042	RNA-Zol Direct Clean-Up Plus Kit (Tri-Zol not included)	50 preps	
RE-043		200 preps	

RNA LATER STABILIZATION REAGENT FS-883

RNA*later* Stabilization Reagent immediately **stabilizes RNA in tissues, cell cultures and blood samples** to preserve the gene expression profile.

RNA Later makes it possible for researchers to postpone RNA isolation for days, weeks, or even months after tissue collection without sacrificing RNA integrity.

The reagent preserves RNA for up to 1 day at 37°C, 7 days at 18–25°C, or 4 weeks at 2–8°C, allowing processing, transportation, storage, and shipping of samples without liquid nitrogen or dry ice.
Alternatively, the samples can also be placed at –20°C or –80°C for archival storage.

Advantages:

In addition for RNA stabilization, RNA Solution can be easily integrated into a modified single-step RNA isolation method. This modified single-step method isolates undegraded RNA from tissues or cells in hours and can be used to process a large number of samples.

Protocol for Tissues

1. (Solution up to 100 mg tissue add 1 ml RNA Later)
Store the tube at –20°C until use.
2.When processing thaw and homogenize tissues in RNA Later .
3.Transfer 0.8 ml of the homogenate mix into a 2 ml tube and add 0.8 ml of the acid-phenol, pH 5.2, and 320 ul of chloroform.
4.Vortex the mixture vigorously by mixing 4 times, 30 sec for each.
5.Centrifuge at 12,000 rpm for 2 min
6. Transfer the upper aqueous phase (containing RNA) to a fresh 2 ml tube, taking care not to disturb the interface (containing DNA/protein).
7. Precipitate the RNA by adding an equal volume (0.8 ml) of isopropanol and 80 ul of 3 M NaAc at –20°C for 30 min.
8. Centrifuge at 12,000 rpm for 15 min and discard the supernatant.
9. Wash the RNA pellet by using 200 ul of 70% ethanol and gentle inverting the tube for several times.
10. After a brief spin and careful removing the supernatant, let the RNA pellet to air dry for about 5-10 min.
11. Dissolve the RNA pellet in 20 ul DEPC-treated TE.
12. Store the samples at –20°C and used for cDNA synthesis.

Protocol for Culture Cells

1. Transfer 107 cells (isolated from cell colture) into 1 ml of RNA Later Solution
Store the tube at –20°C until use.
2. When processing thaw and homogenize tissues in RNA Later
3. Transfer 0.8 ml of the homogenate mix into a 2 ml tube and add 0.8 ml of the acid-phenol, pH 5.2, and 320 ul of chloroform.
4. Vortex the mixture vigorously by mixing 4 times, 30 sec for each.
5. Centrifuge at 12,000 rpm for 2 min.
6. Transfer the upper aqueous phase (containing RNA) to a fresh 2 ml tube, taking care not to disturb the interface (containing DNA/protein).
7. Precipitate the RNA by adding an equal volume (0.8 ml) of isopropanol and 80 ul of 3 M NaAc at –20°C for 30 min
8.Centrifuge at 12,000 rpm for 15 min and discard the supernatant.
9. Wash the RNA pellet by using 200 ul of 70% ethanol and gentle inverting the tube for several times.
10. After a brief spin and careful removing the supernatant, let the RNA pellet to air dry for about 5-10 min.
11. Dissolve the RNA pellet in 20 ul DEPC-treated TE.
12.Store the samples at –20℃ and used for cDNA synthesis.

Protocol for Whole Blood

1. Collect fresh human blood in an anticoagulant-treat collection tube.
2. Transfer up to 300 µl fresh blood to a 1.5ml microcentrifuge tube (not provided). If the sample is more than 300 µl (up to 1ml), add the sample to a sterile 15 ml centrifuge tube.
3. Add 3 x the sample volume of RBC Lysis Buffer and mix by inversion. Do not vortex.
4. Incubate at room temperature for 10 minutes. Centrifuge at 3,000 x g for 5 minutes and completely remove the supernatant.
5.Resuspend the pellet with 100 µl of RBC Lysis Buffer.
6. Store 100µl of RBC Lysis Buffer with 1 ml of RNA Stabilization Solution at –20°C until RNA isolation.
7. When processing, thaw and homogenize tissue in RNA Stabilization Solution.
8.Transfer 0.8 ml of the homogenate mix into a 2 ml tube and add 0.8 ml of the acid-phenol, pH 5.2, and 320 ul of chloroform.
9.Vortex the mixture vigorously by mixing 4 times, 30 sec for each.
10. Centrifuge at 12,000 rpm for 2 min.
11. Transfer the upper aqueous phase (containing RNA) to a fresh 2 ml tube, taking care not to disturb the interface (containing DNA/protein).
12. Precipitate the RNA by adding an equal volume (0.8 ml) of isopropanol and 80 ul of 3 M NaAc at –20°C for 30 min
13. Centrifuge at 12,000 rpm for 15 min and discard the supernatant.
14. Wash the RNA pellet by using 200 ul of 70% ethanol and gentle inverting the tube for several times.
15. After a brief spin and careful removing the supernatant, let the RNA pellet to air dry for about 5-10 min.
16. Dissolve the RNA pellet in 20 ul DEPC-treated TE.
17. Store the samples at –20℃ and used for cDNA synthesis.

Cat. No.	Product Name	Samples	Size	Store at
FS-883	RNA Later Stabilization Reagent	Tissues Cell Cultures	100 ml	Store at +4°C

VIRAL NUCLEIC ACID (DNA & RNA) EXTRACTION KIT I DR-001

Viral Nucleic Acid Extraction Mini Kit I is designed for extraction of Viral DNA or RNA from cell free fluides such as serum, plasma, body fluid and cell cultured supernatant and from transport medium of swabs (covid samples). This method first lyses virus by using a chaotropic salt, then binds nucleic acid to silica-based membranes. After washing with ethanol-contained wash buffer, contaminants and enzyme inhibitors will be removed completely. It takes only 20 min for an entire procedure, the purified nucleic acid is ready for RT-PCR and PCR **.** gel, up to 200 mg. This kit contains carrier RNA for very low viral load samples.

Features:

- Principle: spin column (silica membrane)
- Safe Use: Eliminates the use of phenol, chloroform, ethidium bromide, and cesium chloride, minimizing exposure to, and disposal of hazardous materials.
- High Purity: Complete removal of contaminants and inhibitors for reliable downstream applications

Sample: 140 µl cell-free fluid such of plasma, serum, body fluids , cell cultured supernatant and from transport medium of swabs (covid samples.)

Length of recovery nucleic acid: > 200 bp

Recovery rate: 80-90%

Binding capacity: 30 ug

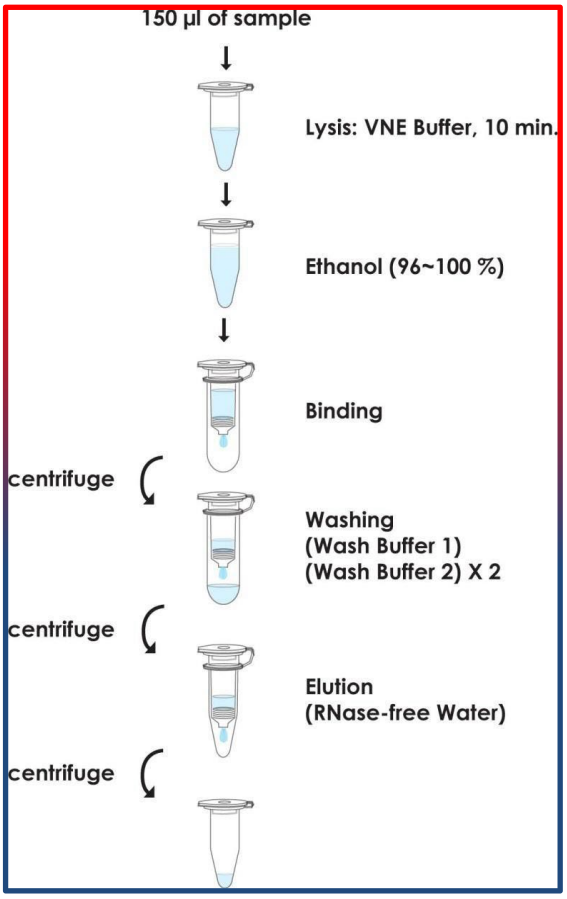
Elution Volume: 40-50 µl

Operation time: 20 minutes

Binding capacity: 60 ug RNA/column

Applications:

- Real-time PCR
- PCR
- RT-PCR
- Real-time RT-PCR



Quality Control:
The quality of our Viral RNA/ Viral Nucleic Acid Mini Kit is tested on a lot-to-lot basis according to ISO quality management system.

Cat. N.	Product Name	Size	Store at
DR-002	Viral Nucleic Acid (DNA/RNA) Extraction Kit I	100 preps	Store at room temperature for 1 year

Environmental DNA/RNA Extraction Kits

Plant Genomic DNA Extraction Mini Kit

Soil DNA Isolation Mini Kit

Stool DNA Isolation Mini Kit

Viral Nucleic acid (DNA& RNA) Extraction Kit

Fungi/Yeast Genomic DNA Extraction Mini Kit

Plant Total RNA Purification Mini Kit

PLANT GENOMIC DNA EXTRACTION MINI KIT DE-021

Plant Genomic DNA Mini Kit provides a fast and simple method to isolate total DNA (genomic DNA, mitochondrial chloroplast and viral DNA) from plant tissue and cells. Plant tissues are ground in liquid nitrogen and lysed by buffer containing detergent. The tissue debris in lysate could be removed by provided filter column. In the presence of a chaotropic salt, the genomic DNA in the lysate binds to the glass fiber matrix in the spin column. After washing off the contaminants, the purified DNA is eluted by low salt elution buffer or waters.

Features

- **High Purity:** DNA is immediately suitable for a variety of applications, including amplification, digestion, PCR etc.
- **High Speed:** Using a column type extraction system to allow a more rapid, more convenient methods compared to the conventional mmethods. Rapid speed for the isolation of genomic DNA from various plants, within 40 minutes.
- **Safe:** The kits use a spin column tube and removes proteins, nucleases in cells, it is notnecessary to treat the sample with harmful organic solvents such as phenol and chloroform.

Applications

- Real-time PCR
- PCR
- RFLP
- Amplification
- Southern blotting

Time Required

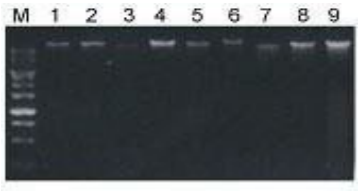
About 30-60 minutes depending upon the sample types.

Sample Size

Mini: up to 100mg fresh sample or 20mg dry sample.

Storage Conditions: Plant Genomic DNA Extraction Kit can be stored at room temperature (15-25 °C). Stable for 1 year at room temperature at 15-25°C.

The Quality of DNA After Purification



7: Populus tremula (Aspen)

8: Flammulina velutipes

9: Oxalis comiculats (Fourleaf clover)

DNA Yield

Sample	DNA yield (ug)	
	Mini	Maxi
	100 mg young leaf	1 g young leaf
Arabidopsis	3 ~ 5	30~50
Rice	10~15	100~150
Tomato	10~15	100~150
Tobacco	20~25	200~250
Chinese Yam	30~60	300~500
Maize	15~20	150~200
Sweet Potato	20~30	200~300
Orchis	5~10	50~100
Campor Tree	15~20	150~200
Spinach	5~10	50~100
Bamboo	10~15	100~150

Cat. N.	Product Name	Size	Store at
DE-021	Plant Genomic DNA Extraction Mini Kit	50 preps	Store at RT for 1 years Store RNAse A at -20°C
DE-022		100 preps	

SOIL DNA ISOLATION MINI KIT DE-025 & DE-026

Our Soil DNA Isolation Mini Kit is suitable to isolate DNA from different environmental samples improving DNA recovery in terms of DNA yield.
The technology operates through our high-quality beads- beating disruption method and is perfect for use with different ***soil samples of up to 0.5 g.*** The silica membrane technology, and spin column along with beads- beating method guarantee the high-quality purification and isolation of DNA that can be used for PCR, genotyping, arrays, etc. The inhibitors of downstream PCR or enzymatic reactions will be removed with the sequent buffers in this kit. Phenol/chloroform is not required in the whole procedure; all operation can be finished within 60 minutes. The purified DNA is ready-to-use for downstream applications.

Specifications:

Principle: Spin Column (silica membrane)

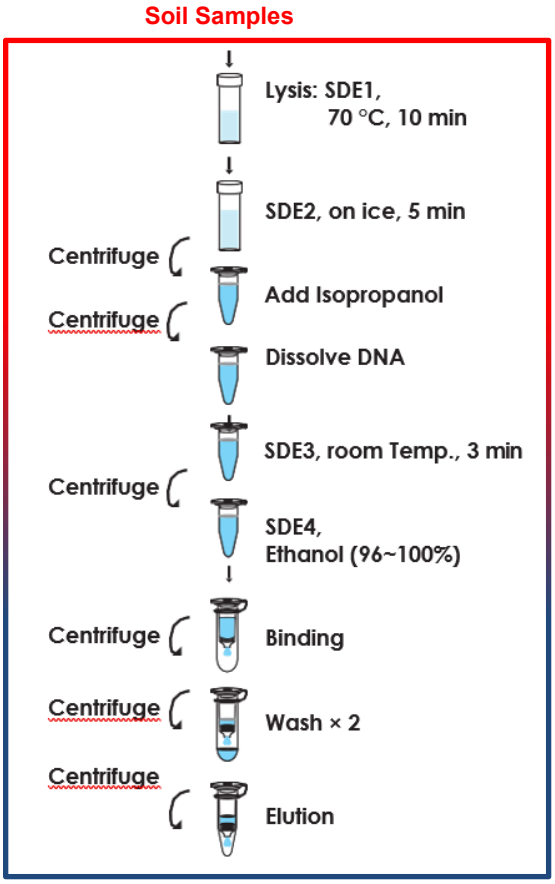
Sample: **0,25 -0,5 g**

Operation time: < 60 min

Elution volume: 50~200 µl

Important Notes:

1. Buffers provided in this system contain irritants. Wear gloves, safety glasses
and lab coat when handling these buffers.
2. Check FSDE1 Buffer before use, Warm FSDE1 Buffer at 60°C for 10 minutes if
any precipitate formd.
3. Add indicated volume of ethanol (96~100%) to Wash Buffer before use.
4. Prepare a heating block or a water bath to 70°C. If DNA is isolated from
gram positive bacteria, prepare a heating block or a water bath to 95°C
for another incubation.
5. All centrifuge steps are done at full speed (~18,000 xg) in a microcentrifuge.
6. Preheat Elution Buffer or ddH2 O to 60°C for elution step



Cat. N°	Product Name	Size
DE-025	Soil DNA Isolation kit	50 Reactions
DE-026		100 Reactions

STOOL DNA ISOLATION MINI KIT DE-023 & DE-024

Stool DNA Isolation Mini Kit is designed for the isolation of high-quality total DNA from 50~200 mg of fresh or frozen stool samples. The inhibitors, such as polysaccharides and humic acid, will be removed with the sequent buffers in this kit. High quality DNA for sensitive downstream applications including PCR, qPCR, Sequencing and microarray

Specifications:

Principle: Spin Column (silica membrane)

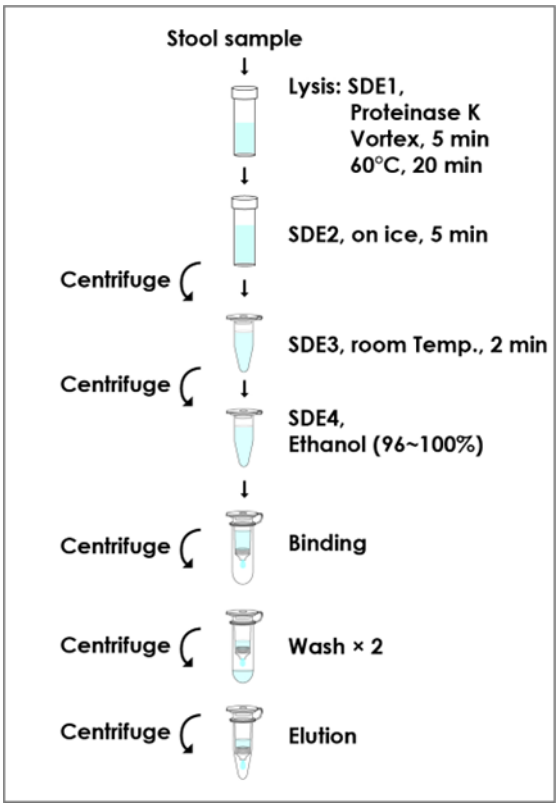
Sample: 50~200 mg

Operation time: < 60 min

Elution volume: 50~200 µl

Important Notes

1. Buffers provided in this system contain irritants. Wear gloves and lab coat when handling these buffers.
2. Check FSDE1 Buffer before use, warm FSDE1 Buffer at 60°C for 10 mins if any precipitate formed.
4. Add indicated volume of ethanol (96~100%) to Wash Buffer before use.
5. Prepare a heating block or a water bath to 60°C. If DNA is isolated from gram positive bacteria, prepare a heating block or a water bath to 95°C
for another incubation.
6. All centrifuge steps are done at full speed (~18,000 x g) in a microcentrifuge.
7. Preheat Elution Buffer or ddH2 O to 60°C for elution step.



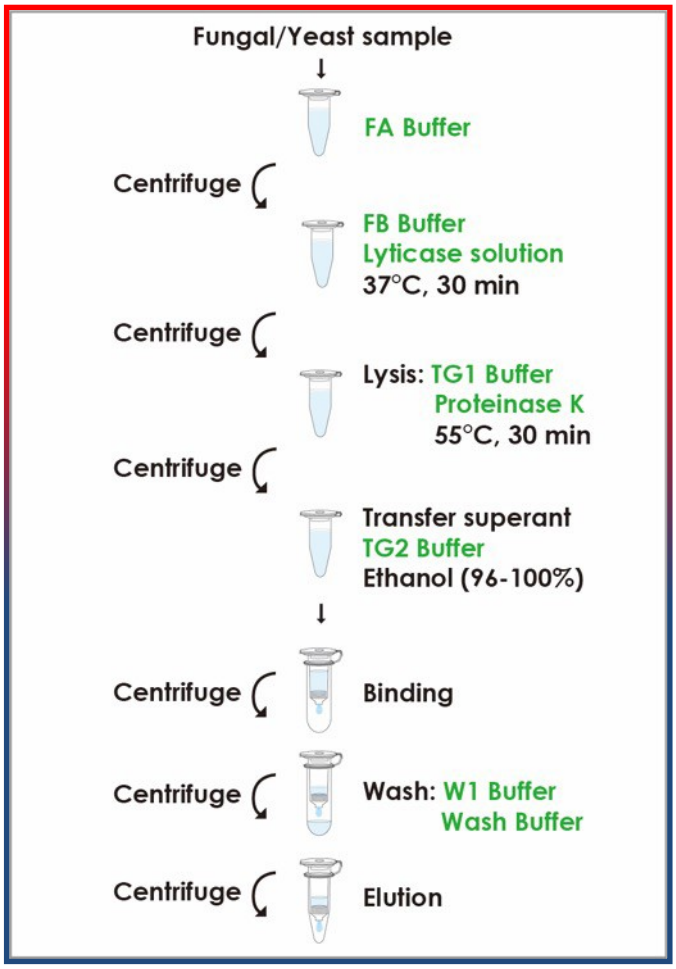
Cat. N°	Product Name	Size
DE-023	Stool DNA Isolation kit	50 Reactions
DE-024		100 Reactions

FUNGI /YEAST GENOMIC DNA EXTRACTION MINI KIT DE-046

Description

The FavorPrepTM Fungi/Yeast Genomic DNA Extraction Mini Kit is designed for the purification of DNA from fungus and yeast cells. The enzyme teatment (lyticase & proteinase K) and **bead- beating homogenization** are applied to lyse samples efficiently and improving DNA yield. This kit provides the most complete and effective method to extract application-ready pure genomic DNA from fungi and yeast samples.

Technology: mini spin column (silica matrix)
Sample size:1~ 5 x10^6 cell culture fungal/yeast cells
Operation time:~ 60 minutes
Binding capacity:60 μg/ column
Column applicability: centrifugation and vaccum



Ordering Information

Cat. No.	Product Name	Size	Store at:
DE-046	Fungi Yeast genomic DNA Extraction Kit	50 preps.	At Room Temperature for 1 year Lyticase : At -20℃.

PLANT TOTAL RNA PURIFICATION MINI KIT RE-007

Description

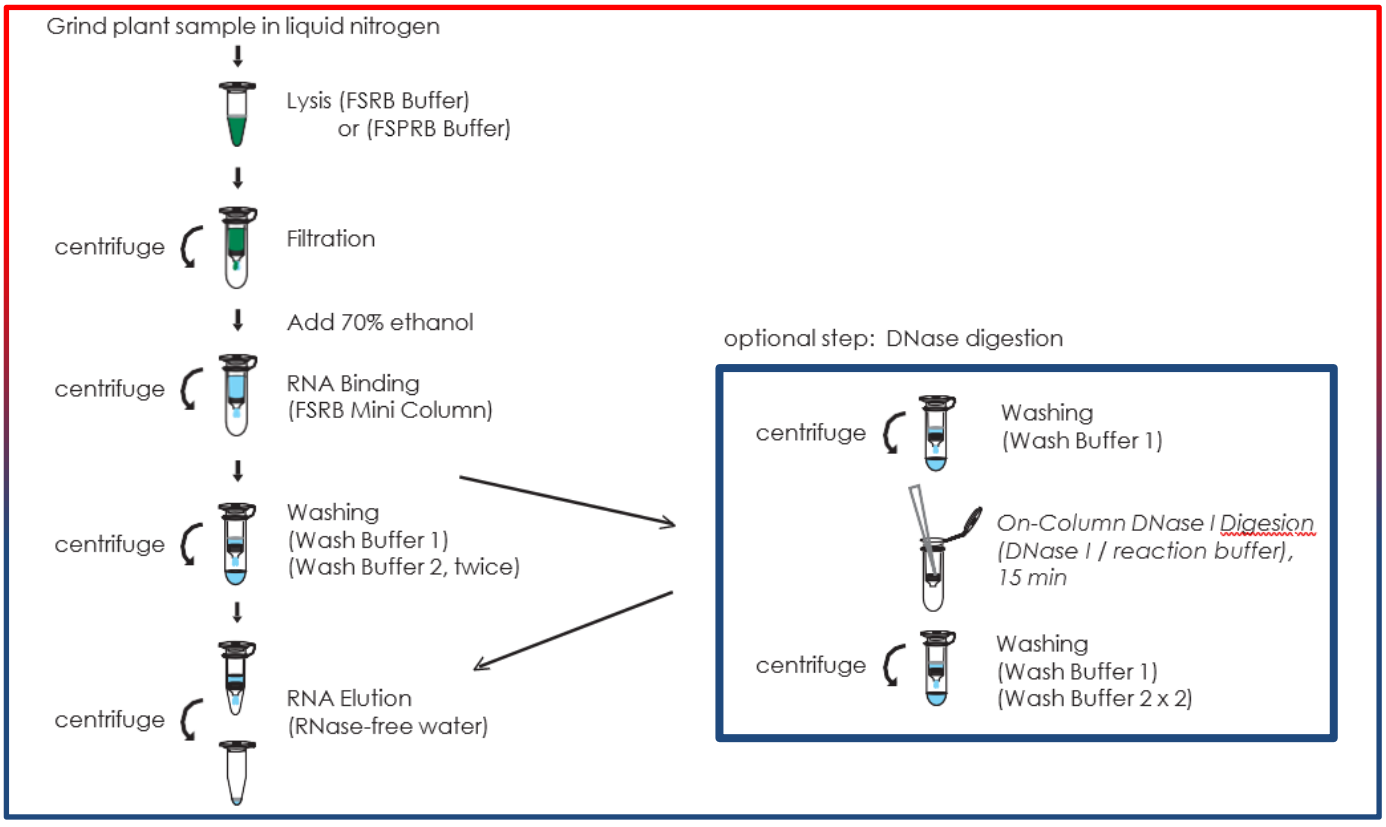
The Plant Total RNA Purification Mini Kit is designed for purification of total RNA from plant tissues and cells using the modified salt precipitation procedure and RNase inhibitors without the use of hazardous solvents such as phenol. Plant RNA is quickly and efficiently isolated and is immediately available for downstream applications, including RT-PCR, Northern blotting, primer extension and cDNA library construction. For RNA Plant Total RNA extraction from woody plant we reccomend RE-015 (50 preps) and RE-016 (100 preps).

Specification:

Principle: spin column (silica membrane)
Sample: up to 100 mg plant tissues or $1x10^7$ plant cells
Operation time: 30- 60 min
Binding capacity: up to 100 ug Total RNA/column
Expected Yield: 5-30 ug of Total RNA from 100 mg of young leave
Column Applicability: Centrifugation and Vacuum
Minimum Volume: 30 ul

Applications

- Nothern blotting hybridizations
- Primer extension
- RT-PCR
- RNase protection assays
- Differential display
- As starting material for purification of mRNA for cDNA synthesis



Ordering Information

Cat. No.	Product Name	Size	Store at
RE-007 RE-008	Plant Total RNA Purification Mini Kit	50 preps 100 preps	Store at room temperature at 4-8°C for 1 year.

QUBIT Reagents

dsDNA Quantification

Qubit dsDNA Broad Range Quantification Assay Kit

Qubit dsDNA High Sensitivity Quantification Assay Kit

Qubit dsDNA Broad Range Quantification Assay Kit FS-QBR-01

The Qubit dsDNA Broad Range Quantification Assay Kit offers sensitive and selective detection of purified dsDNA samples. The kit provides a linear detection range from 2-100ng and is designed for use with handheld fluorometers such as Qubit® fluorometers from ThermoFisher Scientific. The kit is suitable for Qubit® Fluorometer: Qubit 1.0, Qubit 2.0, Qubit 3, and Qubit 4 fluorometers.

Contents and Storage Information

Components	100assays	500assays	Concentration
Qubit PicoGreen dsDNA BR Reagent(Component A)	250μL	1.25mL	200X in DMSO
Qubit dsDNA BR Buffer (Component B)	50mL	2500mL	TE Buffer
Qubit dsDNA BR Standard#1 (Using Component A)	N/A	N/A	0 ng/uL in TE Buffer
Qubit dsDNA BR Standard#2 (Component C)	1mL	5mL	100 ng/uL in TE Buffer

Qubit® Fluorometer: Qubit 1.0, Qubit 2.0, Qubit 3, and Qubit 4 fluorometers.
Quantitation Range: 0.2-100 ng

The PicoGreen reagent is a proprietary, asymmetrical cyanine dye. Free dye does not fluoresce, but upon binding to dsDNA it exhibits a >1000-fold fluorescence enhancement. PicoGreen is 10000x more sensitive than UV absorbance methods, and highly selective for dsDNA over ssDNA and RNA.

Qubit dsDNA High Sensitivity Quantification Assay Kit FS-QHS-02

The Qubit dsDNA High Sensitivity Quantification Assay Kit is designed for use with handheld fluorometers such as the Qubit® fluorometers from ThermoFisher Scientific. The kit is suitable for Qubit 1.0, Qubit 2.0, Qubit 3, and Qubit 4 fluorometers.

Contents and Storage Information：

Components	100assays	500 assays	Concentration
Qubit PicoGreen dsDNA HS Reagent(Component A)	250μL	1.25mL	200X in DMSO
Qubit dsDNA HS Buffer (Component B)	50mL	2500mL	TE Buffer
Qubit dsDNA HS Standard#1 (Using Component B)	N/A	N/A	0 ng/uL in TE Buffer
Qubit dsDNA HS Standard#2 (Component C)	1mL	5mL	10 ng/uL in TE Buffer

Qubit® Fluorometer: Qubit 1.0, Qubit 2.0, Qubit 3, and Qubit 4 fluorometers.
Quantitation Range: 0.2-100 ng

The PicoGreen reagent is a proprietary, asymmetrical cyanine dye. Free dye does not fluoresce, but upon binding to dsDNA it exhibits a >1000-fold fluorescence enhancement. PicoGreen is 10000x more sensitive than UV absorbance methods, and highly selective for dsDNA over ssDNA and RNA.